

Introduction to variant calling

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Norway

BIOS-IN 5K/9K
24th of Oct 2022

@archaeogenomics 

CEES
Centre for Ecological and Evolutionary Synthesis



UiO : University of Oslo



Evolutionary Biologist

specialize in ancient DNA

Archaeogenomics group
(10+ MSc, PhDs & Postdocs)

@archaeogenomics



Multidisciplinary research:

Archaeology

Biology

Ecology

Molecular methods/sequencing

Genomics

Bioinformatics

Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?

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- 2) Variant calling pipelines/methods and limitations

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- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and limitations
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

Introduction

Genetic variation (genomic differences between individuals) is everywhere



Genetic variation at different scales:

1) Biological differences (phenotypes) between species



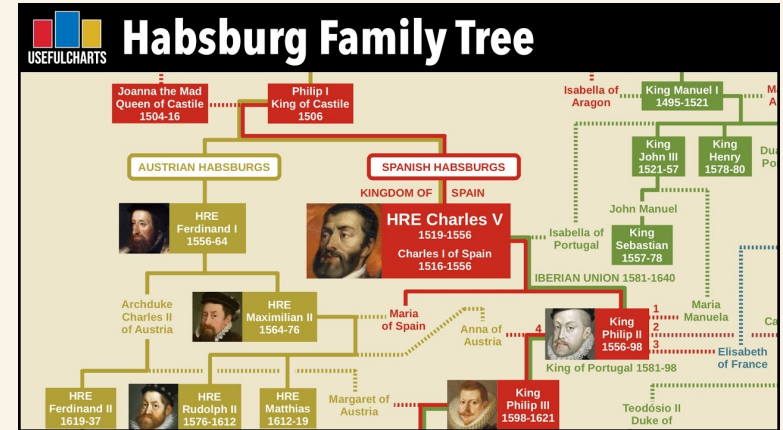
Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
- 2) Biological differences within species



Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
- 2) Biological differences within species
- 3) Patterns of relatedness between individuals/
populations (23 and me)



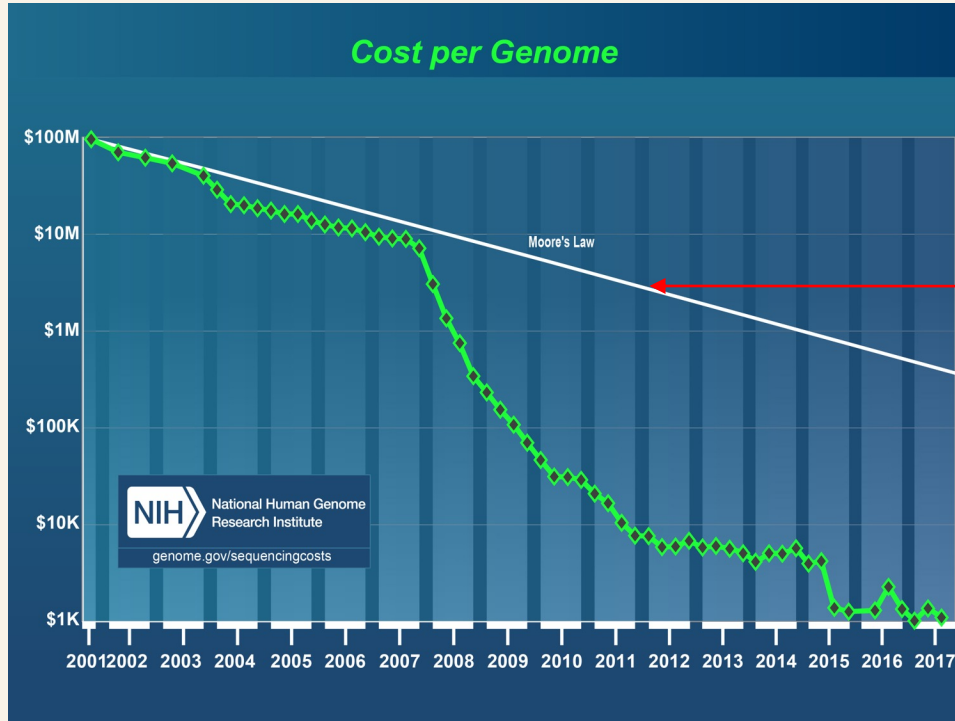
Genetic variation explains many observations within
biology

Genetic variation explains many observations within
biology

Knowing patterns of/quantifying genetic variation has
enormous potential for a wide range of applications in
society

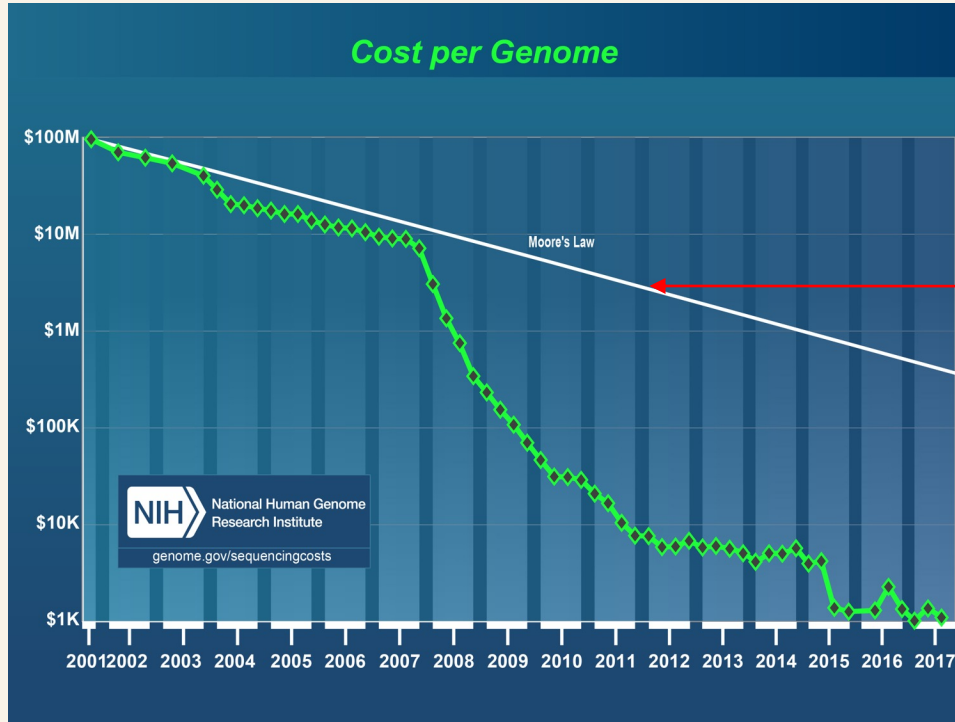
*(e.g. personal medicine, forensic sciences, biodiversity
assessments, crop improvement, animal breeding, conservation
management, history & genealogy, etc etc)*

Why are we here?



Moore's Law

Why are we here?

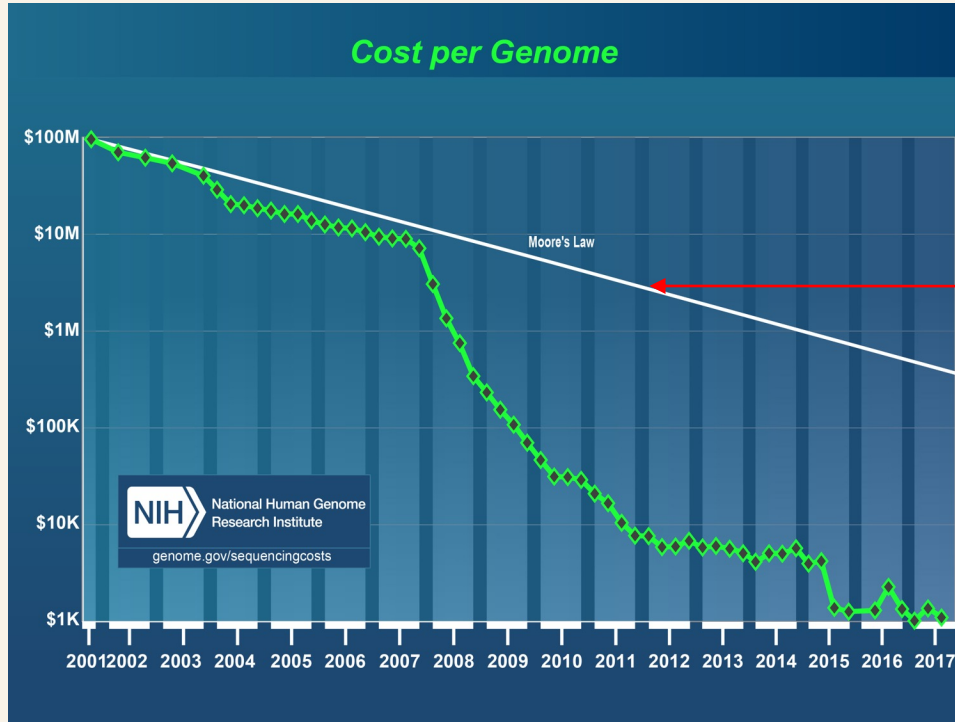


2000



2017

Why are we here? *Phenomenal* technological advances



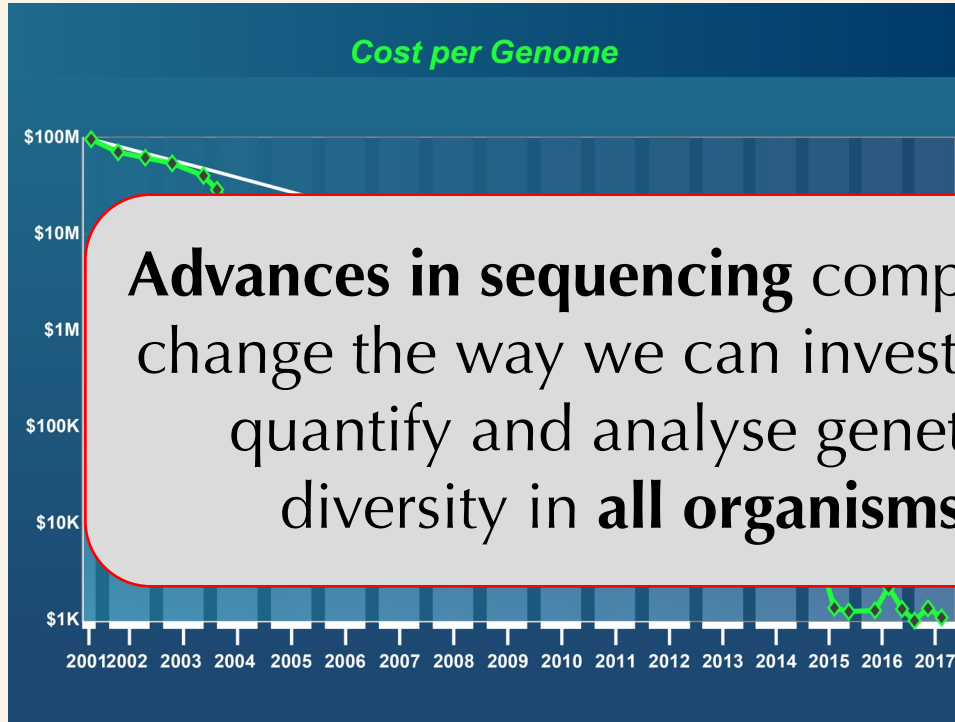
2000



2017

Technological revolution that has *fundamentally* changed the way we do biology

Why are we here? ***Phenomenal*** technological advances



Advances in sequencing completely change the way we can investigate, quantify and analyse genetic diversity in **all organisms**



2004



2017

Technological revolution that has *fundamentally* changed the way we do biology

How has sequencing changed and is changing the world?

Changed healthcare

Sequencing (genome and exome) funded solely by *healthcare* systems

2012

~1%

2017

~20%

2022

>80%

Dag Undlien (OUH)

Today



**2030
Precision medicine**



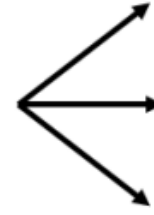
Today



Average



Standard



Improved Outcome



No benefit



Adverse side effects

2030
Precision medicine



Whole genome sequencing is actively used in Norwegian healthcare *and* provides clinical solutions



Population health data



Data analytics



Personalised treatment



Improved Outcome

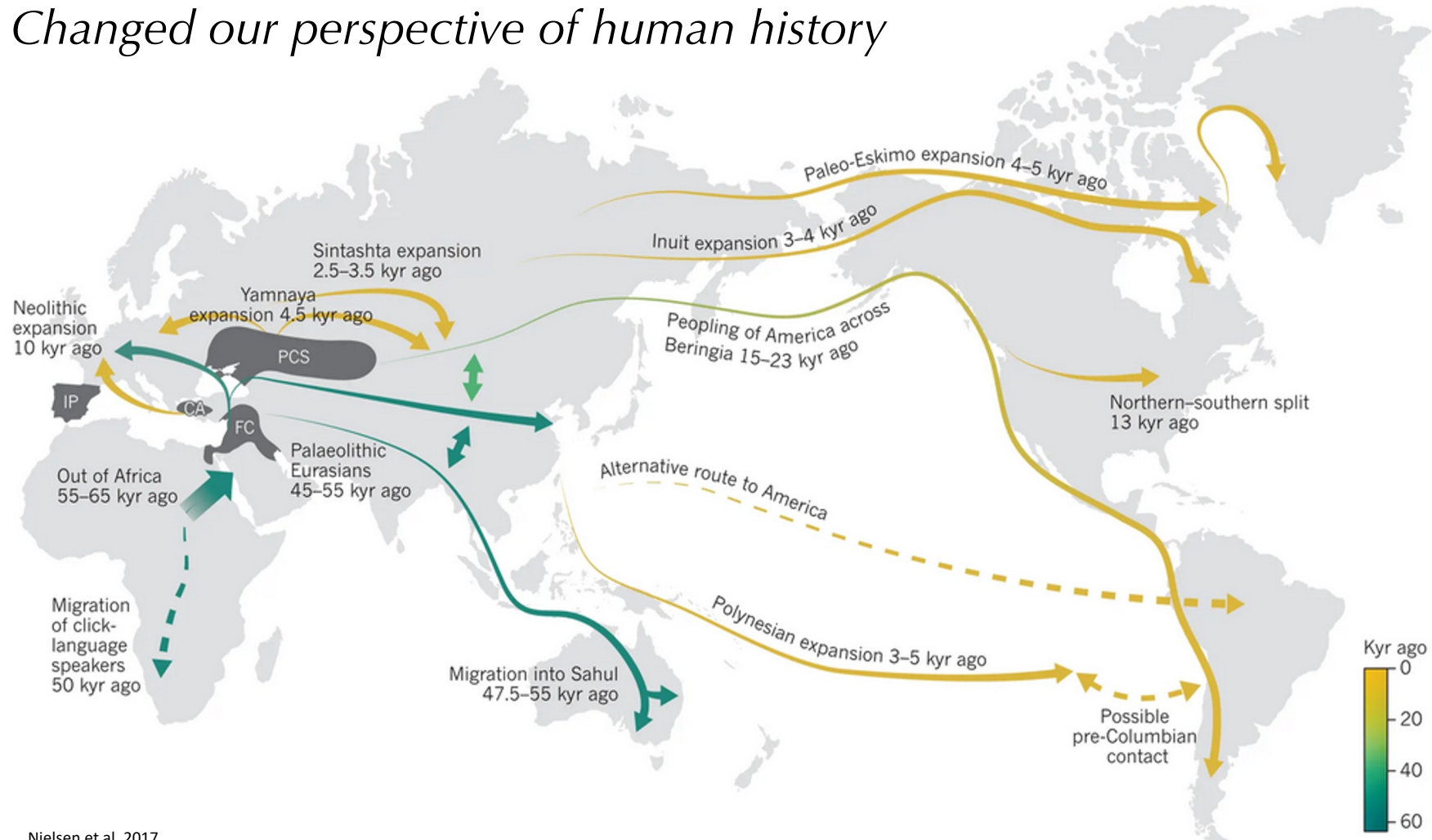


Improved Outcome



Reduced side effects

Changed our perspective of human history



Changed forensic capabilities

Using continuously expanding public genomic databases (e.g. 23 and me)...

The New York Times

Genealogists Turn to Cousins' DNA and Family Trees to Crack Five More Cold Cases

Police arrested a D.J. in Pennsylvania and a nurse in Washington State this week, the latest examples of the use of an open-source ancestry site since the break in the Golden State killer case.

Changed forensic capabilities

Or by the genetic testing of thousands of people!

As the *Times* reports, that law paved the way for a prosecutor in the Verstappen case to call for the voluntary DNA sampling of 21,500 Dutchmen, and the obligatory sampling of 1,500 men who were of “special interest” to investigators.

The alleged killer, 55-year-old Jos Brech, was one of those 1,500 men who were mandated to provide a DNA sample. He never showed up. Dutch officials grew suspicious and took DNA samples from Brech's relatives. The results matched the DNA

Changed vaccine development and disease tracking

Genomic epidemiology of SARS-CoV-2 with subsampling focused globally over the past 6 months

Built with [nextstrain/ncov](#). Maintained by the [Nextstrain team](#). Enabled by data from [GISAID](#).

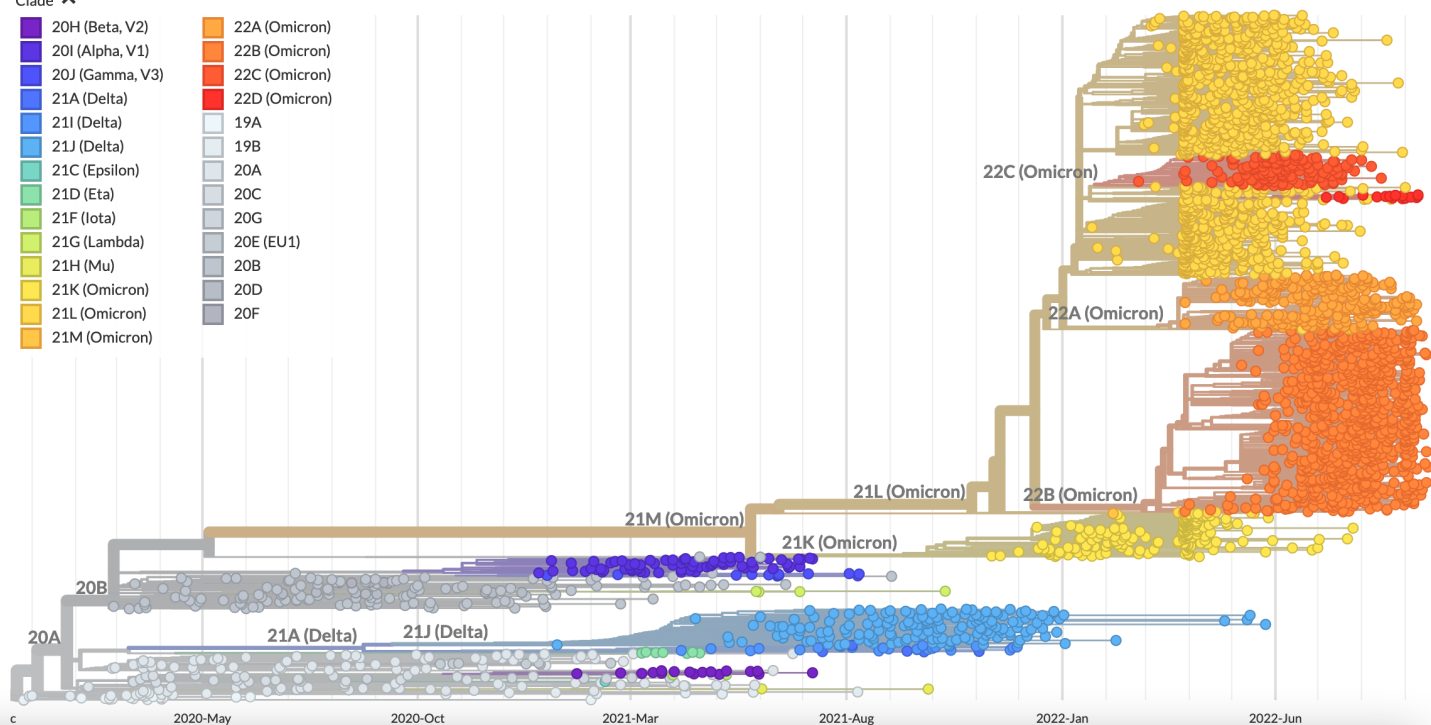
Showing 3074 of 3074 genomes sampled between Dec 2019 and Sep 2022.

Phylogeny

Clade ^

- 20H (Beta, V2)
- 20I (Alpha, V1)
- 20J (Gamma, V3)
- 21A (Delta)
- 21I (Delta)
- 21J (Delta)
- 21C (Epsilon)
- 21D (Eta)
- 21F (Iota)
- 21G (Lambda)
- 21H (Mu)
- 21K (Omicron)
- 21L (Omicron)
- 21M (Omicron)

- 22A (Omicron)
- 22B (Omicron)
- 22C (Omicron)
- 22D (Omicron)
- 19A
- 19B
- 20A
- 20C
- 20G
- 20E (EU1)
- 20B
- 20D
- 20F



Changed improvement and selection of commercial crops

Vitamin D Deficiency



5 IN 10 PEOPLE

globally have a vitamin D insufficiency.

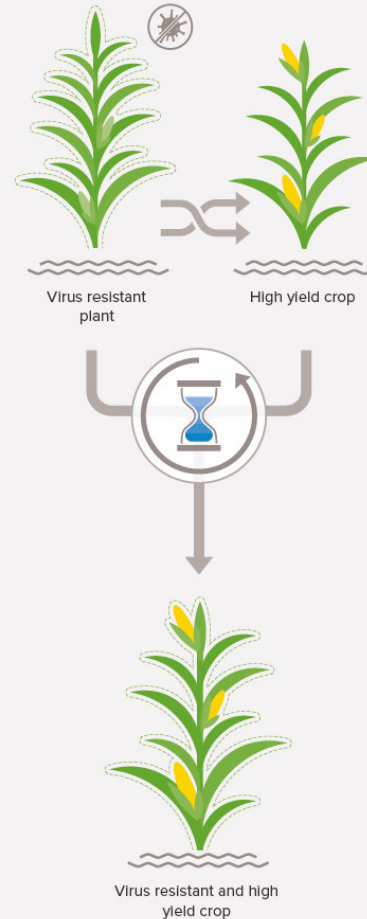
1 billion people worldwide are affected by low levels of vitamin D.

Seed WORLDGROUP

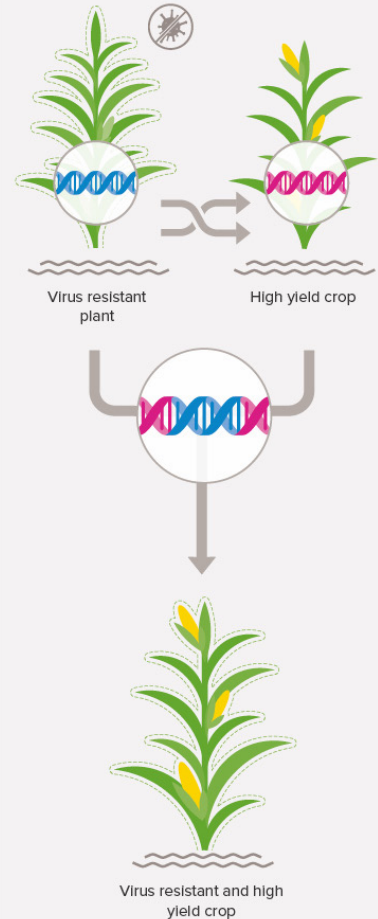


FIGURE 3 Differences between conventional breeding and GM

Conventional breeding



Genetic modification



Changed our understanding of the human microbiome

A map of diversity in the human microbiome

Lactobacillus species (*L. gasseri*, *L. jensenii*, *L. crispatus*, *L. iners*) are predominant but mutually exclusive in the vagina

Staphylococcus epidermidis colonizes external body sites

Streptococcus dominates the oral cavity with *S. mitis* > 75% in the cheek

Propionibacterium acnes lives on the skin and nose of most people

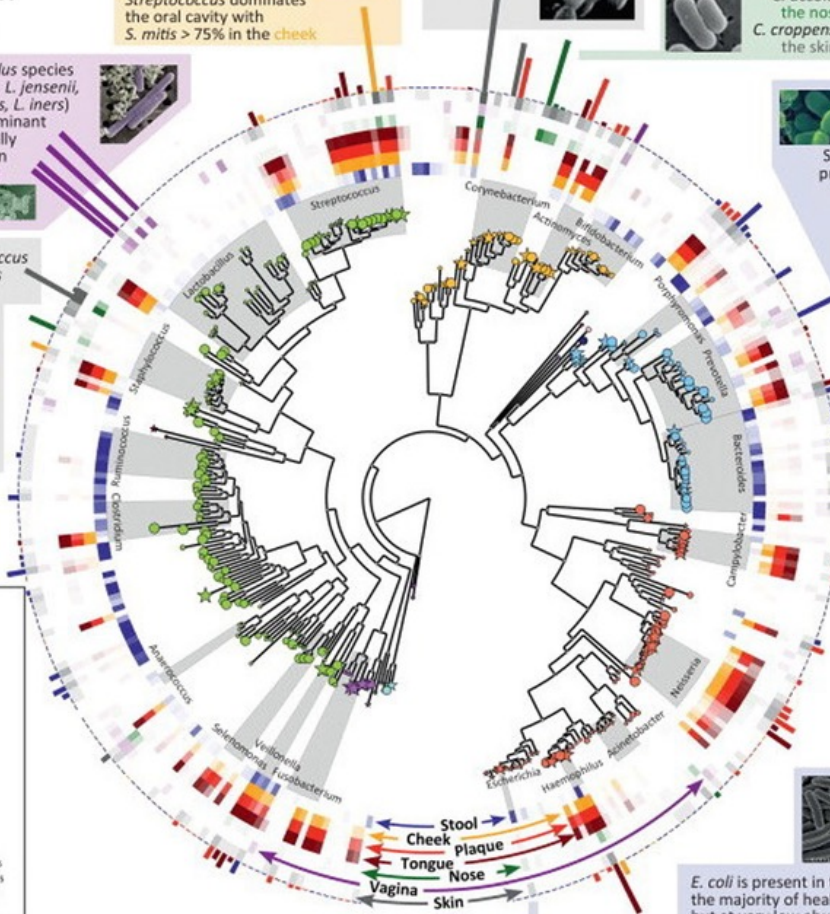
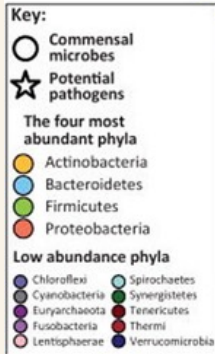
Many *Corynebacterium* species characterize different body sites:
C. matruchoti the plaque
C. accolens the nose
C. croppenstedtii the skin

Several *Prevotella* species are present in the gastrointestinal tract. *P. copri* is present in 19% of the subjects and dominates the intestinal flora when present

Bacteroides is the most abundant genus in the gut of almost all healthy subjects

Campylobacter includes opportunistic pathogens, but members live in the oral cavities of most healthy people in the cohort

E. coli is present in the gut of the majority of healthy subjects but at very low abundance



NIH Human Microbiome Project



Characterization of the microbiomes of healthy human subjects at five major body sites, using 16S and metagenomic shotgun sequencing.

Enter HMP1



Characterization of microbiome and human host from three cohorts of microbiome-associated conditions, using multiple 'omics technologies.

Enter IHMP

Changing our perspective of extinct species

How Woolly mammoths could be brought back from extinction

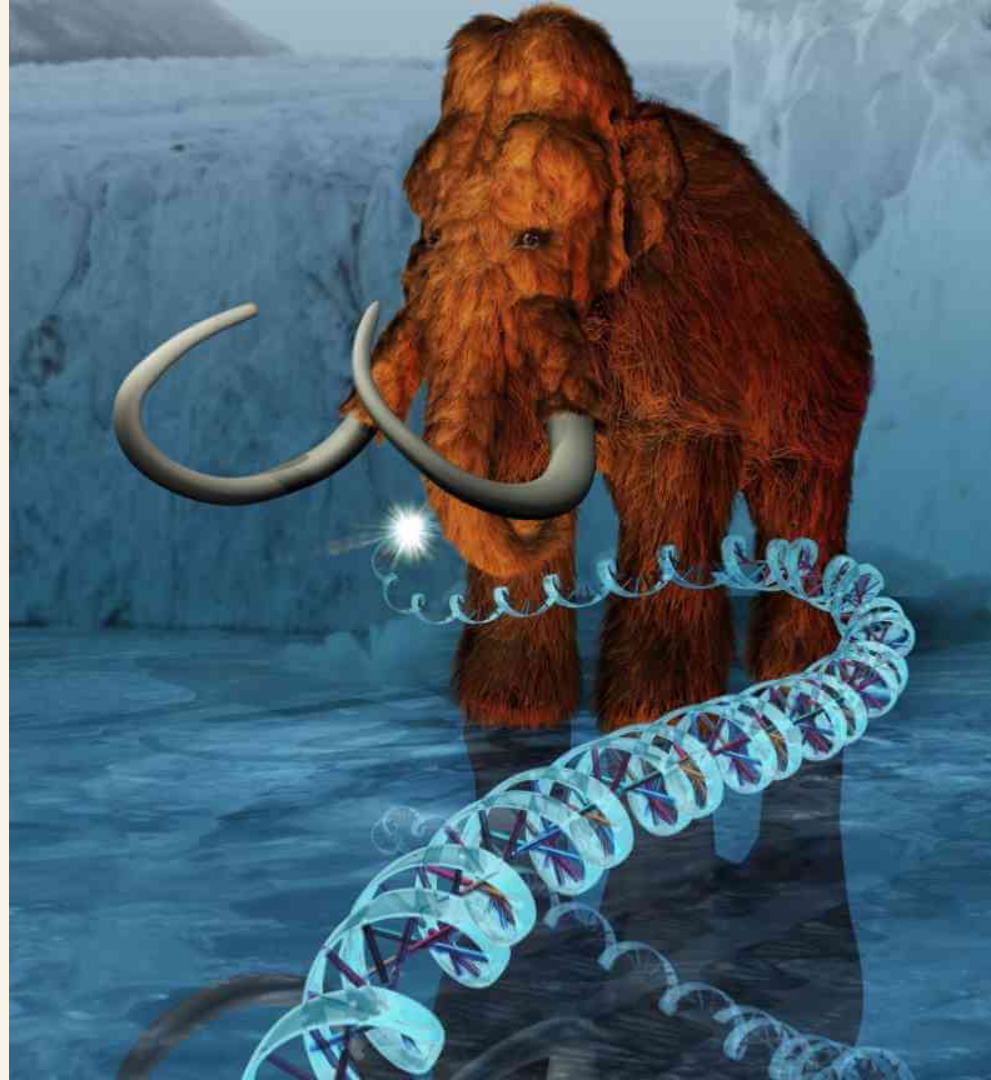
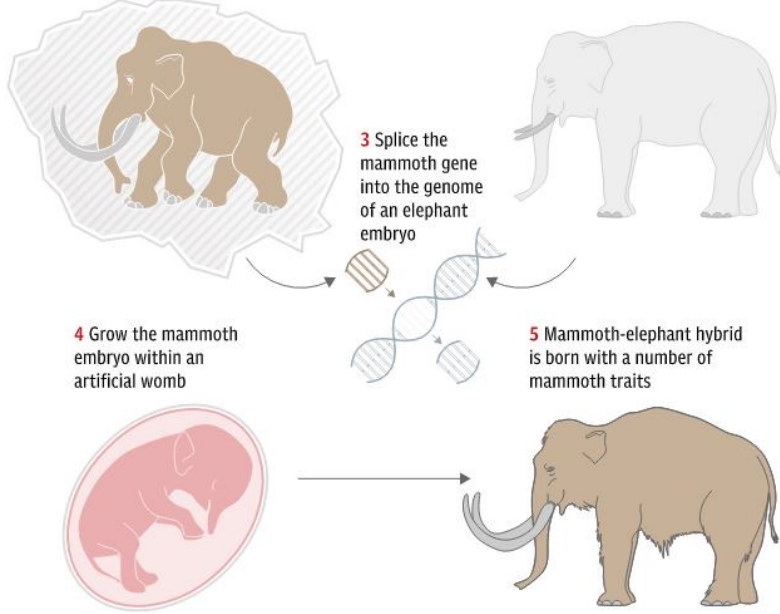
1 DNA extracted from mammoth found in permafrost.

2 Identify genes which separate them from elephants, such as those which code for a shaggy coat, big ears and antifreeze blood.

3 Splice the mammoth gene into the genome of an elephant embryo

4 Grow the mammoth embryo within an artificial womb

5 Mammoth-elephant hybrid is born with a number of mammoth traits



Changing our perspective of extinct species

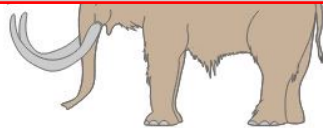
How Woolly mammoths could be brought back from extinction

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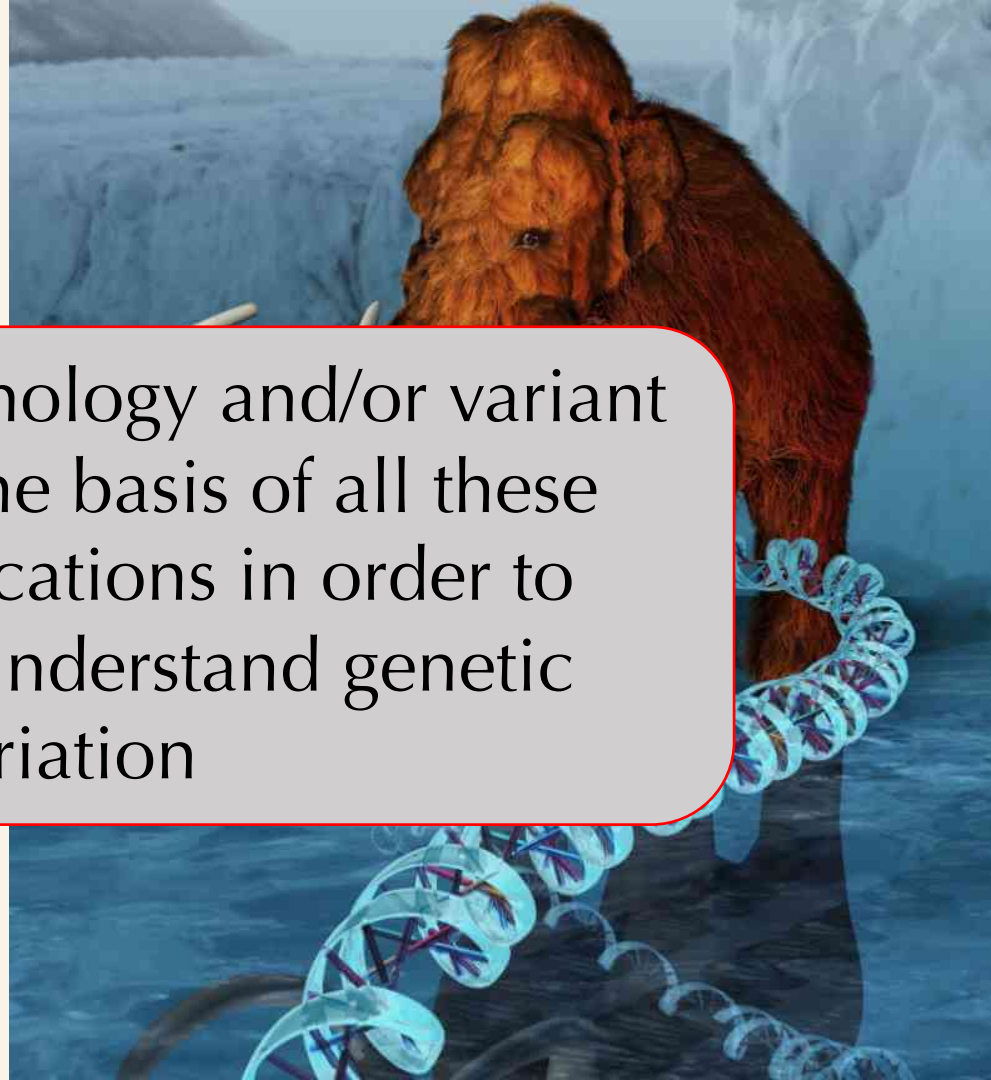
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4 Grow the mammoth embryo within an artificial womb



Sequencing technology and/or variant calling are at the basis of all these different applications in order to quantify and understand genetic variation



Available to
high school
students!
21.10 2022

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High school student is first to sequence the angelfish genome

17-year-old Indeever Madireddy sequenced the genome of his pet angelfish after it died – the first time this species has been sequenced

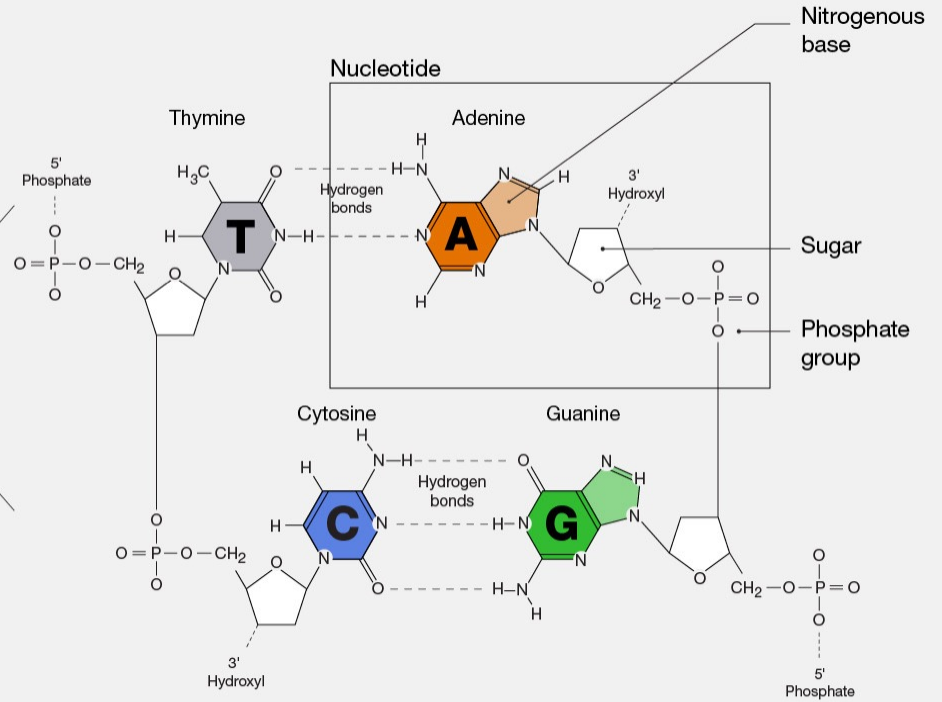
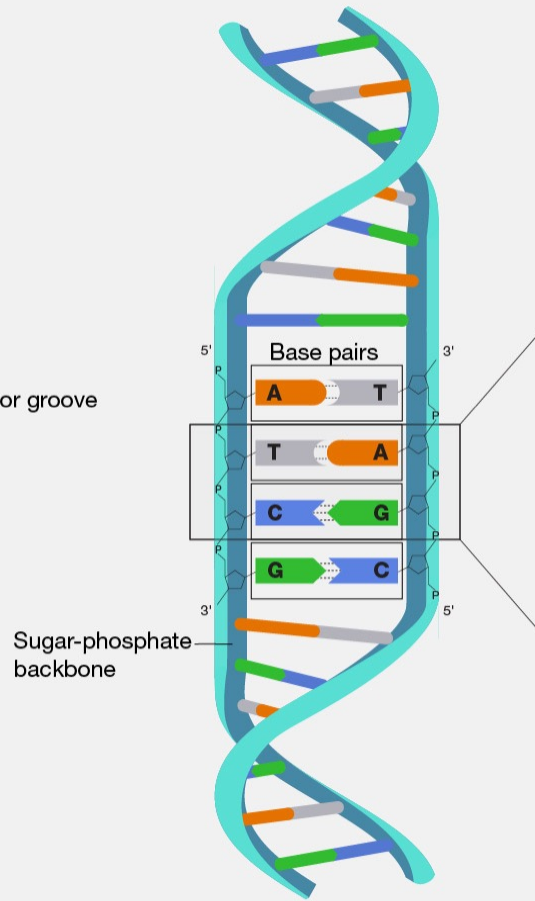


LIFE 21 October 2022

By [Michael Le Page](#)



DNA

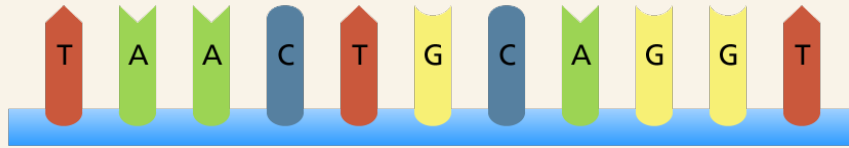


What does genetic variation look like?

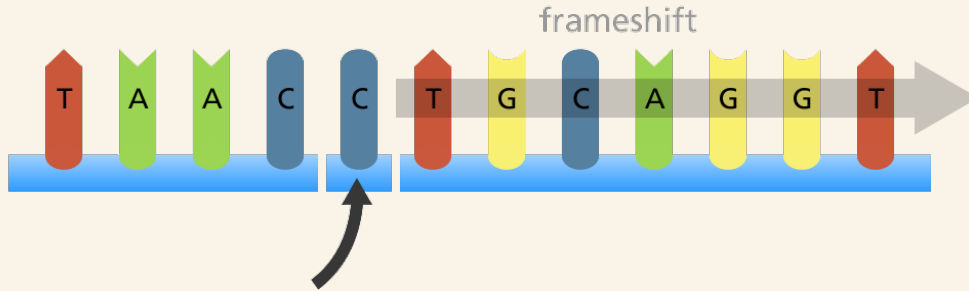
- 1) DNA (nucleotides) can be inserted or deleted (*indels*).

1) Insertion/Deletion (Indel)

Original sequence



Insertion



Can range from 1
base-pair (bp) to
many bp

What does genetic variation look like?

1) DNA (nucleotides) can be inserted or deleted (*indels*).

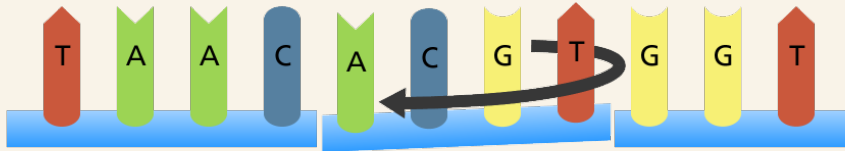
2) DNA can be *structurally rearranged*
(inversions/translocations)

2) Structural rearrangements (inversions/translocations)

Original sequence



Inversion



Can be MILLIONS of
bp long affecting the
order of many genes
simultaneously
(*Supergenes*)

What does genetic variation look like?

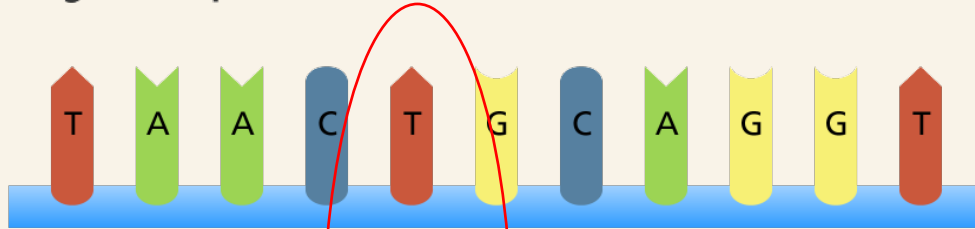
1) DNA (nucleotides) can be inserted or deleted (*indels*).

2) DNA can be *structurally rearranged*
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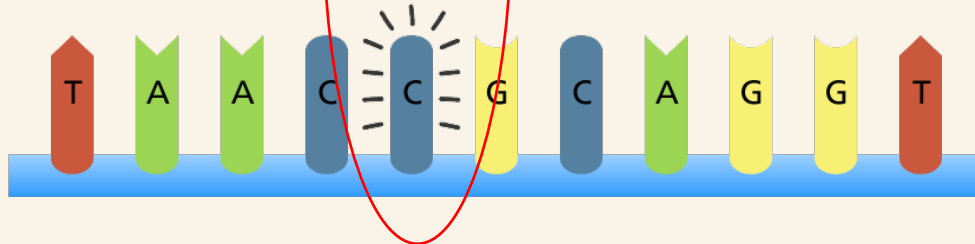
3) DNA can be *altered* at a single base pair (Single Nucleotide Polymorphism or SNP)

3) Single Nucleotide Polymorphism (SNP)

Original sequence



Point mutation



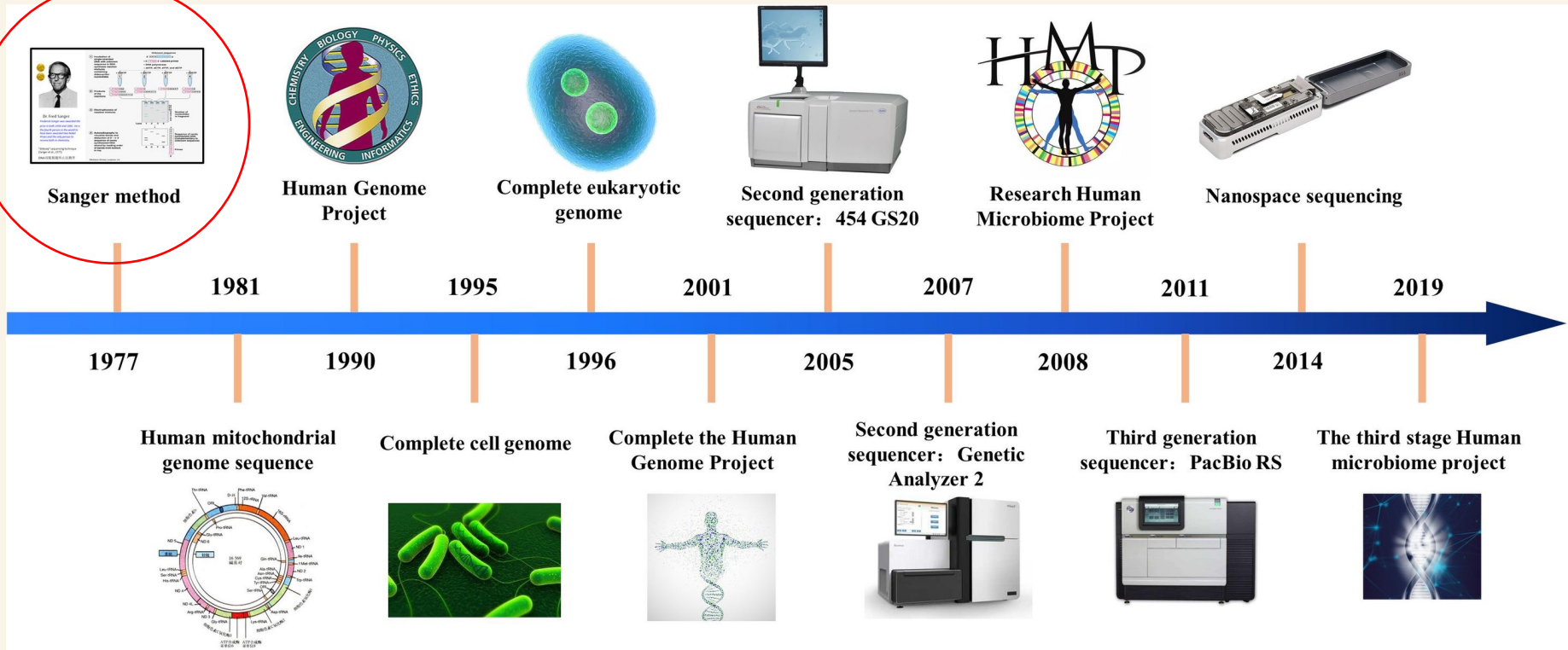
Extremely common

> 150 million known SNPs
in humans (2015)

~100 SNPs unique in
EVERY human

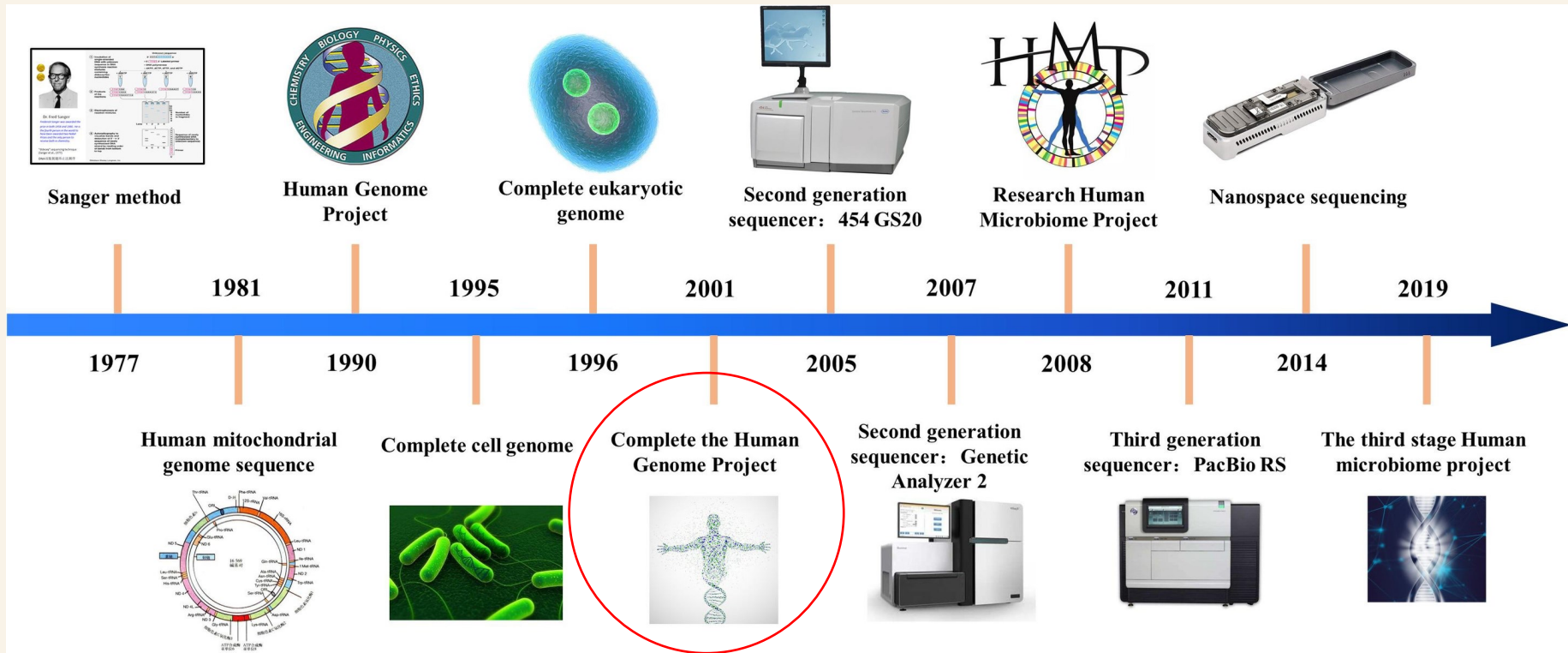
*Putatively EVERY base in
the human genome*

How do we observe and quantify genetic variation?



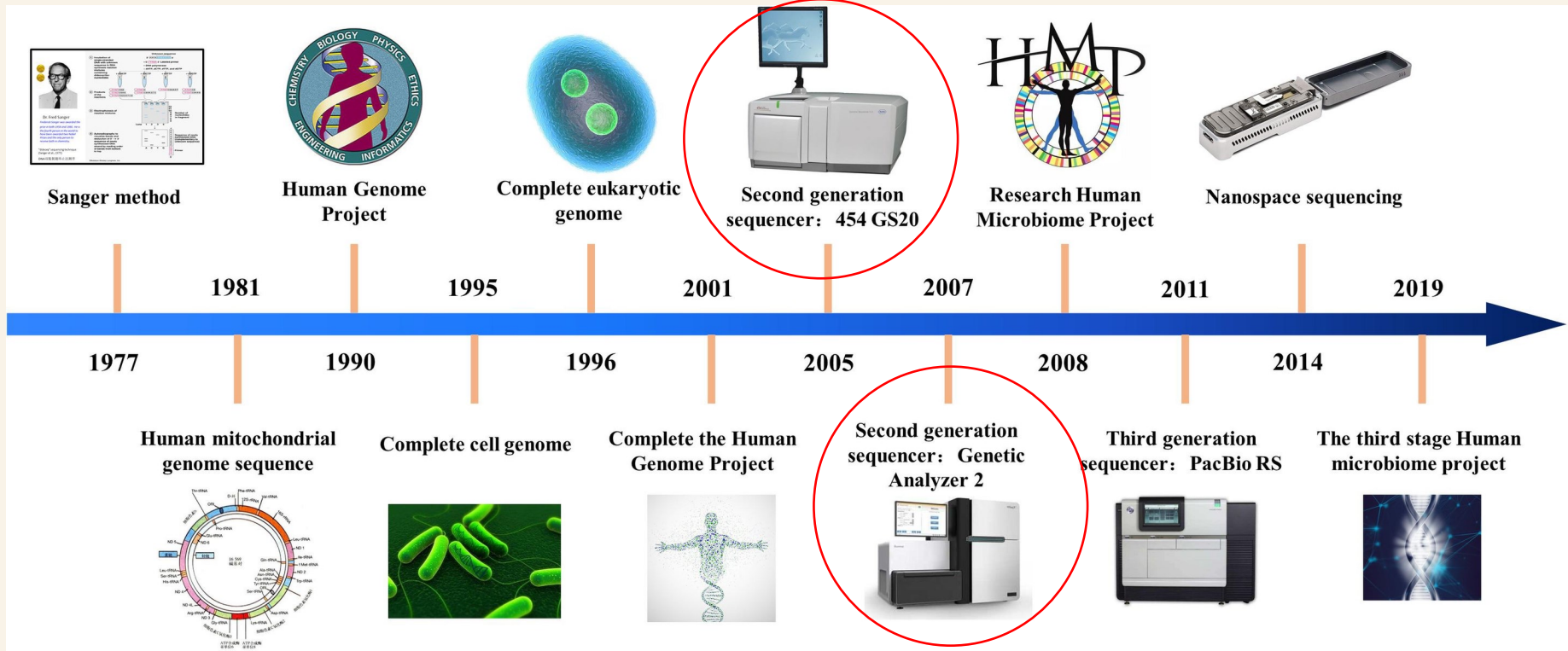
Sanger sequencing – leading sequencing technology for decades

How do we observe and quantify genetic variation?



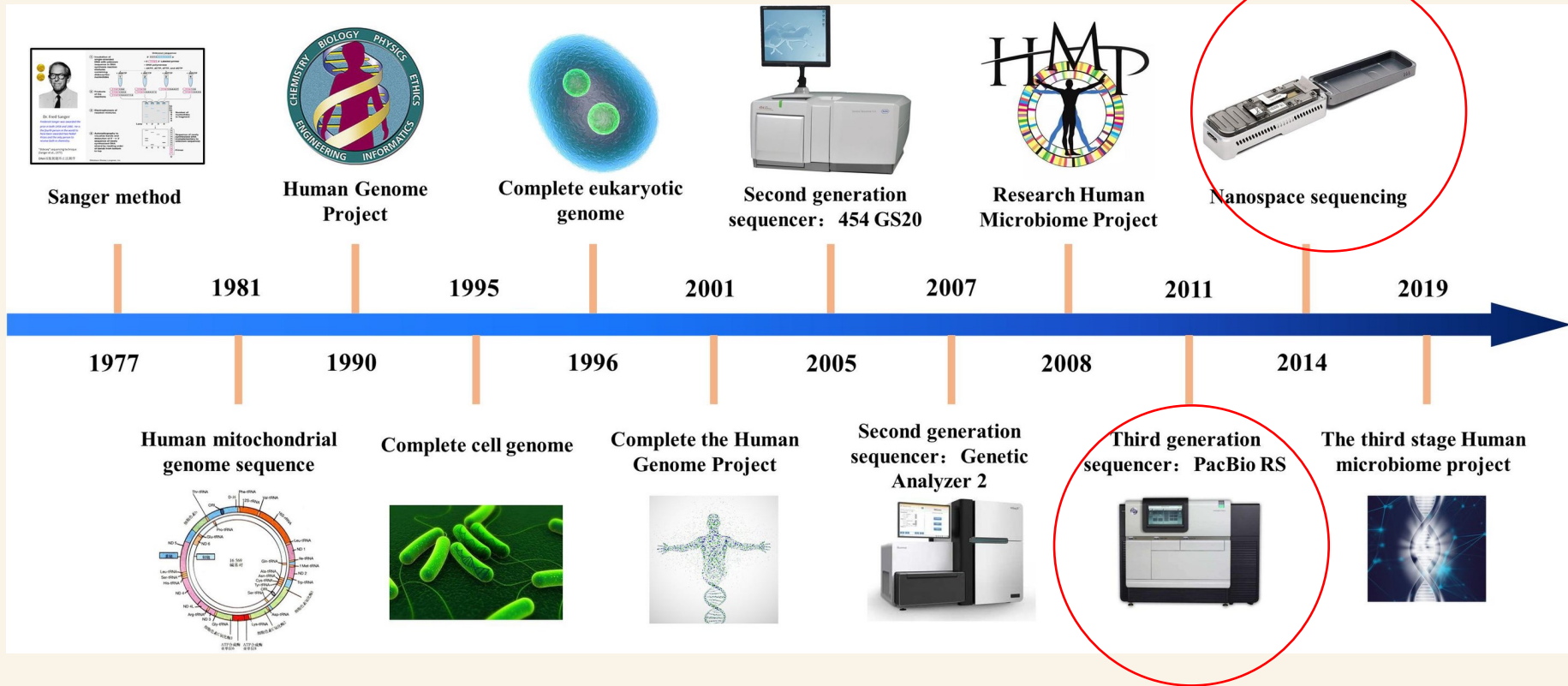
Human genome project: sparked a novel industry

How do we observe and quantify genetic variation?



“New” sequencing technologies (already outdated!)

How do we observe and quantify genetic variation?



Latest sequencing technologies that focus on long read sequencing

Two dominant technologies today



PacBio

Long read length (10k bp +)

More expensive

Specific applications

Two dominant technologies today



PacBio
Long read length (10k bp +)
More expensive
Specific applications



Illumina
Short read length (150-250 bp)
Cheap!
Workhorse of sequencing

Practical considerations: size matters!



PacBio
Long read length (10k bp +)
More expensive
Specific applications



Illumina
Short read length (150-250 bp)
Cheap!
Workhorse of sequencing

What variation can you assess with these different types of reads?

Type of variant	Short reads	Long reads
Indel	Only if small (~few bp)	Yes
Structural (inversion)	Difficult	Yes
SNP	Yes	Yes

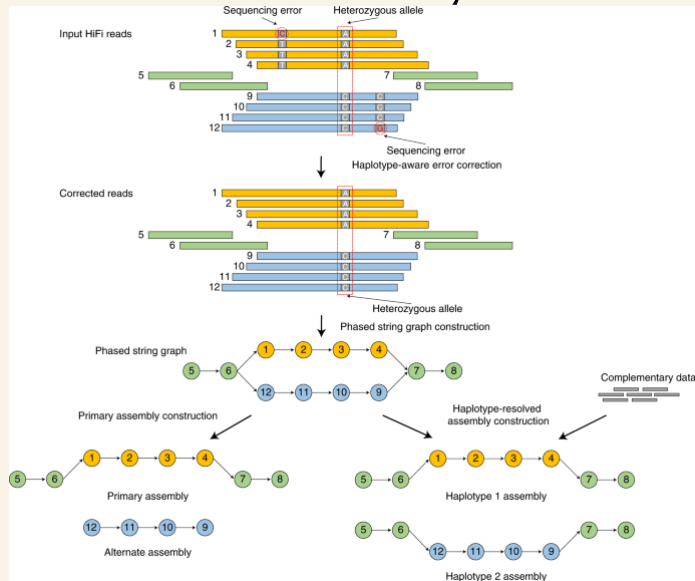
What variation can you assess with these different types of reads?

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Structural (inversion)	Difficult	Yes
SNP	Yes	Yes

Illumina ***re-sequencing*** domination means that SNPs are most reliably targeted and are most studied type of genetic variation

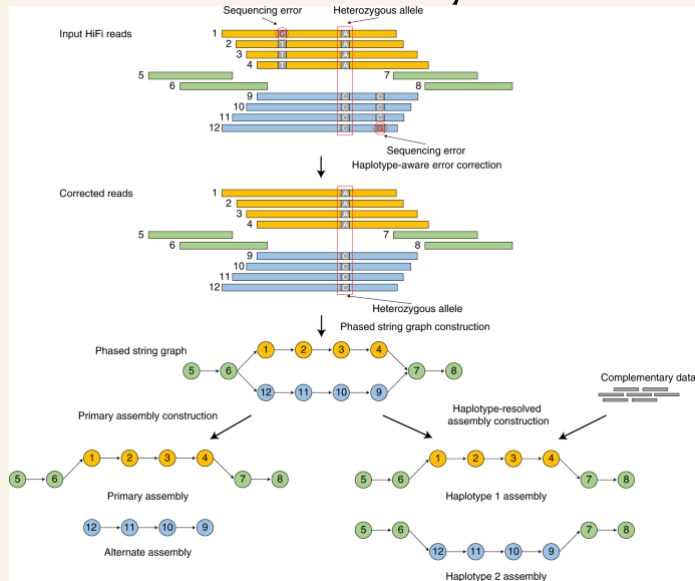
Yet there *are* different ways of assessing genetic variation

i.e. *de novo*
haplotype aware
assembly

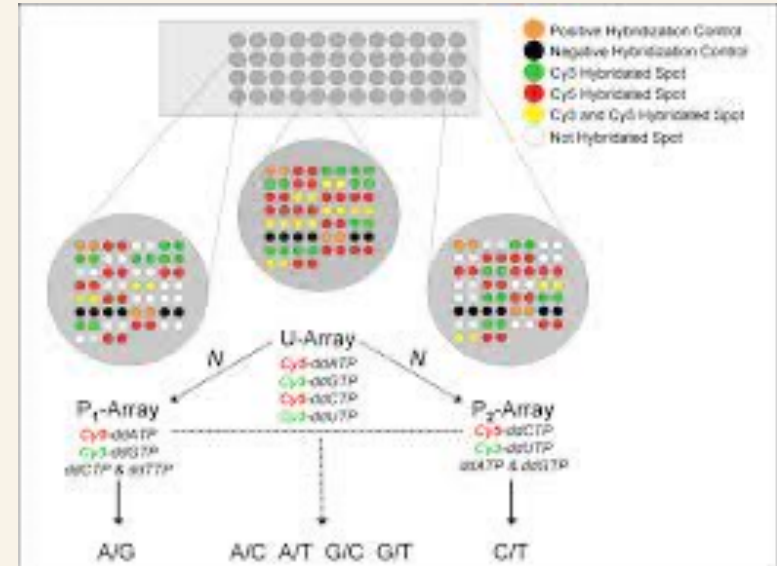


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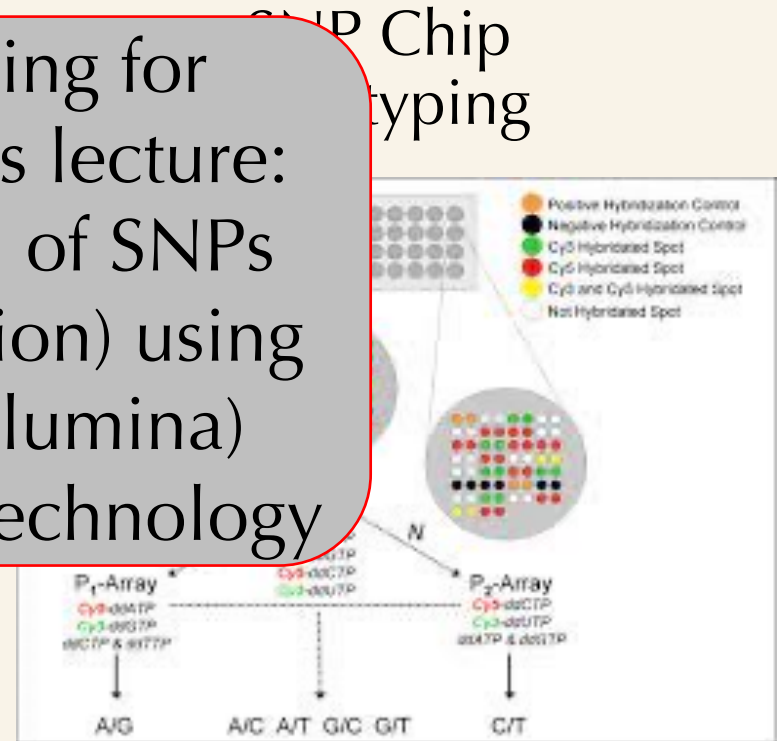
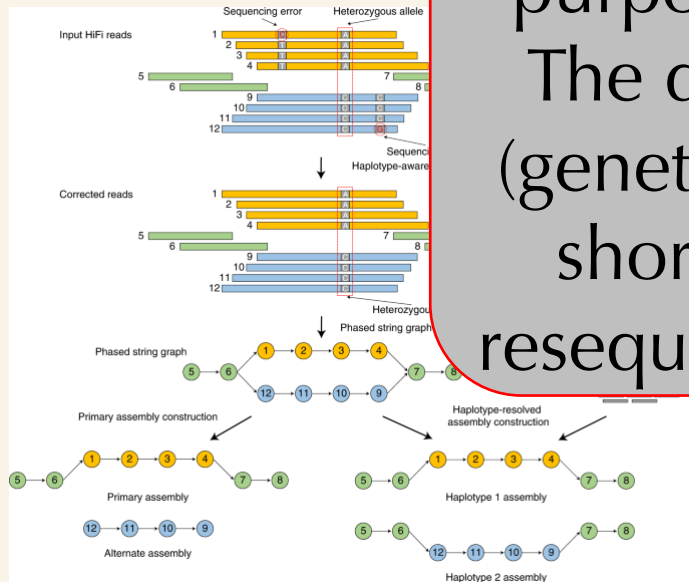
SNP Chip/DNA micro
array genotyping



Yet there are different ways of assessing genetic variation

i.e. *de novo*
haplotype assembly

Variant calling for
purpose of this lecture:
The detection of SNPs
(genetic variation) using
short read (Illumina)
resequencing technology



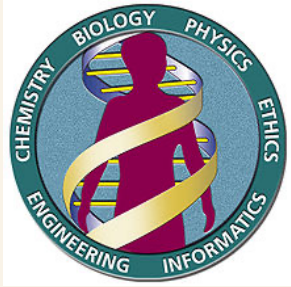
Questions?



2) Variant calling pipelines/methods and limitations

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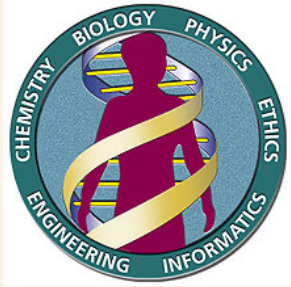
Variant calling **always** starts with a reference genome



Assembly of the first complex vertebrate genome
Human genome assembly project (2003)
Not easily repeated: it was massive task
Nowadays; much cheaper and faster

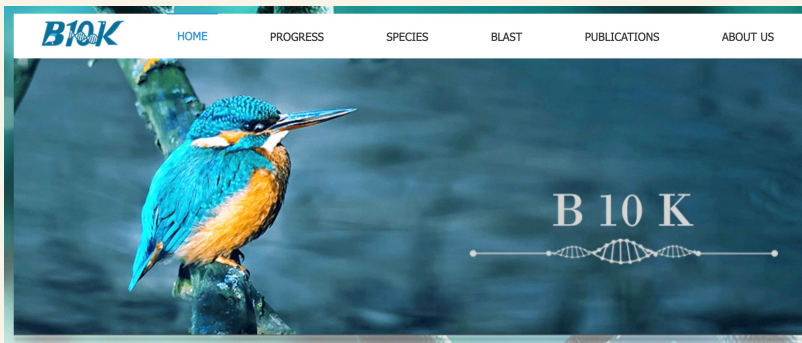
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Great push to provide reference genomes for many organisms!



B10 K: 10.000 bird genomes

*Deep evolutionary understanding
of the entire living avian class*

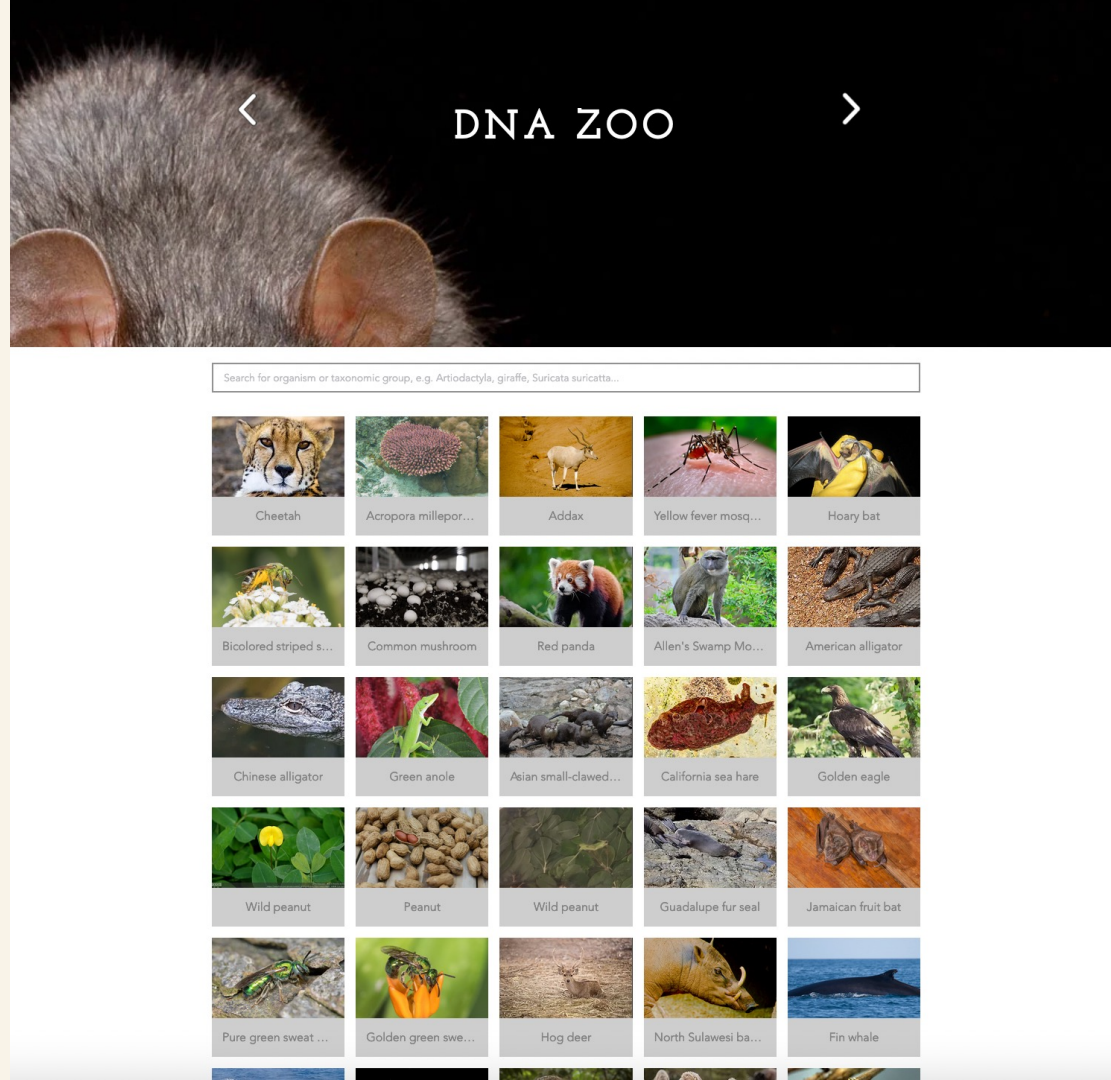
<https://b10k.genomics.cn/>



The *DNA Zoo*

facilitates conservation efforts by releasing high-quality genomics resources.

<https://www.dnazoo.org>



The most ambitious: Earth Biogenome Project



[ABOUT EBP](#) [GOVERNANCE](#) [COMMITTEES](#) [REPORTS](#) [MEDIA](#) [CONTACT](#)

CREATING A NEW FOUNDATION FOR BIOLOGY

**Sequencing Life for the
Future of Life**

<https://www.earthbiogenome.org/>

The most ambitious: Earth Biogenome Project



ABOUT EBP GOVERNANCE COMMITTEES REPORTS MEDIA CONTACT

EBP: moonshot for biology, aims to characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.

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The most ambitious: Earth Biogenome Project



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EBP: moonshot for biology, aims to characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.

The vision: to create a new foundation for biology, with new solutions for preserving biodiversity and sustaining human societies.

But what
is a
reference
genome?

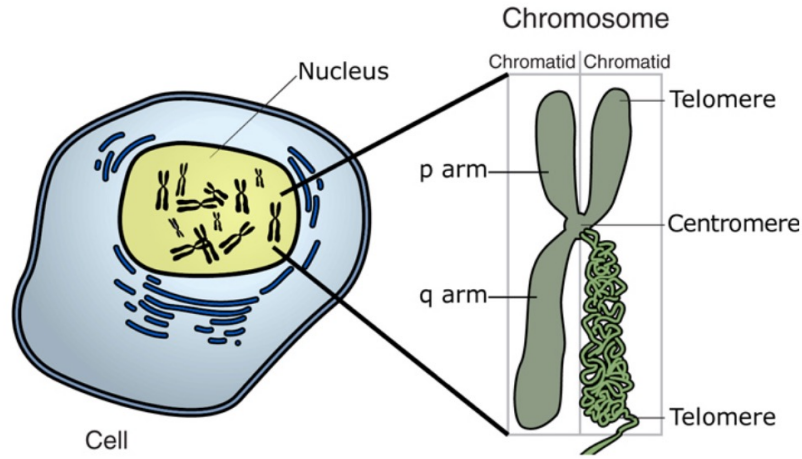
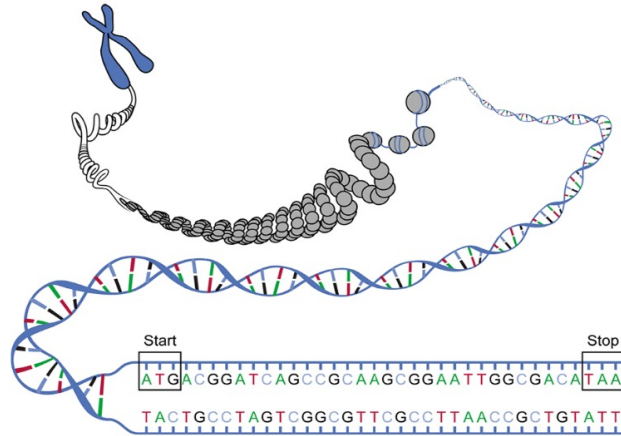
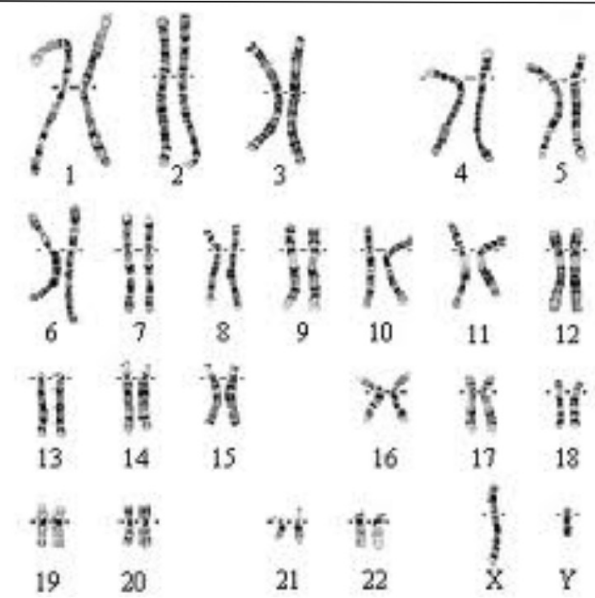


Image adapted from: National Human Genome Research Institute.



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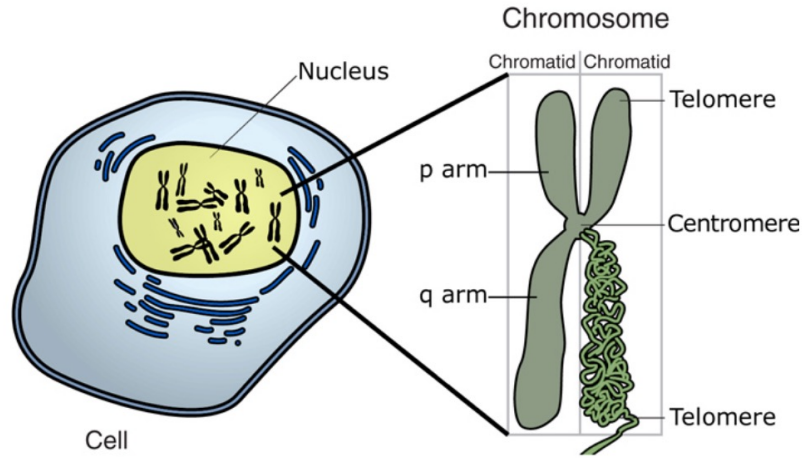
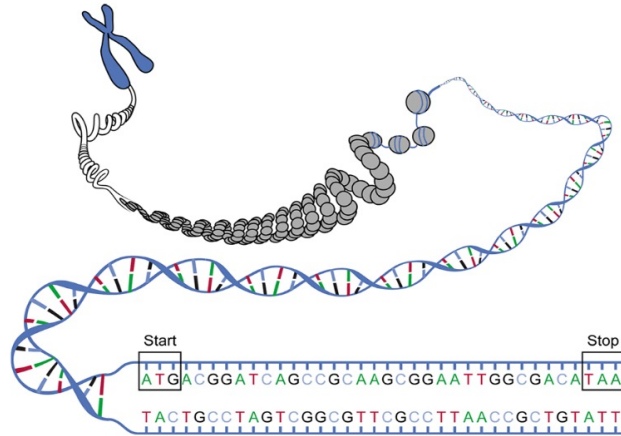
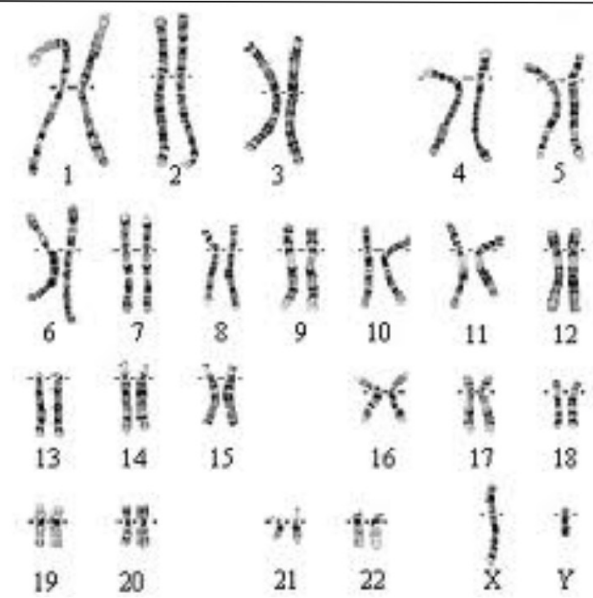


Image adapted from: National Human Genome Research Institute.



Digital representation /
abstraction of a physical,
biological phenomenon

A reference genome is ...

Usually from a single individual

Result of a genome assembly process -> errors are introduced

Of varying quality, that can vary from organism to organism

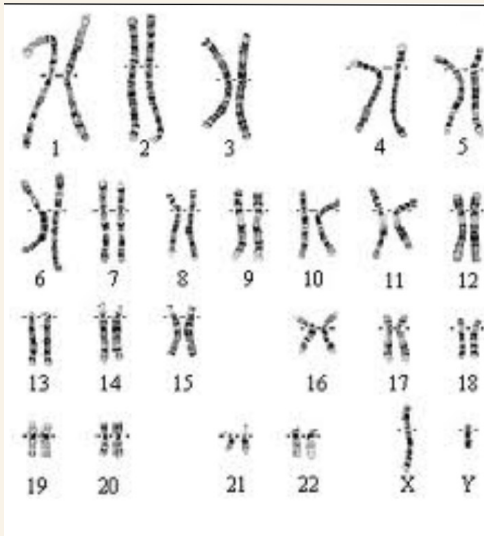
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Of varying quality, that can vary from organism to organism

Digital version of the
genome



Sequencing
and
assembly

>Chr01
ACTACGTATATAGCATGATCATGCATGATACATGGCTAGT...
>Chr02
ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...
>Chr03
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...
>Chr04
CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...
... ..

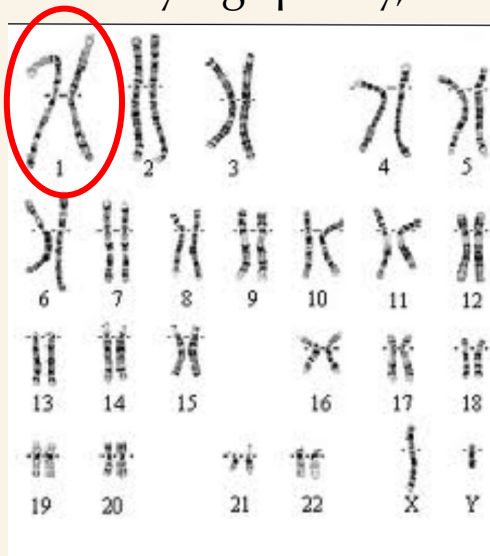
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>Chr02

ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...

>Chr03

ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...

>Chr04

CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...

... ..

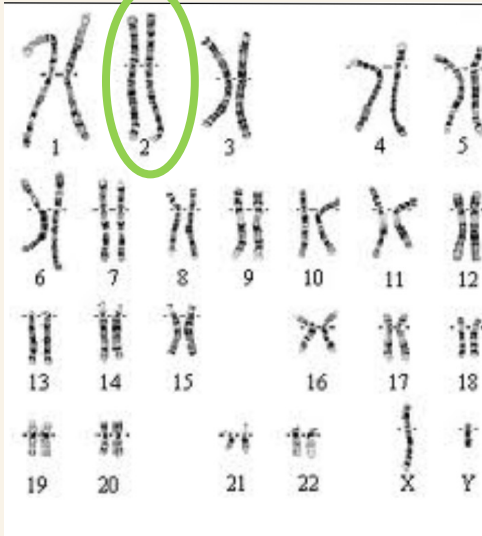
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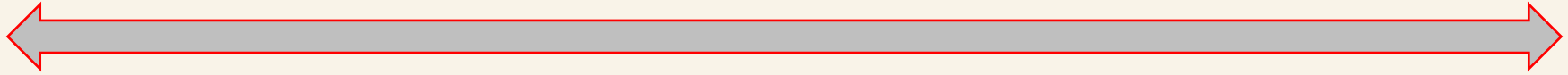
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and
assembly

>Chr01
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>Chr02
ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...
>Chr03
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...
>Chr04
CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...
... ..

Quality scale of reference genomes

Poor

Good

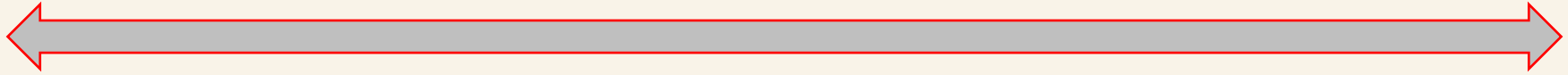


Chromosomes unclear
Thousands of loose fragments
Gaps (*nnnnn*) in sequences
Missing nucleotides

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Chromosomes unclear
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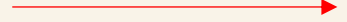
Chromosomes resolved
Continuous sequences
No gaps
Most nucleotides covered,
including centromeres and
repetitive regions

A reference genome has a 2D coordinate system

>Chr01

ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...

123456789.....



Millions of nucleotides/bases

Note: some different coordinate systems exist (i.e. starting at 0 or 1)
or using the base or space as “location”

A reference genome has a 2D coordinate system

```
>Chr01
```

```
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
```

```
123456789.....
```

Note: some different coordinate systems exist (i.e. starting at 0 or 1)
or using the base or space as “location”

i.e. A-C-T-A-C-G-T-A

```
 1 2 3 4 5 6 7 8  
 1 2 3 4 5 6 7
```

Such different systems are usually automatically recognized by different software

A multiple alignment towards a reference

>Chr01
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...

Start (12) Stop (31)

Read 1 AGCATGATCATGCATGATGA

Read 2 GCATGATGATCATGCATGATACATGG

Read 3 TGATGATCATGCATGATACATGGCTAGT

Short read sequencing data is compared to the reference (looking for a "match")

We first need such alignment before we can analyse variation

Read variation is analysed within this alignment context

The diagram illustrates sequence alignment with two SNPs and three reads. The reference sequence is >Chr01 ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT... The first SNP is at position 23 (G/T) and the second is at position 40 (G/A). Read 1 (AGCATGATCATTCATGATGA) has a T at position 23. Read 2 (GCATGATGATCATGCATAATACATGG) has an A at position 40. Read 3 (TGATGATCATGCATAATACATGGCTAGT) has an A at position 40. Red arrows indicate the start and stop of the alignment context. Green arrows indicate the positions of the SNPs.

Start

SNP (G/T, 23)

Stop

SNP (G/A, 40)

>Chr01
ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCAT**G**ATACATGGCTAGT...

Read 1 AGCATGATCAT**T**CATGATGA

Read 2 GCATGATGATCATGCAT**A**ATACATGG

Read 3 TGATGATCATGCAT**A**ATACATGGCTAGT

An accurate alignment is *essential* before we can trust any variant

Read variation is analysed within this alignment context

Diagram illustrating read alignment and SNP identification:

Reference sequence: >Chr01
ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCAT**G**ATACATGGCTAGT...

SNP (G/T, 23) and SNP (G/A, 40) are indicated by green arrows pointing to the variant bases in the reference sequence.

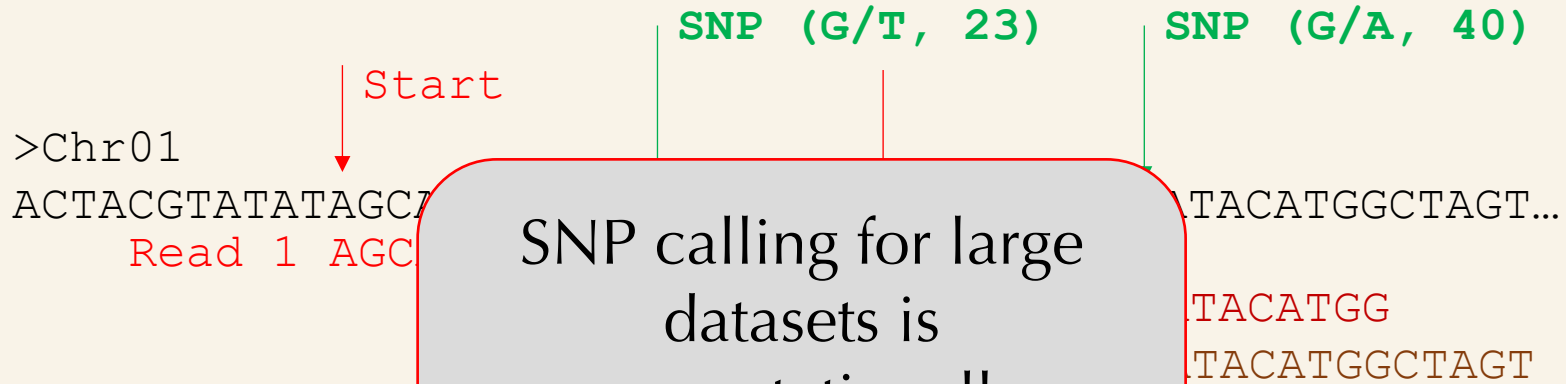
Read 1: AGCATGATCAT**T**CATGATGA

Read 2: GCATGATGATCATGCAT**A**ATACATGG

Read 3: TGATGATCATGCAT**A**ATACATGGCTAGT

We usually analyse *millions to billions* of reads and compare these to reference genomes that consist of **billions** of nucleotides/bases (Human genome ~3Gb)

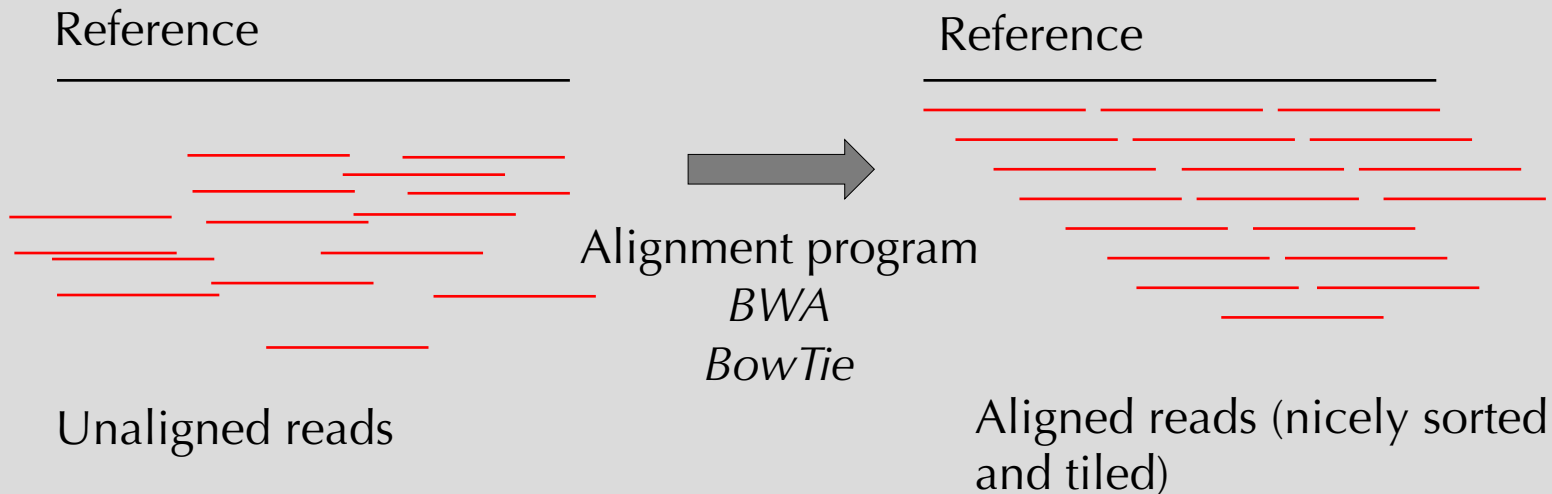
Read variation is analysed within this alignment context



We usually compare the reference sequence with millions of reads and that consist of **billions** of nucleotides/bases (Human genome ~3Gb)

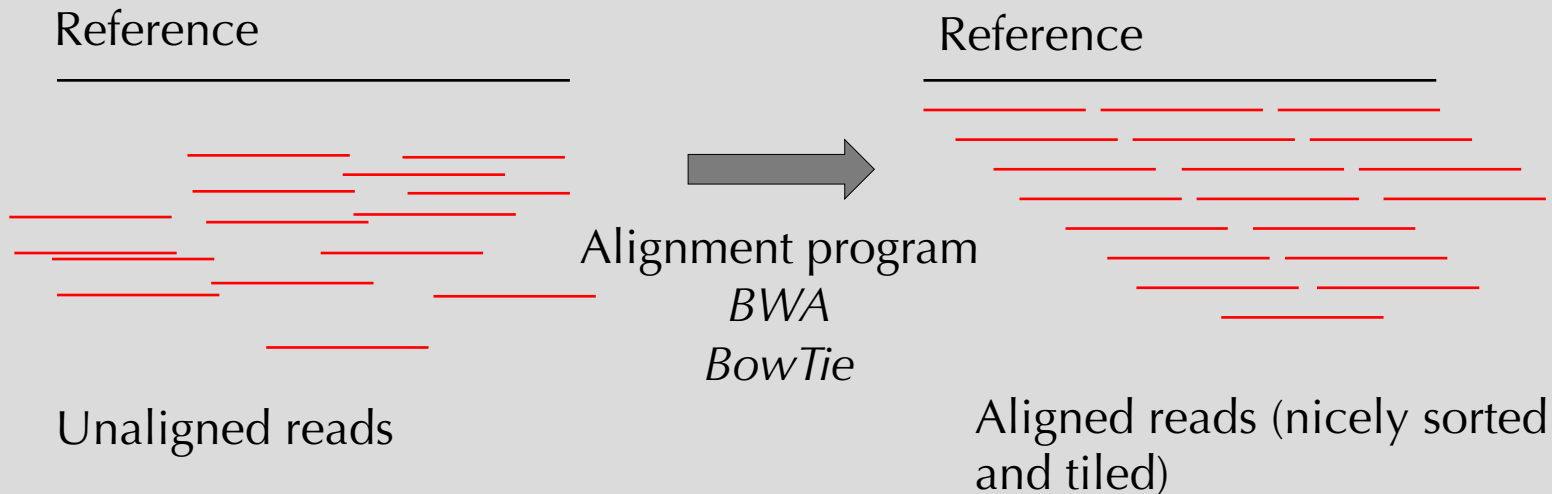
Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!



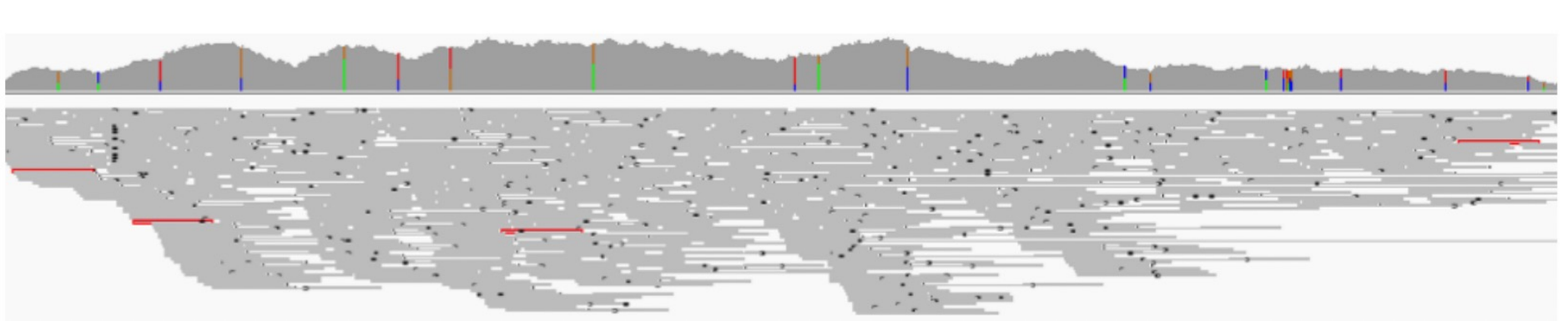
Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!



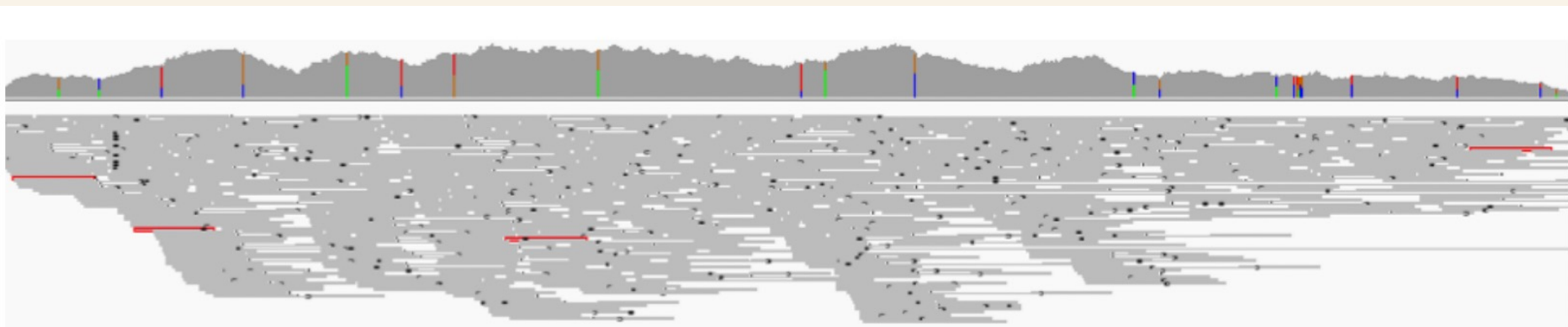
Standard program settings are usually sufficient

Visualisation of thousands of reads



Visualisation of thousands of reads

Genetic variation (SNPs) colours reflect which bases are variable (A-C, A-T, G-C, etc)



After aligning, we need another program to determine which bases are variable:

A SNP caller

SNP calling programs

Table 1. A brief summary of different tools.

caller	Bcftools	16GT	Freebayes	VarScan2	GATK
Code	C	Perl	C++	Java	Java
Model	HMM & MAQ	16-genotype probabilistic	Bayesian	heuristic algorithm	Bayesian
Sampling	Single & multiple	Single	Single	Single & multiple	Single & multiple
Variants	SNPs & indels	SNPs & indels	SNPs & indels&MNPs	SNPs & indels	SNPs & indels
Features	Sorting, indexing, etc.	easy to use, timesaving	straightforward	meet desired thresholds for read depth, base quality, variant allele frequency, and statistical significance	Realignment, per base recalibration, VQSR
Reference	Danecek et al., 2017 [15]	Luo et al., 2017 [19]	Garrison and Marth, 2012 [18]	Koboldt et al., 2012 [16]	Mckenna et al., 2010 [14]

<https://doi.org/10.1371/journal.pone.0262574.t001>

Liu J, Shen Q, Bao H (2022)

Many programs exist, and there is *continuous* development

For instance Bcftools/16GT are now recommended

Yet use of GATK is wide-spread (oldest, developed by Broad institute, good documentation)

What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

>Chr01

ACTACGTATATAGCATGATCAT**G**ATGATGATCATGCATGATACATGGCTAGT...

Read 1 AGCATGATCAT**T**ATGATGA

Is this real or not?

What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

```
>Chr01
```

```
ACTACGTATATAGCATGATCATGATGATGATCATGCATGATACATGGCTAGT...
```

```
Read 1 AGCATGATCATTATGATGA
```

Is this real or not?

Sequencing data (as any type of data) comes with errors (wrong bases called) and/or uncertainty (low quality of bases) in the call

Solution? Generate LOTS more data!

What does a variant caller do?

With more data (read), more certainty is obtained: ***fold coverage***

```
>Chr01
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
  Read 1 AGCATGATCATTCATGATGA
    Read 2 ATGATCATTCATGATGATCAT
      Read 3 GATCATTCATGATGATCATGCATGAT
        Read 4 TCATTCATGATGATCATGCAT
          Read 5 CATTCATGATGATCATGCATGATACATGG
```



5-fold coverage, all the same, we are pretty certain about this call (note: we usually strive for ~20 fold coverage)

What does a variant caller do?

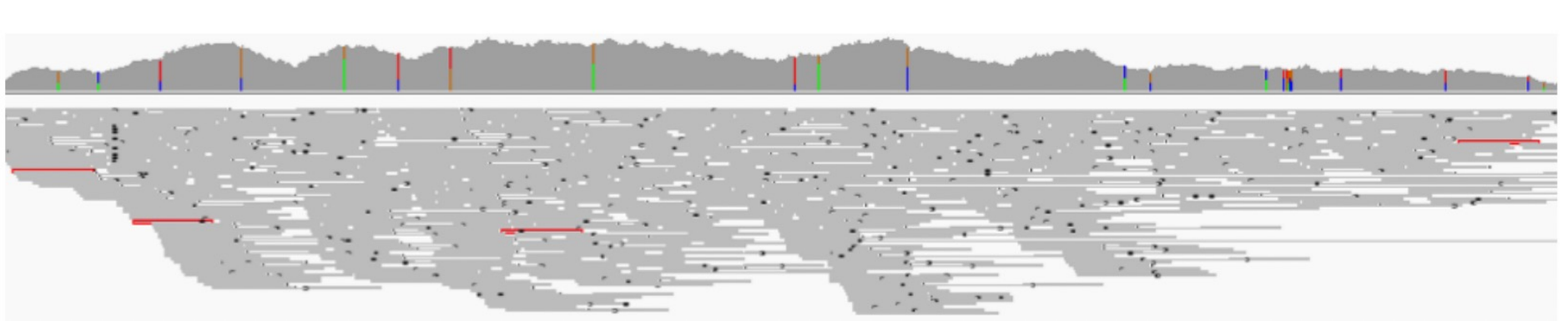
Another example

```
>Chr01
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
Read 1 AGCATGATCATTCATGATGA
Read 2 ATGATCATTCATGATGATCAT
Read 3 GATCATTCATGATGATCATGCATGAT
Read 4 TCATACATGATGATCATGCAT
Read 5 CATTCATGATGATCATGCATGATACATGG
```

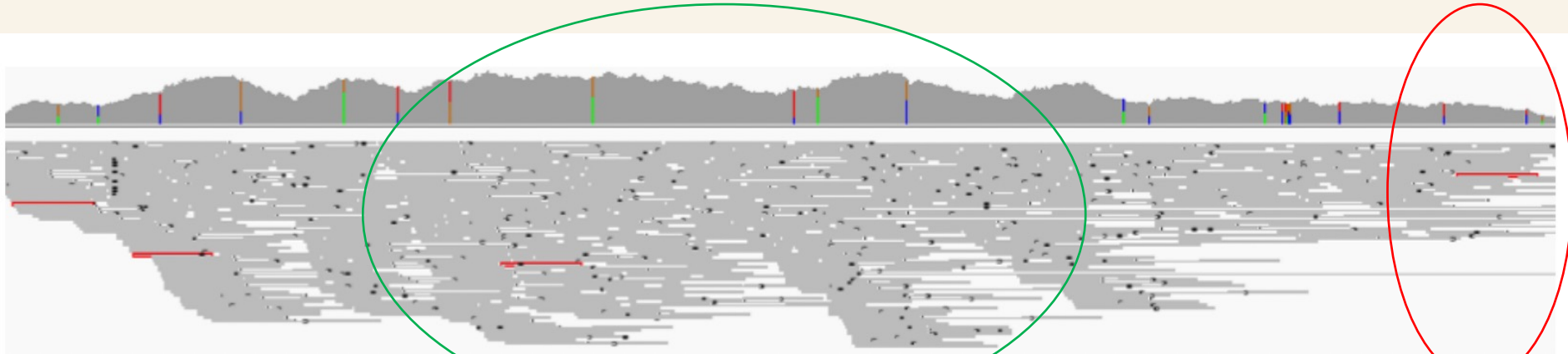
We cannot be so certain about the A, until we get more data

Coverage is the most important determinant for the quality of your data

Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity



Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity



Higher coverage

Lower coverage

Yet along a reference, you'll obtain variable coverage due to random complexity

SNP callers run complex statistical models (e.g. Bayesian or HMM models) to provide confidence in SNP calls and if they are "TRUE". They often assume correct read alignment **and** require sufficient read coverage in order to provide high-quality calls



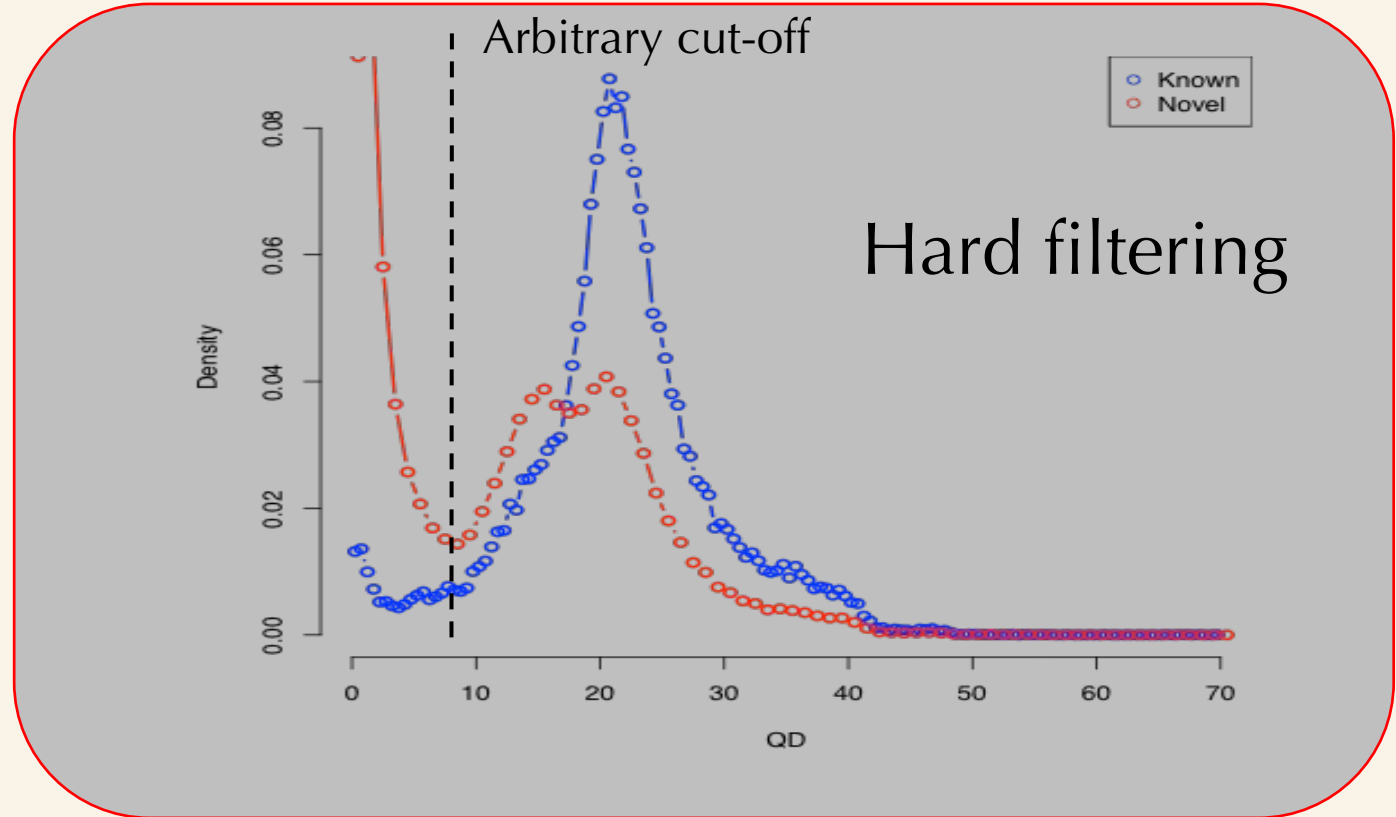
Higher coverage



Lower coverage

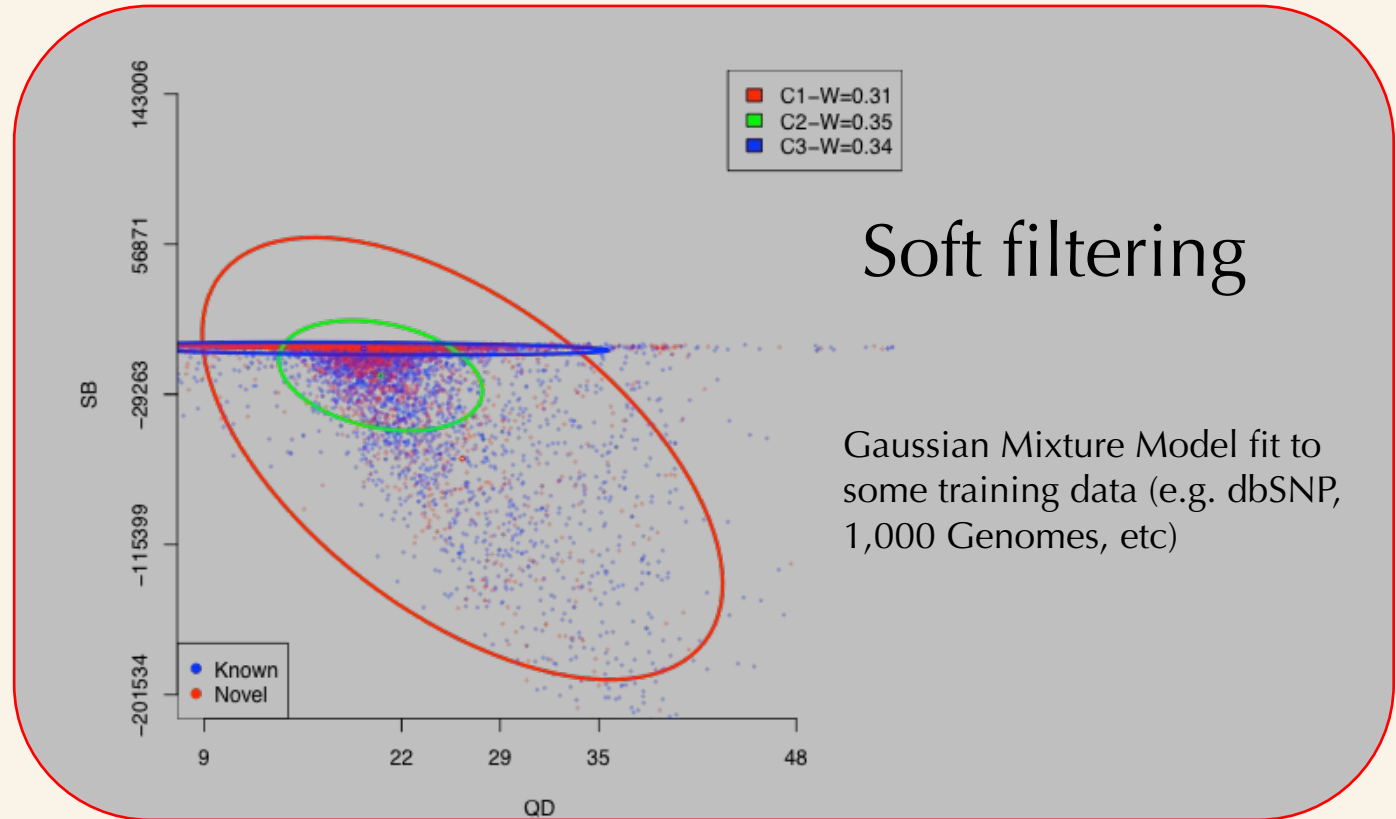
SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

We need to ***filter*** our data to only retain the high quality part of the data

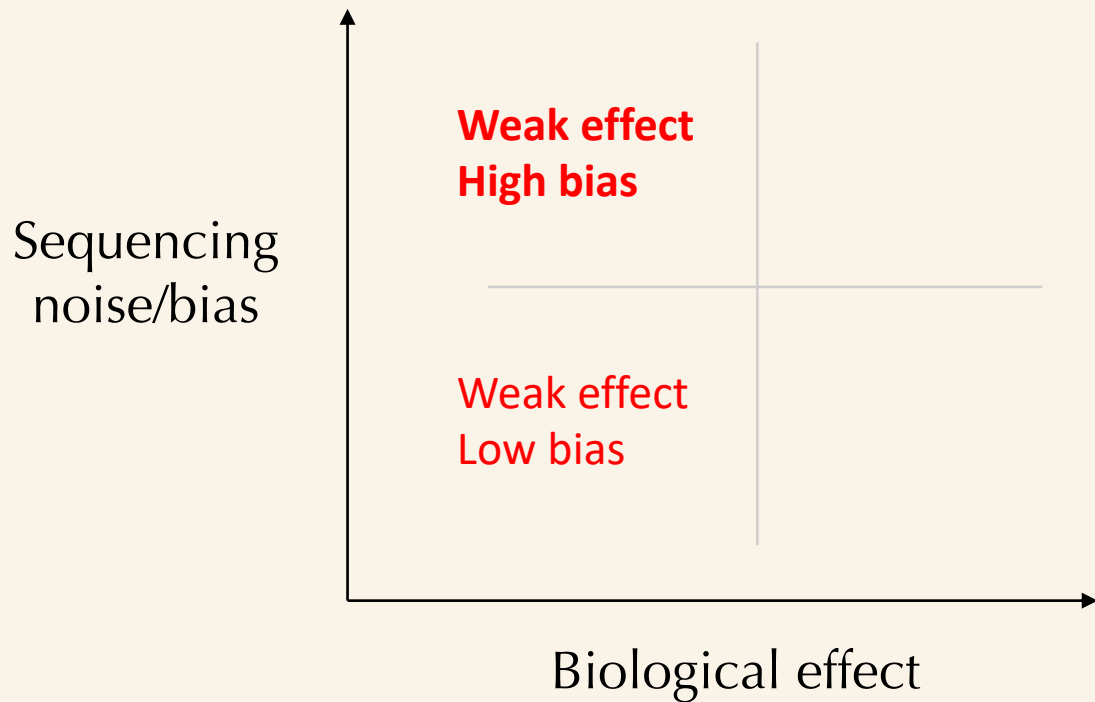


SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

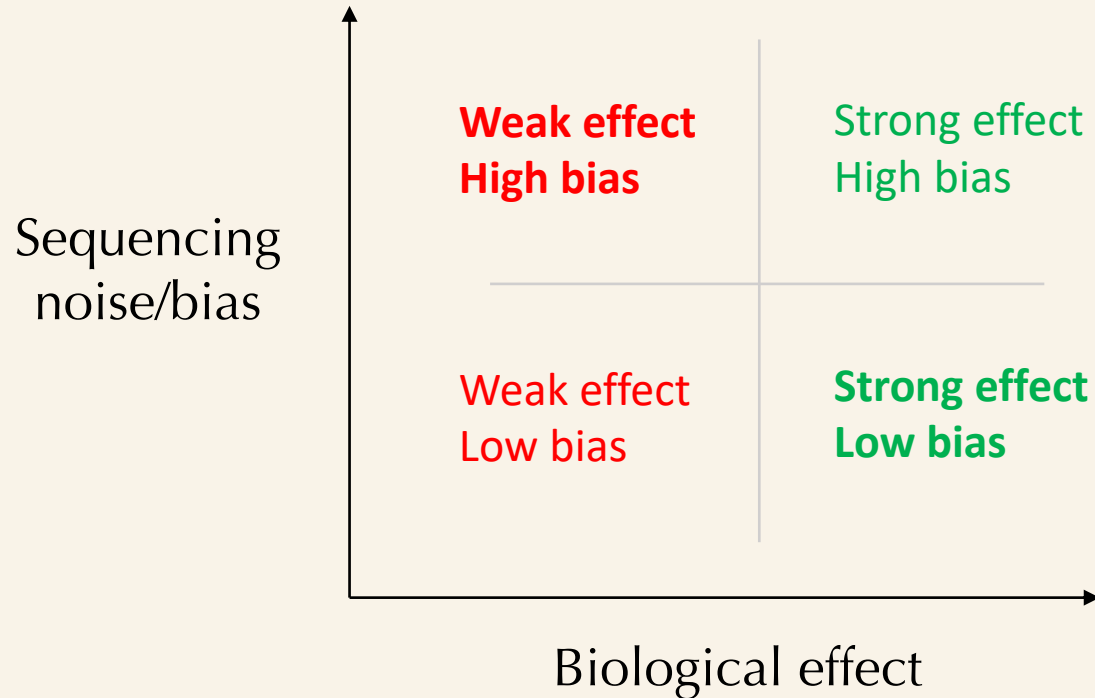
We need to ***filter*** our data to only retain the high quality part of the data



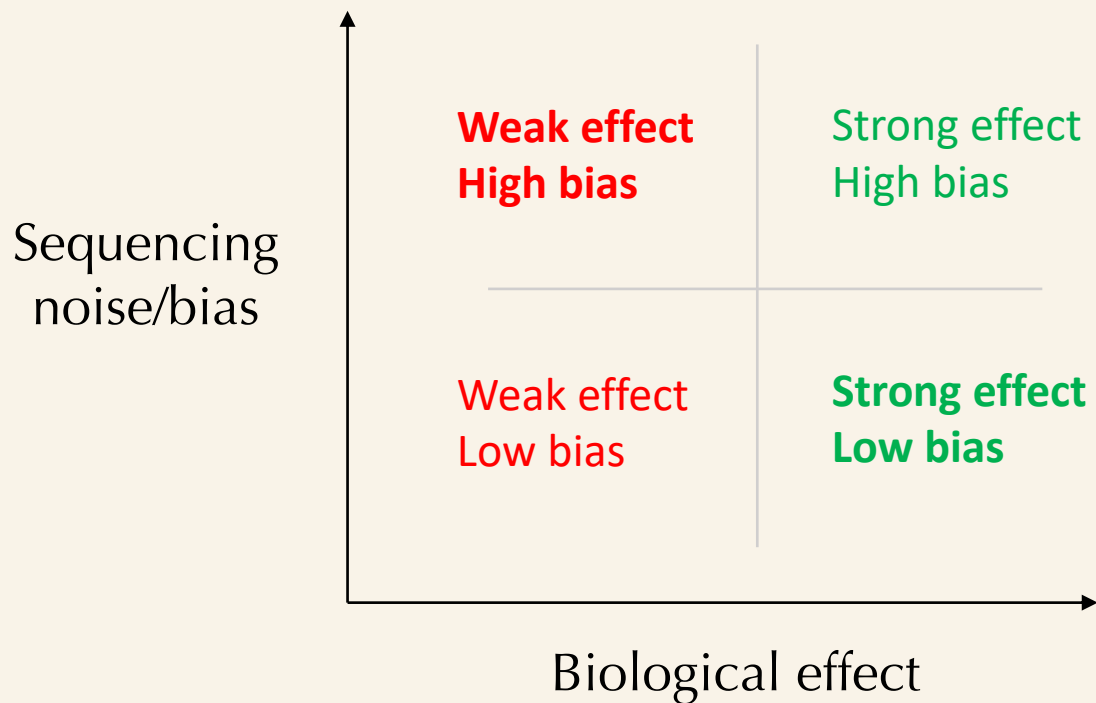
Yet there is no “fixed” approach to filtering your data



Yet there is no “fixed” approach to filtering your data



Yet there is no “fixed” approach to filtering your data



It is not always clear from the outset where you are! You need to explore your data and use preliminary analyses

Questions?



After all this, what does a variant calling pipeline look like?



Reference

Reads



e.g.
population
data

After all this, what does a variant calling pipeline look like?



Reference

Mapping/aligning

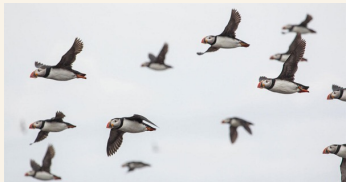
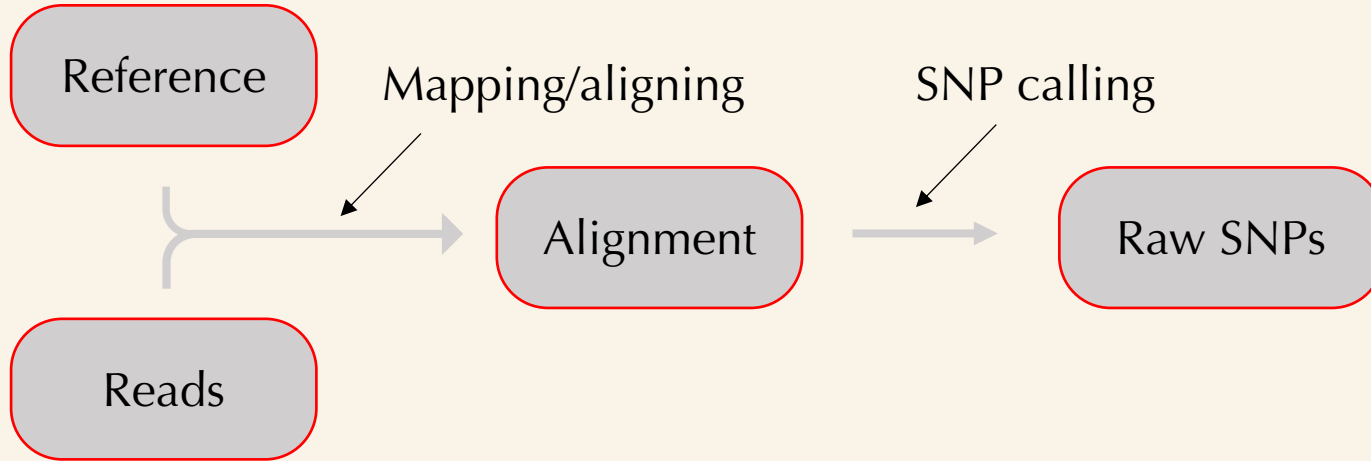
Alignment

Reads



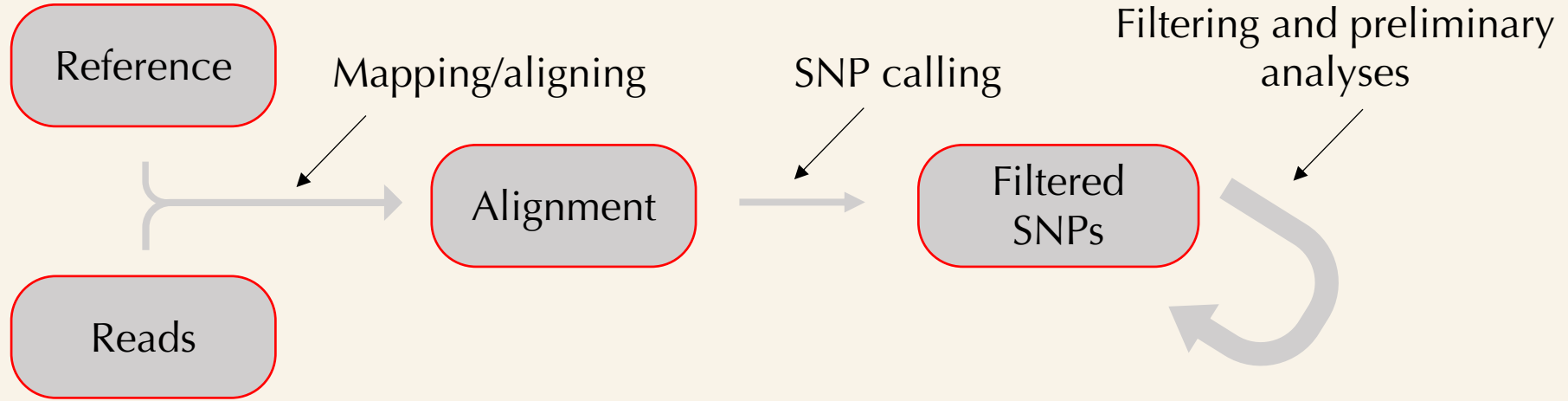
e.g.
population
data

After all this, what does a variant calling pipeline look like?



e.g.
population
data

After all this, what does a variant calling pipeline look like?

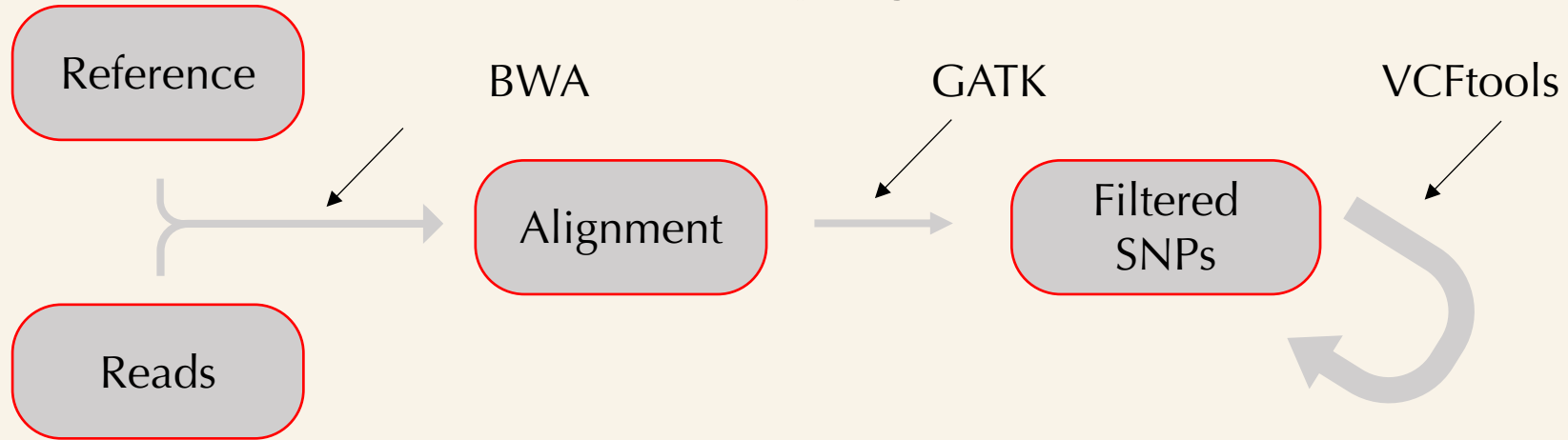


e.g.
population
data

After all this, what does a variant calling pipeline look like?

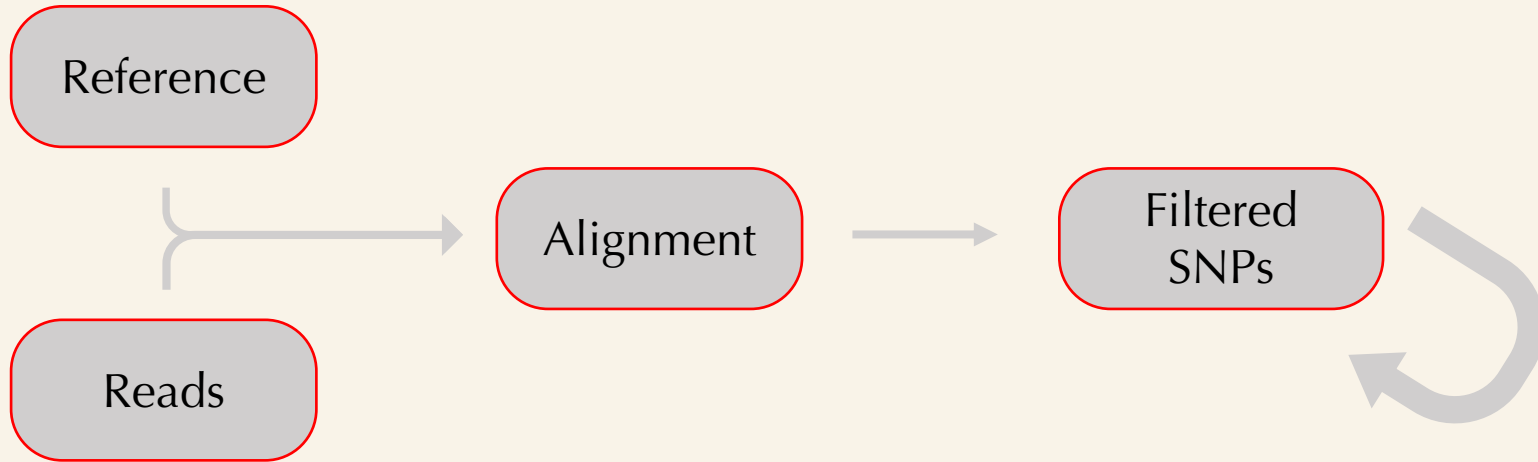


A selection of programs that can be used

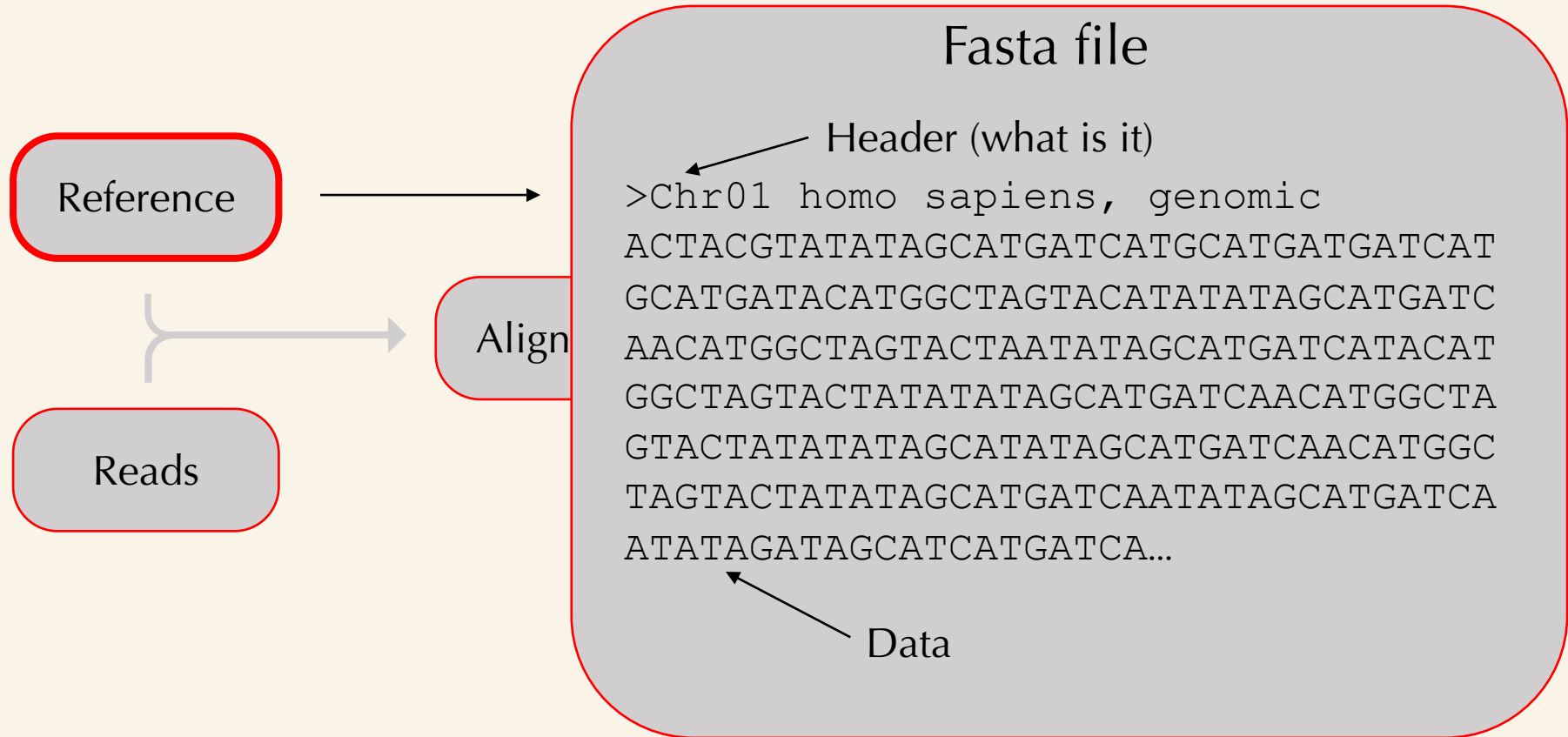


e.g.
population
data

Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!

Alignment



BAM file (binary alignment file)

Kind of data

“Header” with information about the file

```
@HD VN:1.5 GO:none SO:coordinate
@SQ SN:NC_004029.2 LN:16565
@RG ID:L1i1_AGAACCG SM:WLR001 LB:L1i1_AGAACCG PU:Nistelberger-DNA1-2016-04-15
@RG PL:ILLUMINA PG:bwa
@PG ID:bwa PN:bwa VN:0.7.17-r1188 CL:bwa samse Orosvlmt.fasta
@PG ID:GATK IndelRealigner VN:3.6-0-g89b7209 CL:knownAlleles=[] targetIntervals=WLR001/Wal_m
@PG ID:samtools CL:samtools view -H WLR001.Wal_mt.realigned.bam
```

Reference

Sample name

Each of these steps requires specific files to work with!

Alignment

BAM file (binary alignment file)

Readname

Follow by data with information about each read alignment

M_D00564:55:C9FG3ANXX:7 0
M_D00564:55:C9FG3ANXX:7 0
M_D00564:55:C9FG3ANXX:7 0

NC_004029.2
NC_004029.2
NC_004029.2

419 37 91M
474 37 58M
515 37 56M

Start of alignment

Matching bases

TAAAAAGCTGCCGCTAATACAAAAATATACTACGAAAGTGACT
TTACACGACAGCTAAGACCCAAACTGGGATTAGATACCCCA
CTATGCTTAGCCATAAACACAAATAATTTGCACAACAAAAATT

Reference name

Quality of alignment (37 is max)

CIGAR string (56 matching bases)

Each of these steps requires specific files to work with!

SNP data



VCF file (Variant call format)

Again, a “Header” with lots of information about the file

```
##fileformat=VCFv4.2
##ALT=<ID=NON_REF,Description="Represents any possible alternative allele not already represented at this location">
##FILTER=<ID=LowQual,Description="Low quality">
##FILTER=<ID=PASS,Description="All filters passed">
##FORMAT=<ID=AD,Number=R,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth (reads with MQ=255 or with bad mates are
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=MIN_DP,Number=1,Type=Integer,Description="Minimum DP observed within the GVCF block">
##FORMAT=<ID=PL,Number=G,Type=Integer,Description="Normalized, Phred-scaled likelihoods for genotypes as defined in
##FORMAT=<ID=RGQ,Number=1,Type=Integer,Description="Unconditional reference genotype confidence, encoded as a phred
##FORMAT=<ID=SB,Number=4,Type=Integer,Description="Per-sample component statistics which comprise the Fisher's Exac
##GATKCommandLine=<ID=GenomicsDBImport,CommandLine="GenomicsDBImport --genomicsdb-workspace-path Walrus_DB --varian
##GATKCommandLine=<ID=GenotypeGVCFs,CommandLine="GenotypeGVCFs --output Walrus_MT.vcf.gz --variant gendb://Walrus_D
##GATKCommandLine=<ID=HaplotypeCaller,CommandLine="HaplotypeCaller --sample-ploidy 1 --emit-ref-confidence GVCF --c
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##INFO=<ID=BaseQRankSum,Number=1,Type=Float,Description="Z-score from Wilcoxon rank sum test of Alt Vs. Ref base qu
```

Each of these steps requires specific files to work with!

SNP data



VCF file (Variant call format)

Followed by the data:

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO
NC_004029.2	131	.	T	C	356.22	.	AC=1;AF=0.022;AN=45;DP=143;FS=0.000;MLEAC=1;MLEAF=0
NC_004029.2	162	.	T	C	18479.23	.	AC=15;AF=0.333;AN=45;BaseQRankSum=0.00;DP=543;FS=0
NC_004029.2	198	.	C	T	608.22	.	AC=1;AF=0.022;AN=45;DP=410;FS=0.000;MLEAC=1;MLEAF=0
NC_004029.2	387	.	G	A	547.22	.	AC=1;AF=0.022;AN=45;DP=408;FS=0.000;MLEAC=1;MLEAF=0
NC_004029.2	616	.	T	C	235.62	.	AC=1;AF=0.022;AN=45;DP=406;FS=0.000;MLEAC=1;MLEAF=0
NC_004029.2	741	.	C	T	819.22	.	AC=1;AF=0.022;AN=45;DP=412;FS=0.000;MLEAC=1;MLEAF=0
NC_004029.2	743	.	C	T	819	.	AC=1;AF=0.022;AN=45;DP=413;FS=0.000;MLEAC=1;MLEAF=0



Reference name

Each of these steps requires specific files to work with!

SNP data

VCF file (Variant call format)

Followed by the data:

GenoType: Allele Depth: Read Depth (DP): Genotype Quality: Phred-scaled Likelihood

FORMAT	WLR001	WLR002	WLR003	WLR004	WLR005
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:2,0:2:90:0,90	0:0,0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:4,0:4:99:0,135	0:1,0:1:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:5,0:5:46:0,46	0:0,0:0:0:0:0,0	0:2,0:2:90:0,90	0:2,0:2:90:0,90
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0:0,0	0:2,0:2:45:0,45	0:2,0:2:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0:0,0	0:1,0:1:42:0,42	0:1,0:1:42:0,42
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:1,0:1:42:0,42	0:1,0:1:42:0,42
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:3,0:3:99:0,119	0:3,0:3:99:0,119
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:3,0:3:99:0,119	0:3,0:3:99:0,119
GT:AD:DP:GQ:PL	0:1,0:1:0:0:0,0	0:1,0:1:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:0:0:0,0	0:1,0:1:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0

After all this, what does a variant calling pipeline look like?



Fasta file

Reference

Mapping/aligning

Alignment

SNP calling

Filtered
SNPs

Filtering and preliminary
analyses

Reads

FastQ file

BAM file

VCF file

e.g.
population
data



Questions?



Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and pitfalls
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

