

Introduction to variant calling

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Department of Biosciences, University of Oslo (UiO)
Norway

BIOS-IN 5K/9K
4th of Nov 2024

@archaeogenomics



UiO : University of Oslo



Evolutionary Biologist

specialize in ancient DNA

Historical Ecology and Conservation Genomics
(10+ MSc, PhDs & Postdocs)

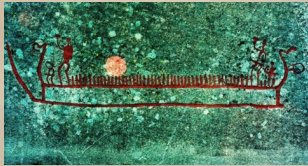
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My background:

Historical Marine Ecology

Stone Age



Subsistence



Viking Age



Early Trade



Middle Ages



"Fish Horizon"



Modern Age



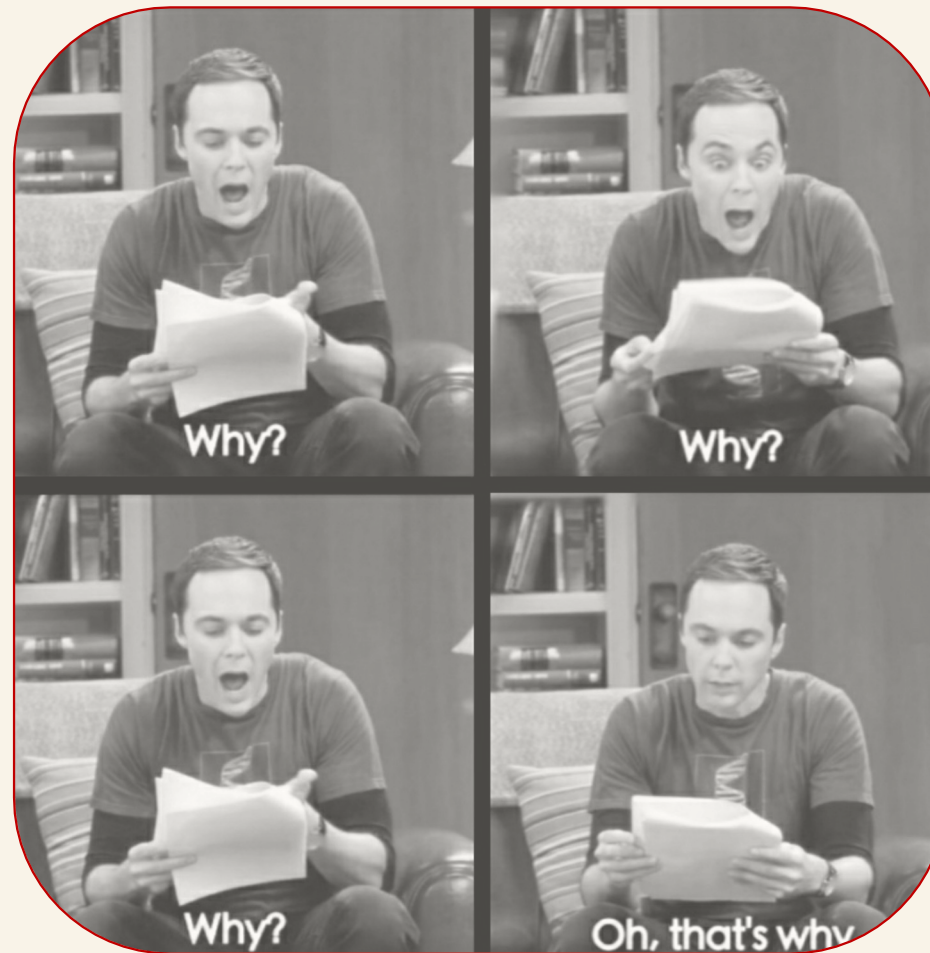
Globalisation



@archaeogenomics



As an evolutionary biologist - *Why* am I here?





Multidisciplinary research:
Archaeology
Biology
Ecology
Molecular methods/sequencing
Genomics
Bioinformatics

Today:

1) Introduction: variant calling, why do we want to do this, and what it is?

Today:

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- 2) Variant calling pipelines/methods and limitations

Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and limitations
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

Introduction

What is genetic variation?

What does genetic variation do?

Why are we interested in
genetic variation?

*Please discuss between
yourselves (two or three) for a
few minutes*

Introduction

Genetic variation (genetic differences between individuals) is everywhere!



Genetic variation at different scales:

- 1) Biological differences (phenotypes) between species



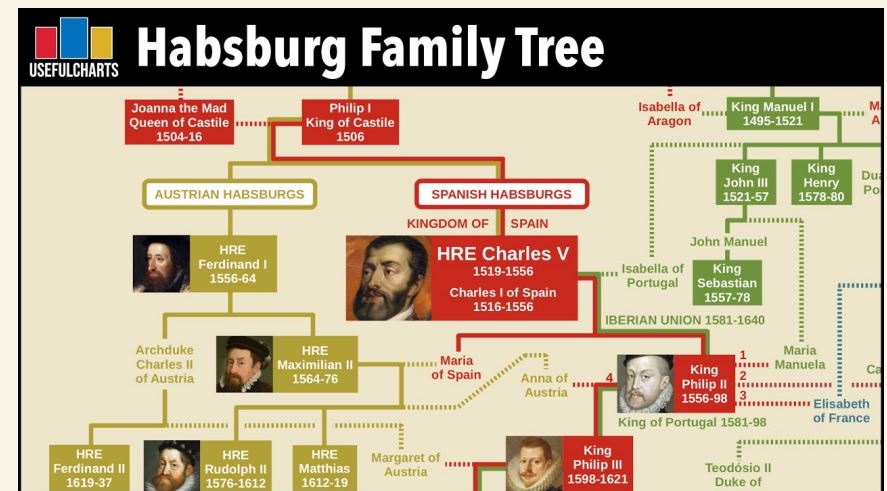
Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
- 2) Biological differences within species



Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
- 2) Biological differences within species
- 3) Patterns of relatedness between individuals/ populations (23 and me)



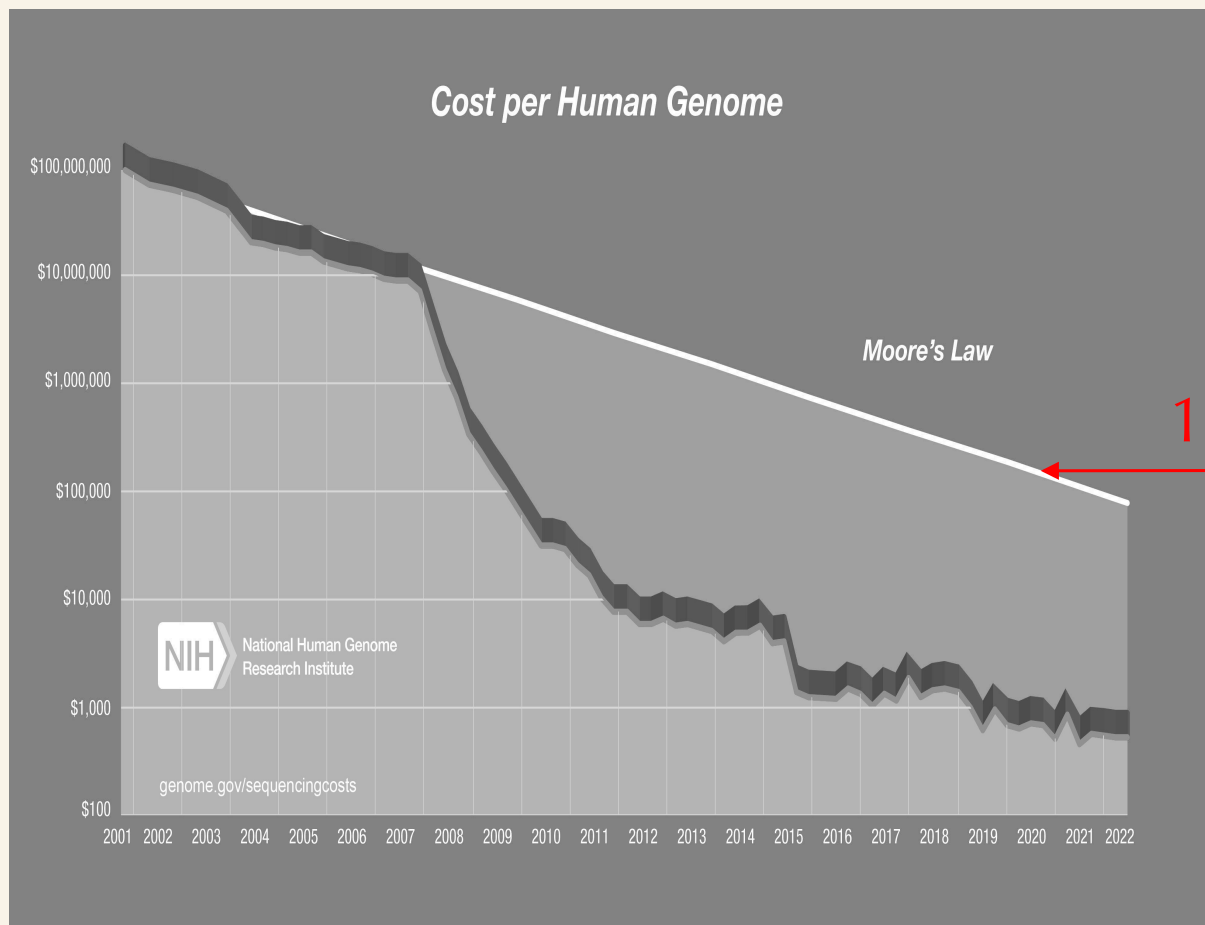
Genetic variation explains many observations within
biology

Genetic variation explains many observations within
biology

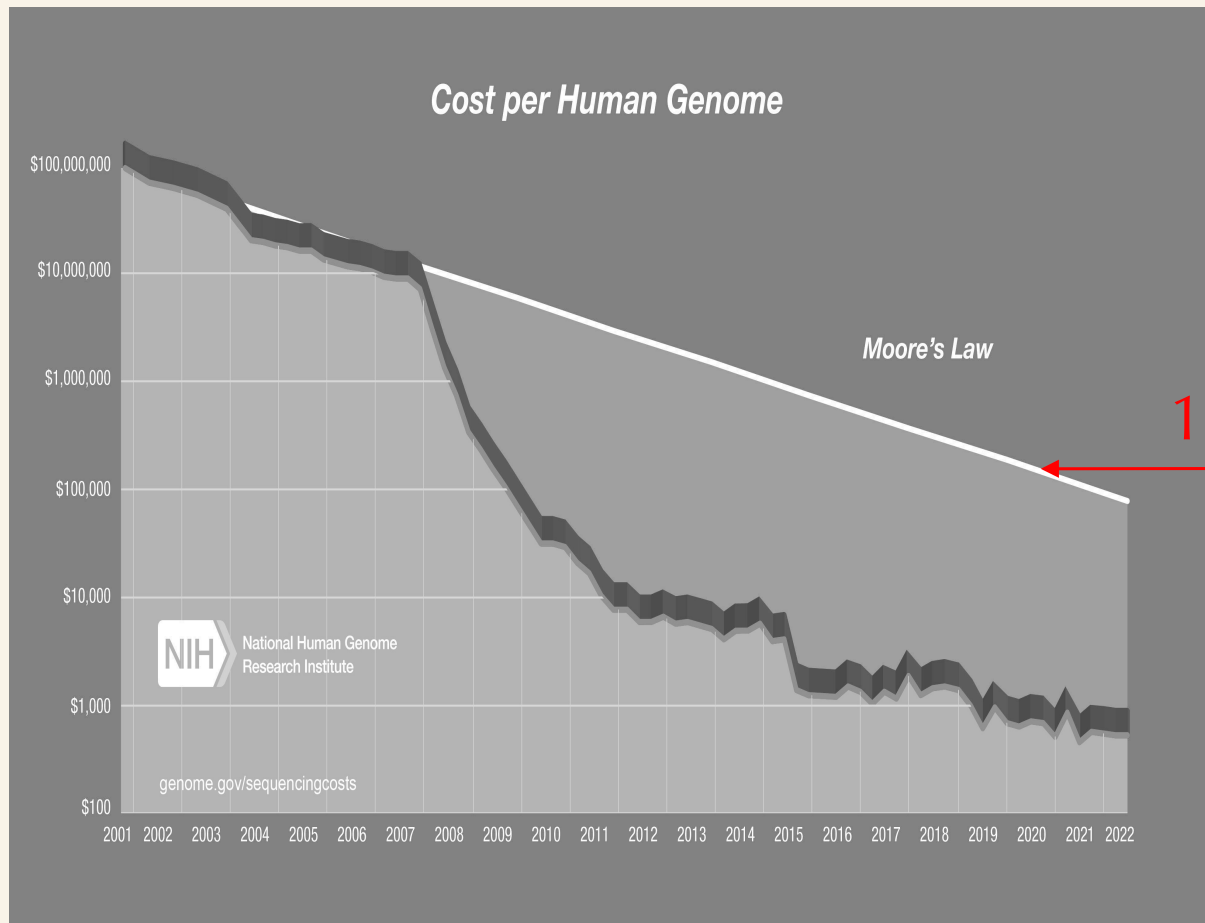
*Knowing patterns of/quantifying genetic variation has
enormous potential for a wide range of applications in
society*

*(e.g. personal medicine, forensic sciences, biodiversity
assessments, crop improvement, animal breeding, conservation
management, history & genealogy, etc etc)*

Why are we here?



Why are we here?



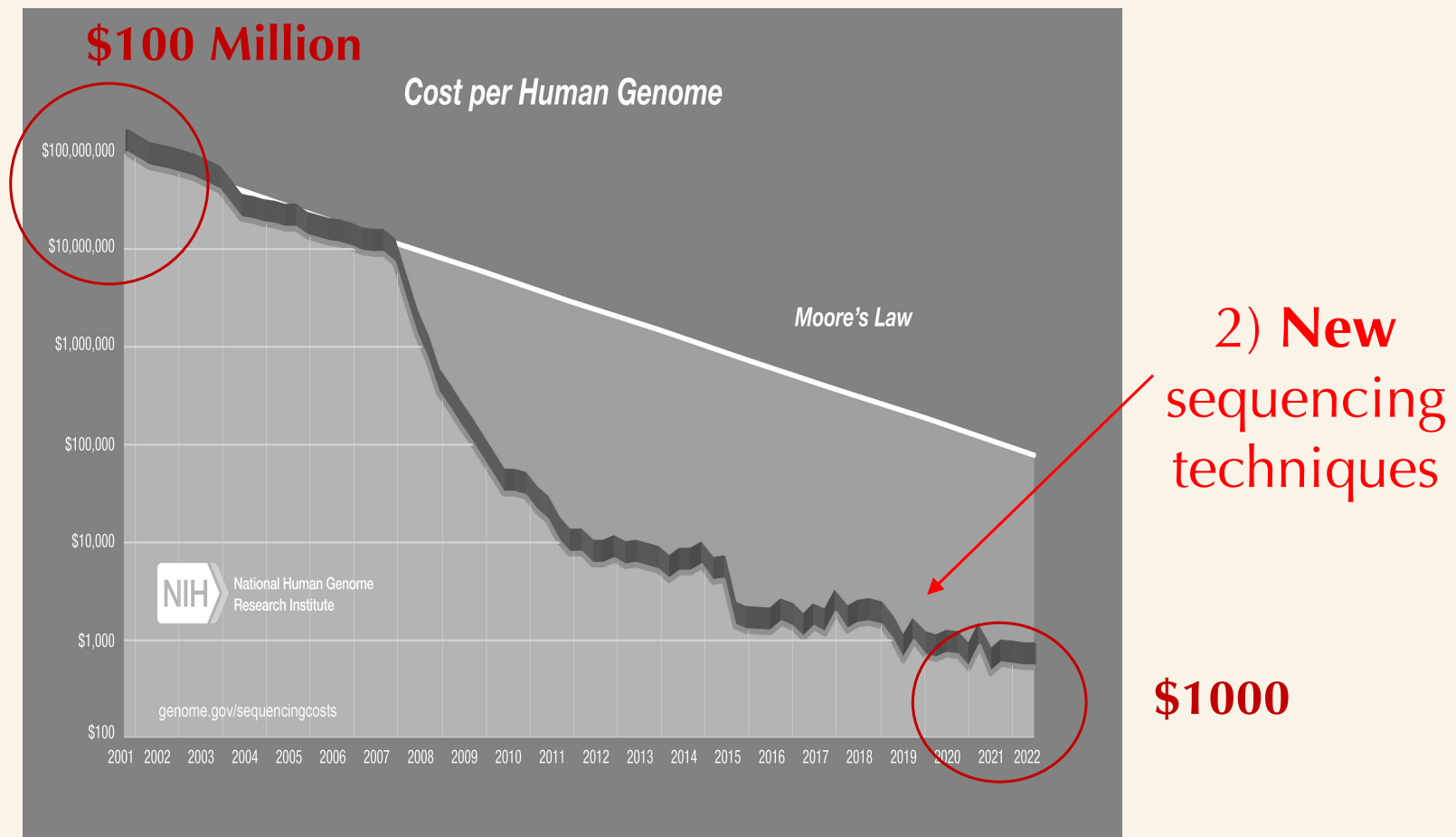
2000

1) Moore's Law

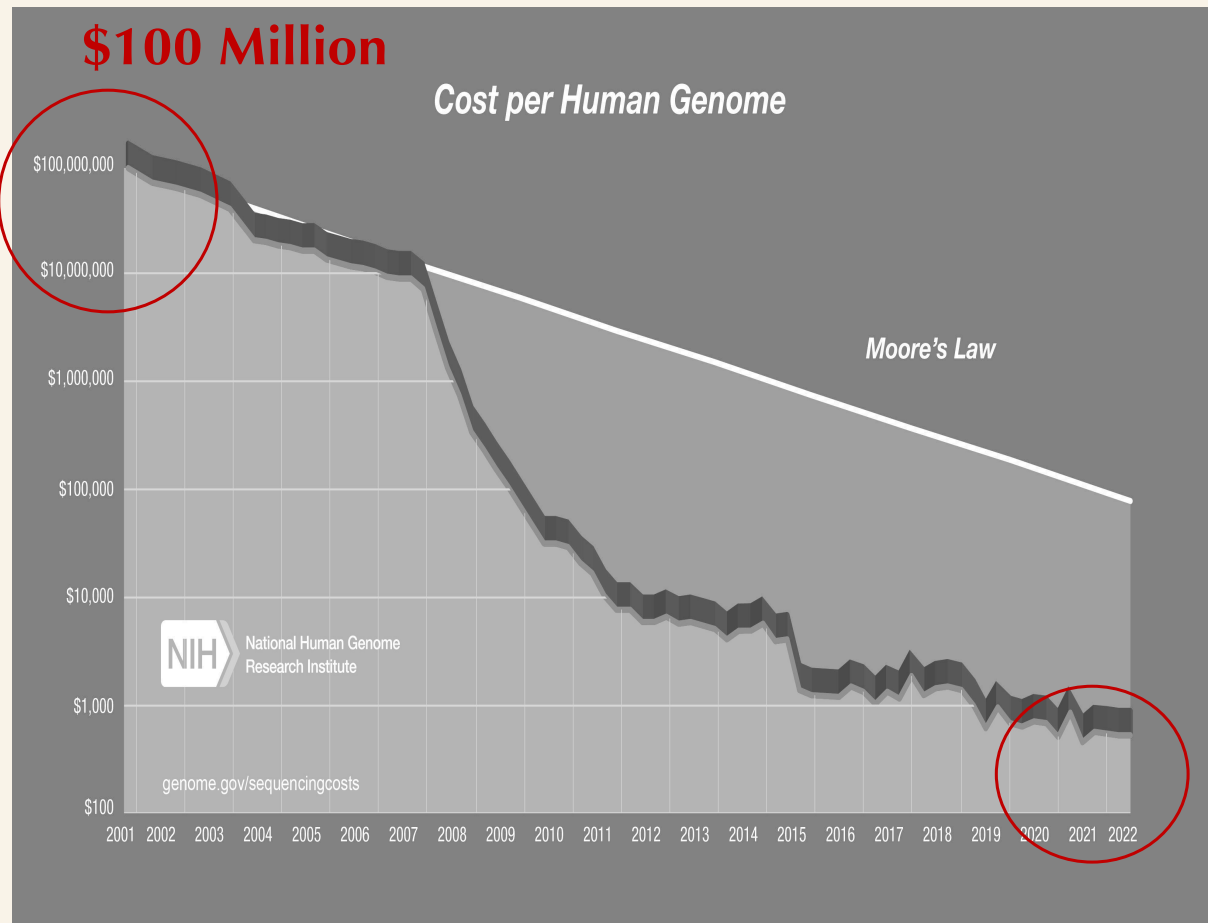


2022

Why are we here?



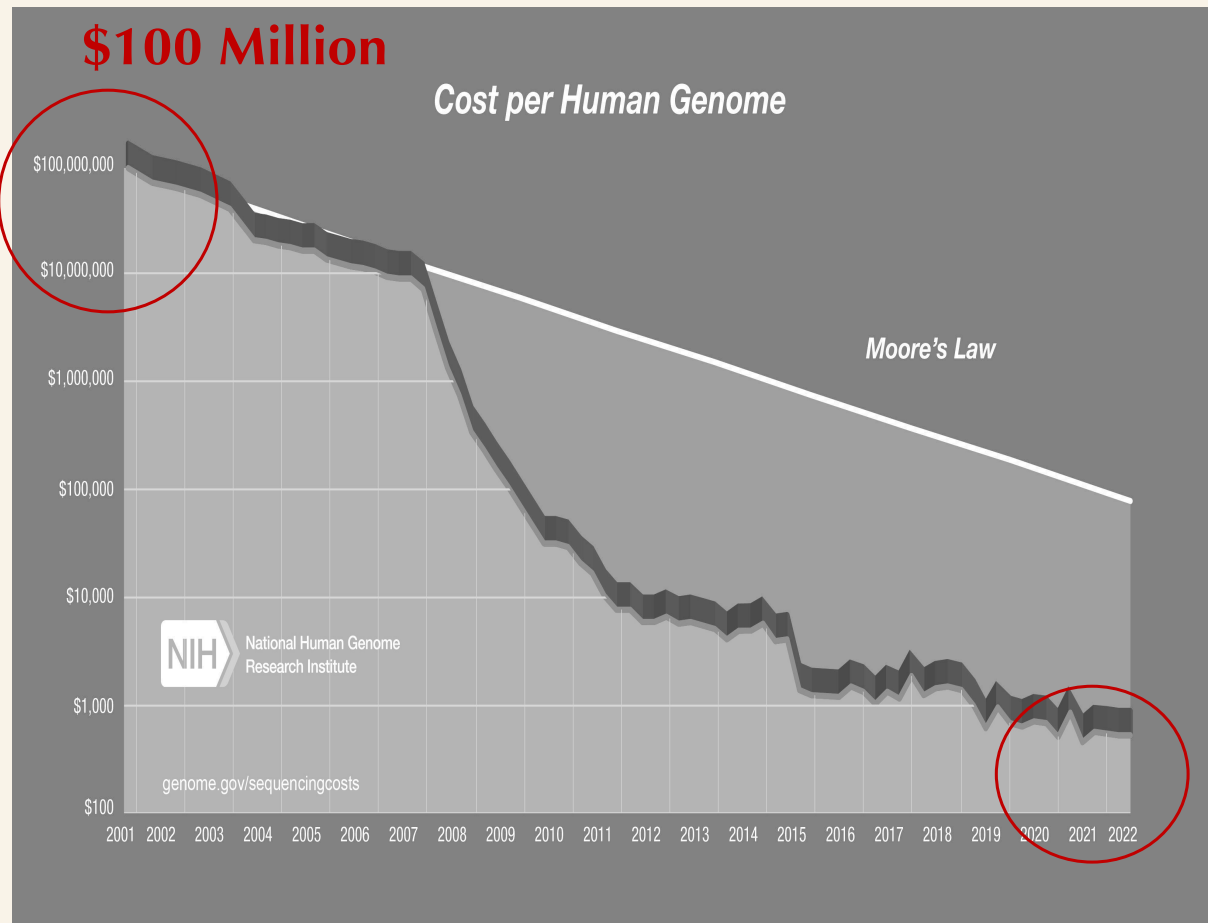
Why are we here?



Quick question!
What would this
Maserati at
~1 million NOK
in 2000 cost now
if similarly
devaluated?

\$1000

Why are we here? Phenomenal technical advances

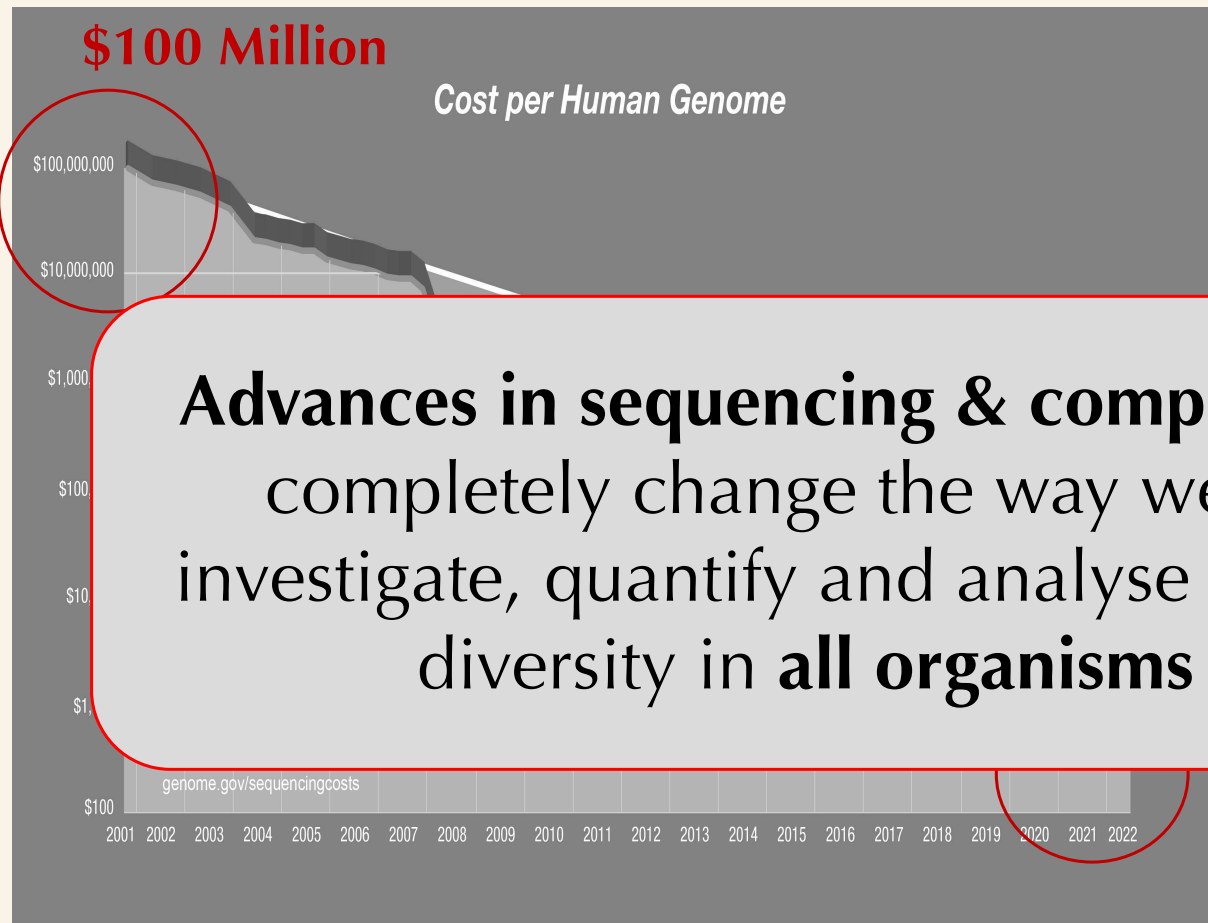


Quick question!
What would this
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10 NOK

\$1000

Has fundamentally changed the way we do biology

Why are we here? Phenomenal technical advance



Question!
this
at
OK
now
y
d?
K

\$1000

Has fundamentally changed the way we do biology

How has sequencing changed and is changing the world?

Changed healthcare

Sequencing (genome and exome) funded solely by *healthcare* systems

2012

~1%

2017

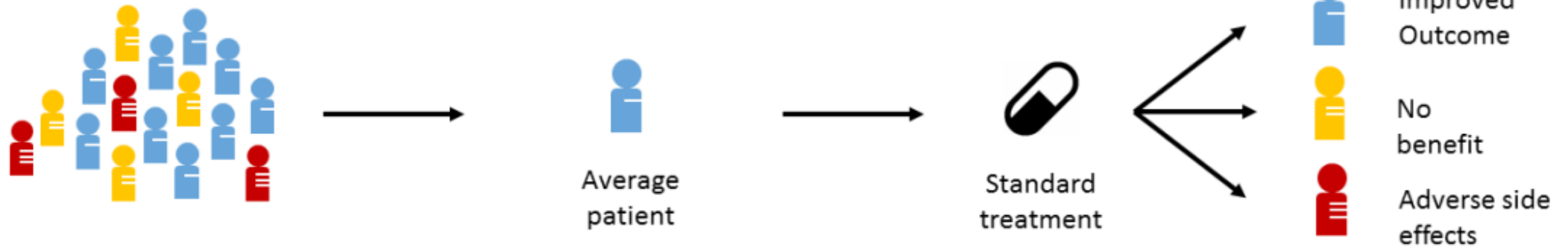
~20%

2022

>80%

Dag Undlien (OUH)

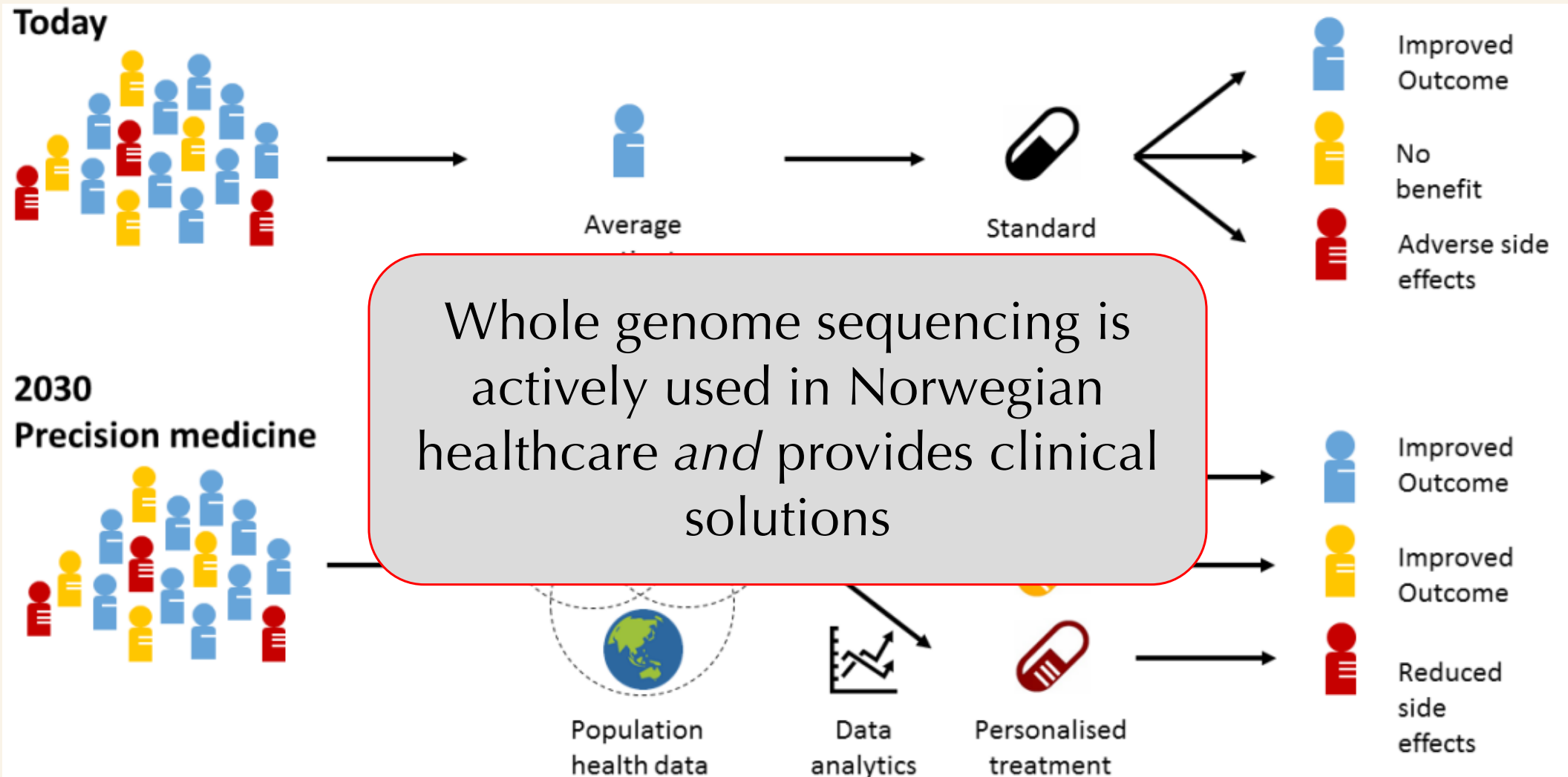
Today



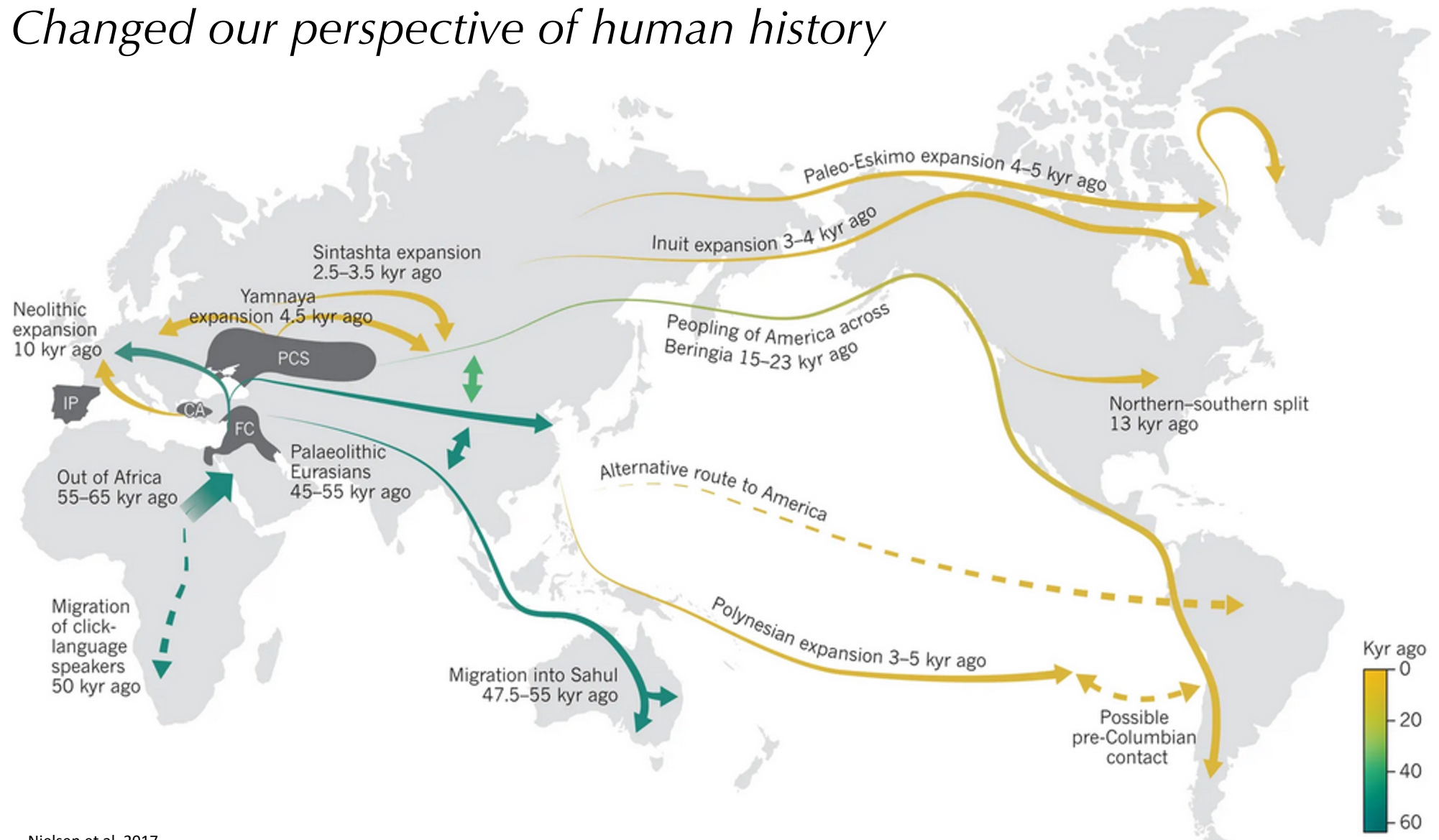
**2030
Precision medicine**



Dag Undlien (OUH)



Changed our perspective of human history



Changed forensic capabilities

Using continuously expanding public genomic databases (e.g. 23 and me)...

The New York Times

Genealogists Turn to Cousins' DNA and Family Trees to Crack Five More Cold Cases

Police arrested a D.J. in Pennsylvania and a nurse in Washington State this week, the latest examples of the use of an open-source ancestry site since the break in the Golden State killer case.

Changed forensic capabilities

Or by the genetic testing of thousands of people!

As the *Times* reports, that law paved the way for a prosecutor in the Verstappen case to call for the voluntary DNA sampling of 21,500 Dutchmen, and the obligatory sampling of 1,500 men who were of “special interest” to investigators.

The alleged killer, 55-year-old Jos Brech, was one of those 1,500 men who were mandated to provide a DNA sample. He never showed up. Dutch officials grew suspicious and took DNA samples from Brech's relatives. The results matched the DNA

Changed vaccine development and disease tracking

Genomic epidemiology of SARS-CoV-2 with subsampling focused globally over the past 6 months

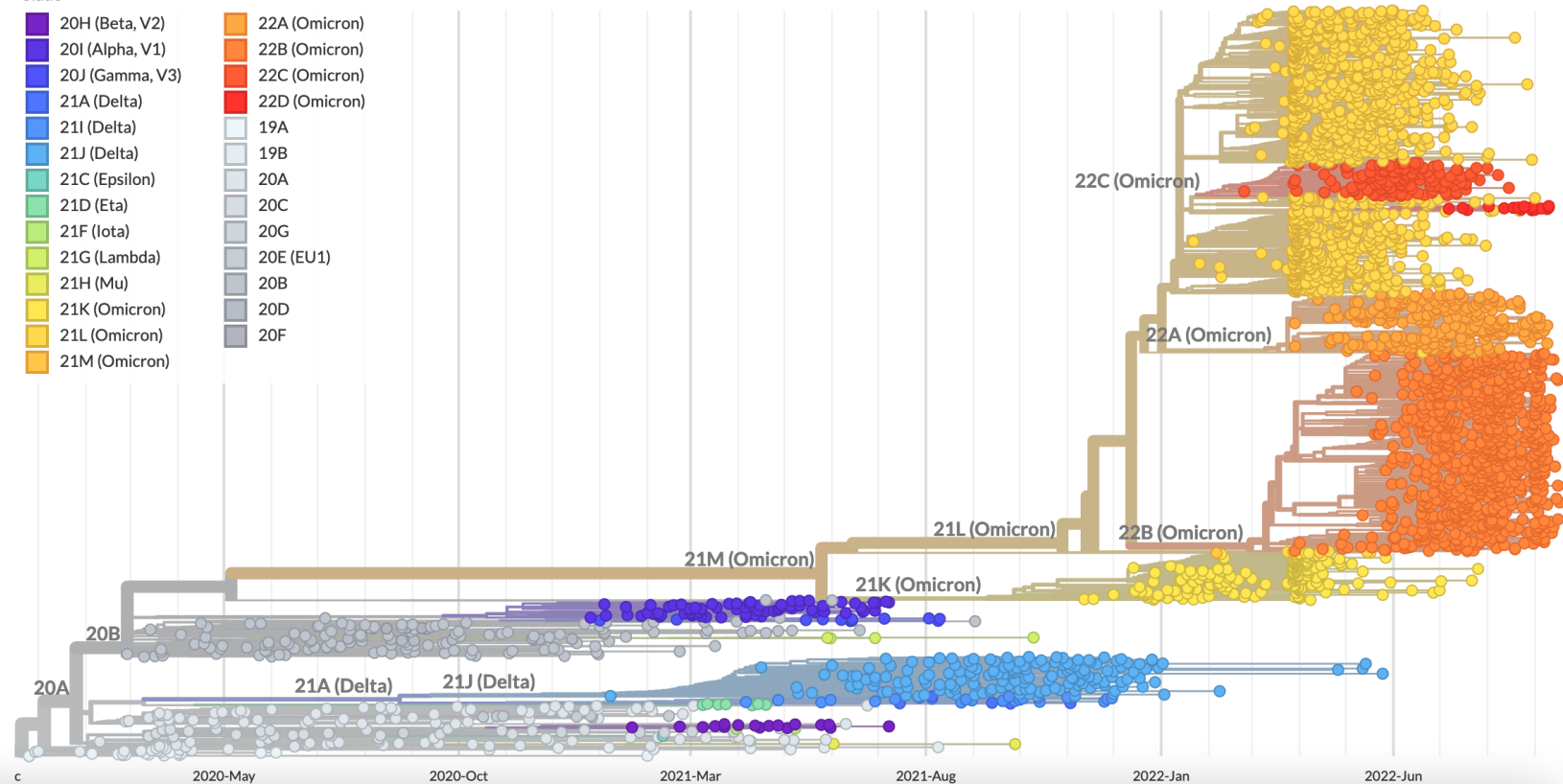
Built with [nextstrain/ncov](#). Maintained by [the Nextstrain team](#). Enabled by data from [GISAID](#).

Showing 3074 of 3074 genomes sampled between Dec 2019 and Sep 2022.

Phylogeny

Clade ^

- | | |
|-----------------|---------------|
| 20H (Beta, V2) | 22A (Omicron) |
| 20I (Alpha, V1) | 22B (Omicron) |
| 20J (Gamma, V3) | 22C (Omicron) |
| 21A (Delta) | 22D (Omicron) |
| 21I (Delta) | 19A |
| 21J (Delta) | 19B |
| 21C (Epsilon) | 20A |
| 21D (Eta) | 20C |
| 21F (Iota) | 20G |
| 21G (Lambda) | 20E (EU1) |
| 21H (Mu) | 20B |
| 21K (Omicron) | 20D |
| 21L (Omicron) | 20F |
| 21M (Omicron) | |



Changed improvement and selection of commercial crops

Vitamin D Deficiency



5 IN 10 PEOPLE

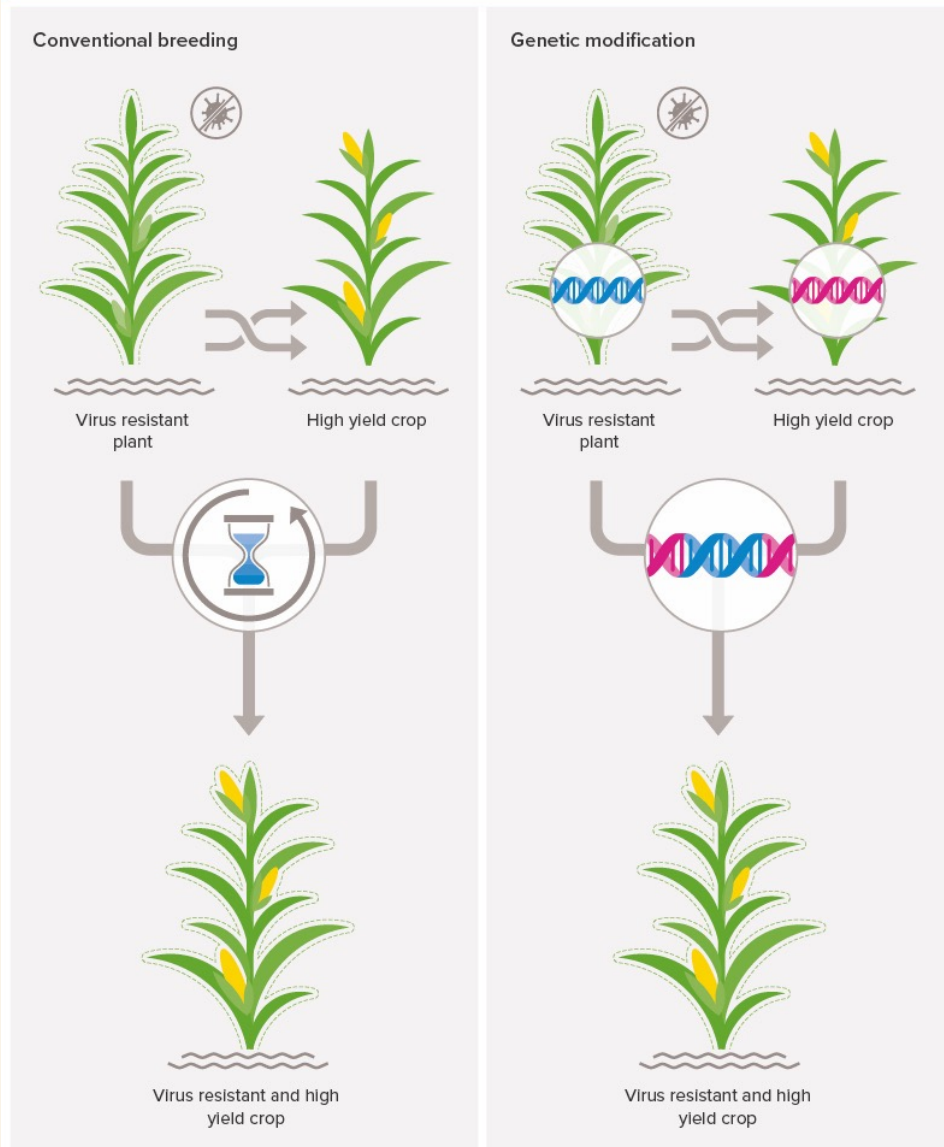
globally have a vitamin D insufficiency.

1 billion people worldwide are affected by low levels of vitamin D.

SeedWORLDGROUP



FIGURE 3 Differences between conventional breeding and GM



Changed our understanding of the human microbiome

NIH Human Microbiome Project



Characterization of the microbiomes of healthy human subjects at five major body sites, using 16S and metagenomic shotgun sequencing.

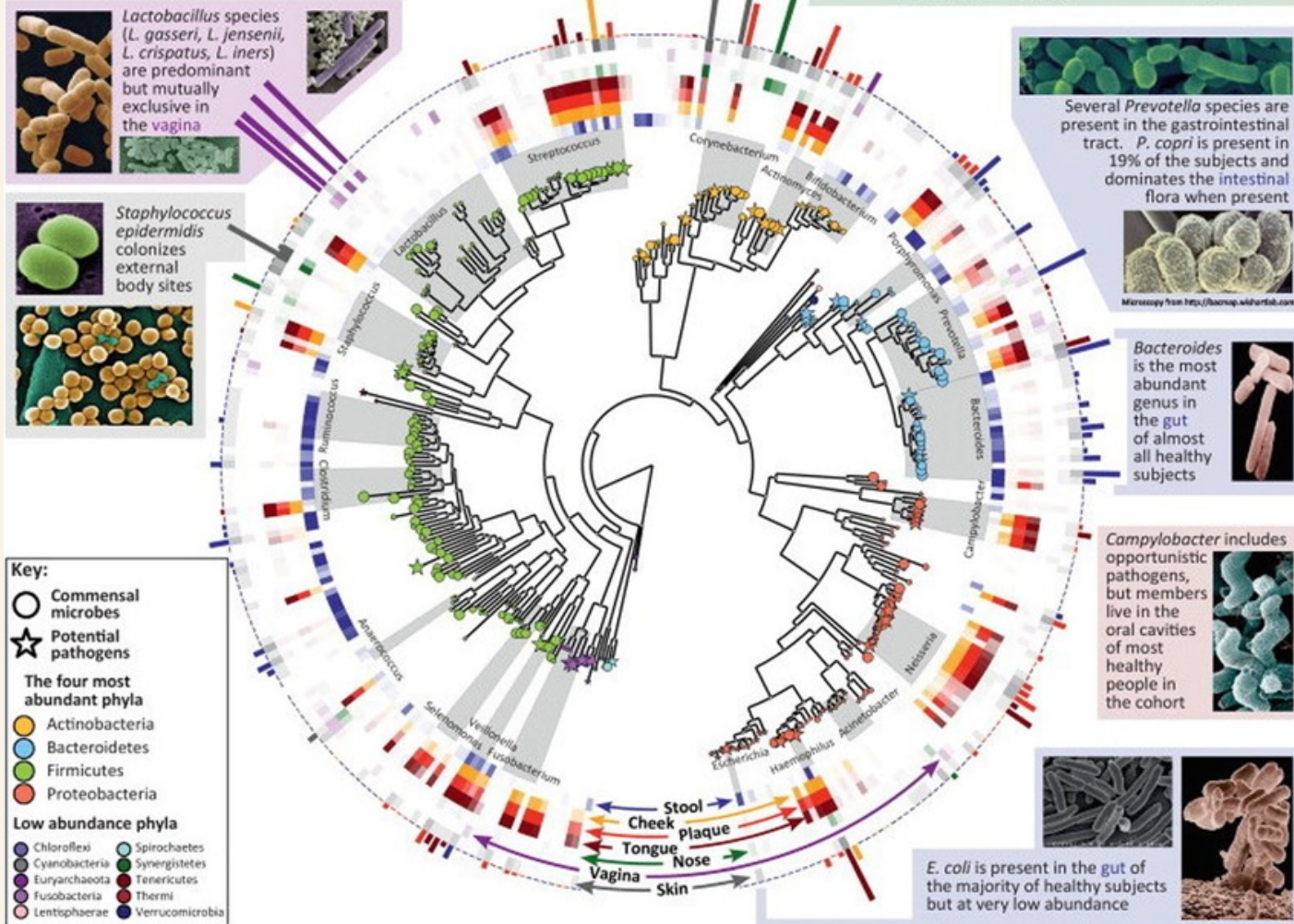
Enter HMP1



Characterization of microbiome and human host from three cohorts of microbiome-associated conditions, using multiple 'omics technologies.

Enter iHMP

A map of diversity in the human microbiome



Changing our perspective of extinct species

How Woolly mammoths could be brought back from extinction

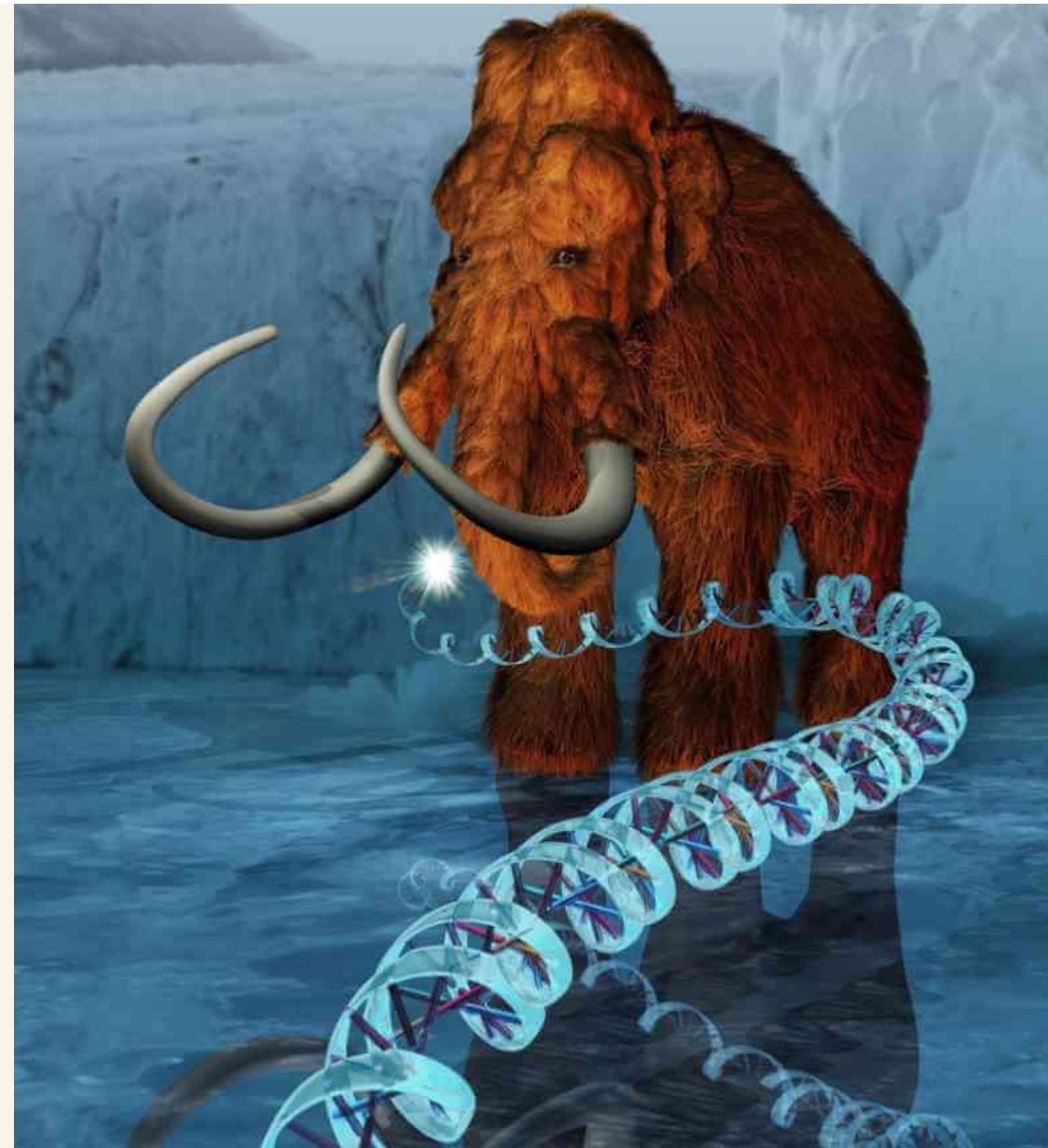
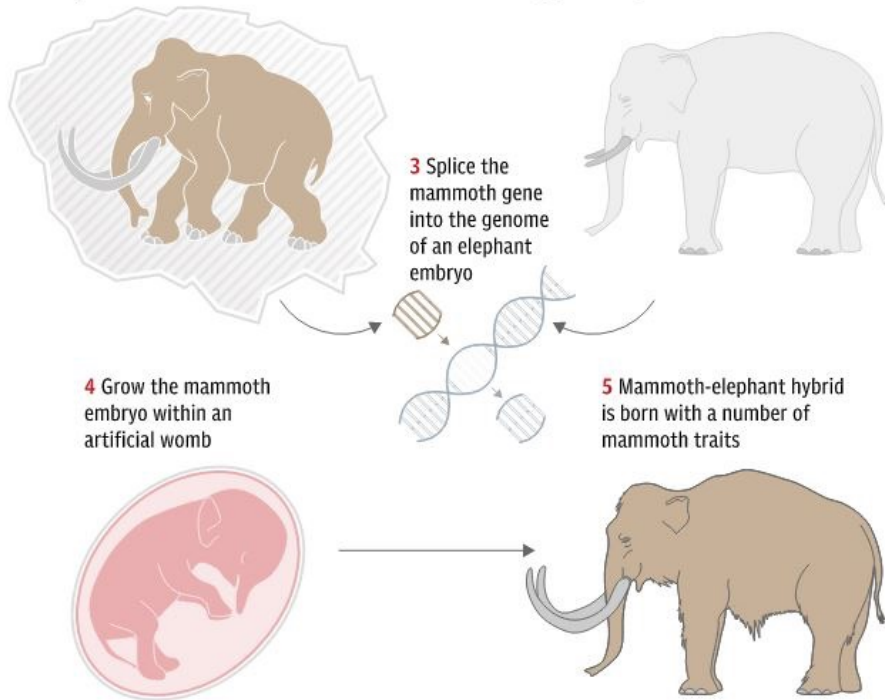
1 DNA extracted from mammoth found in permafrost.

2 Identify genes which separate them from elephants, such as those which code for a shaggy coat, big ears and antifreeze blood.

3 Splice the mammoth gene into the genome of an elephant embryo

4 Grow the mammoth embryo within an artificial womb

5 Mammoth-elephant hybrid is born with a number of mammoth traits



Changing our perspective of extinct species

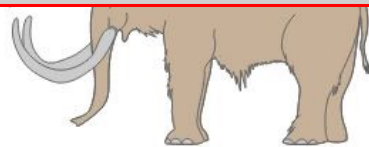
How Woolly mammoths could be brought back from extinction

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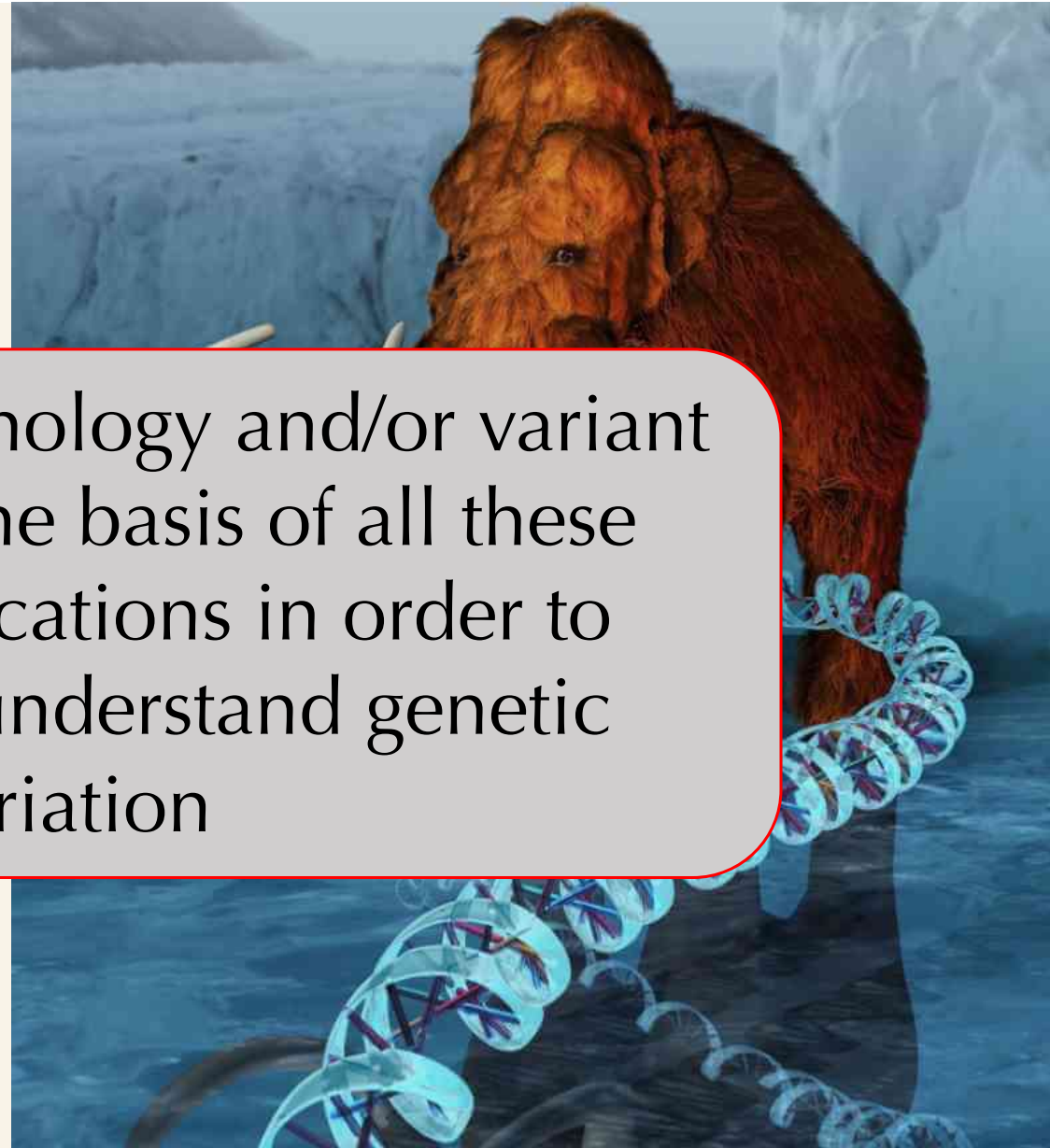
2 Identify genes which separate them from elephants, such as those which code for a shaggy coat, big ears and antifreeze blood.



4 Grow the mammoth embryo within an artificial womb



Sequencing technology and/or variant calling are at the basis of all these different applications in order to quantify and understand genetic variation



Available to
high school
students!
21.10 2022

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High school student is first to sequence the angelfish genome

17-year-old Indeever Madireddy sequenced the genome of his pet angelfish after it died – the first time this species has been sequenced

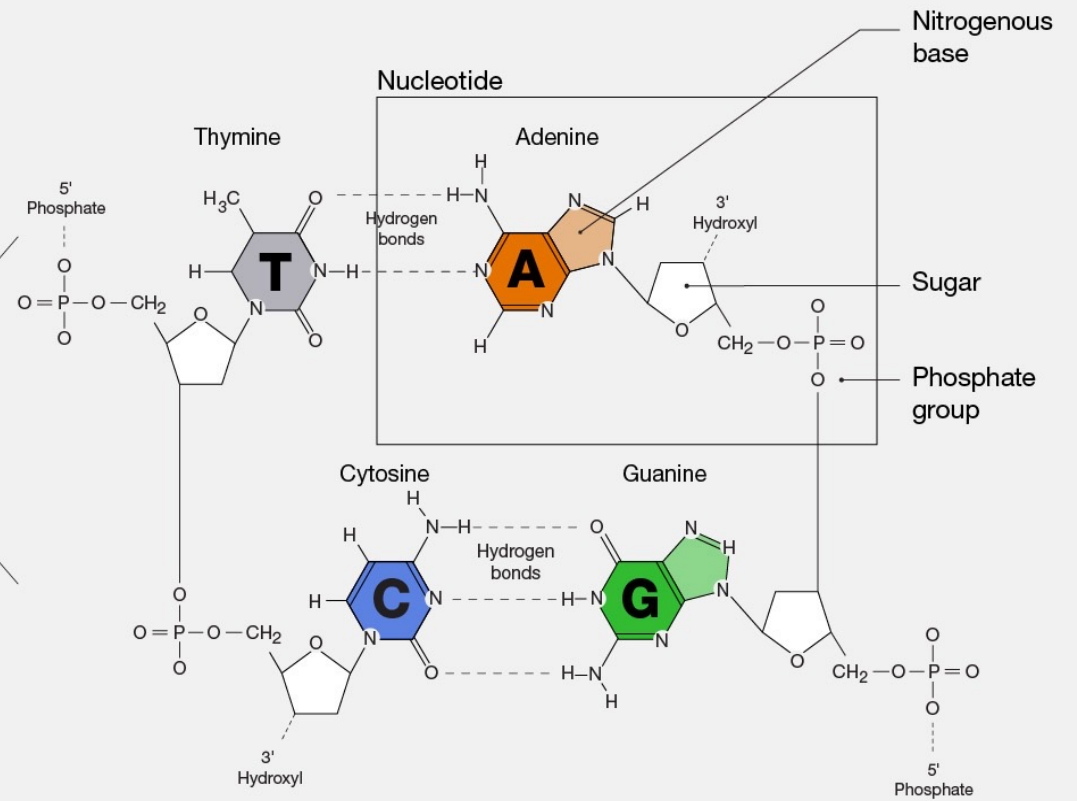
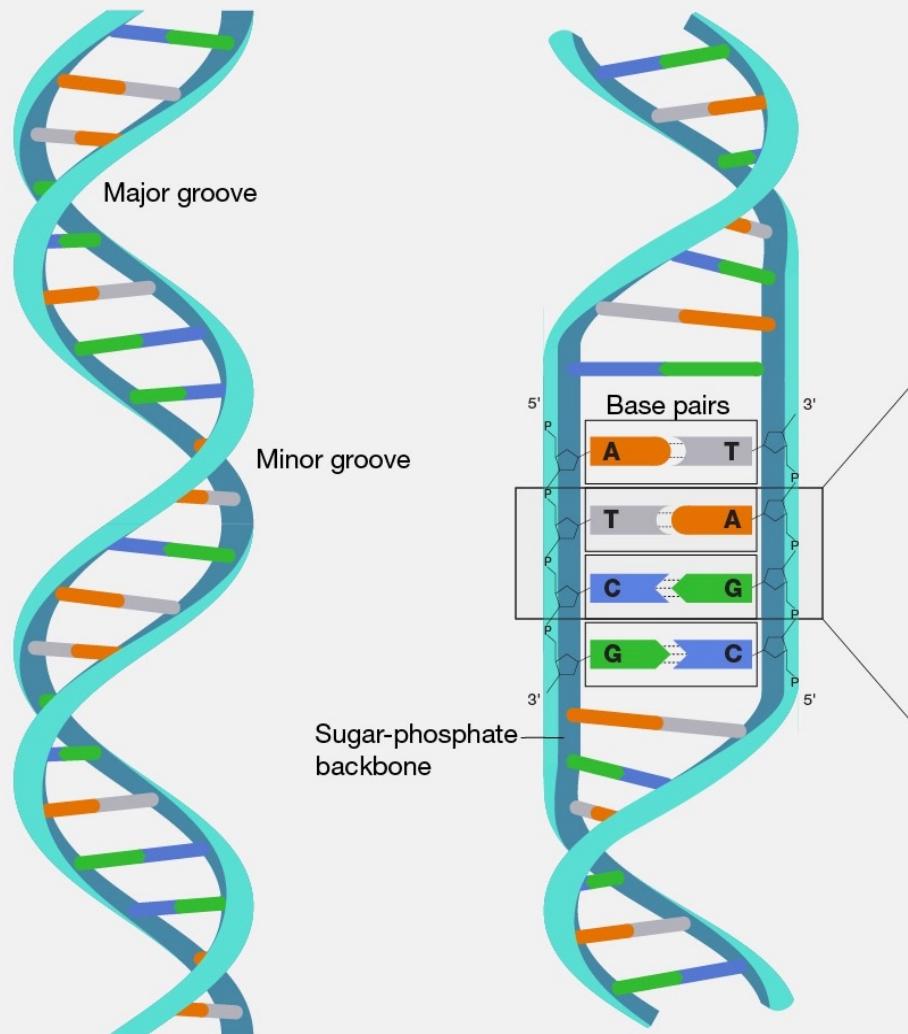


LIFE 21 October 2022

By [Michael Le Page](#)



DNA



What does genetic variation look like?

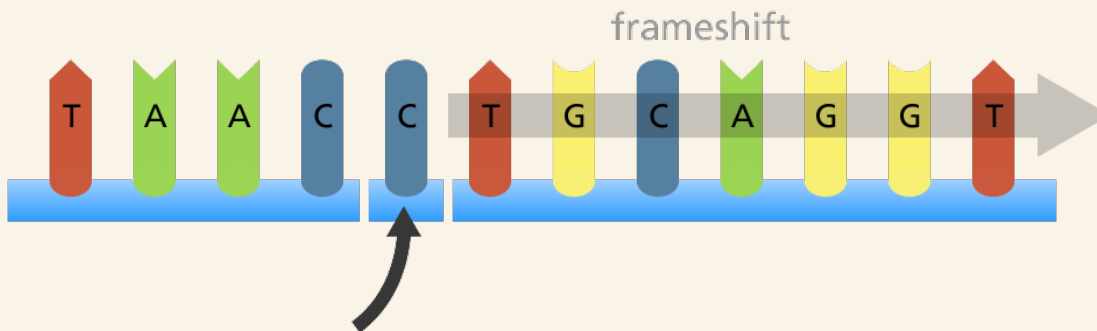
1) DNA (nucleotides) can be inserted or deleted (*indels*).

1) Insertion/Deletion (Indel)

Original sequence



Insertion



Can range from 1 base-pair (bp) to many bp

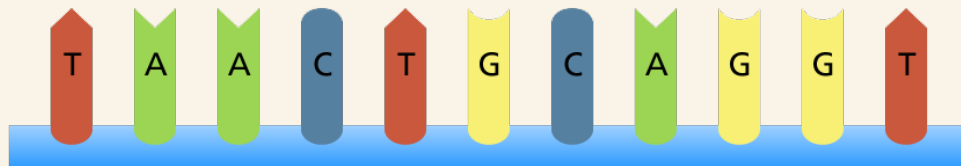
What does genetic variation look like?

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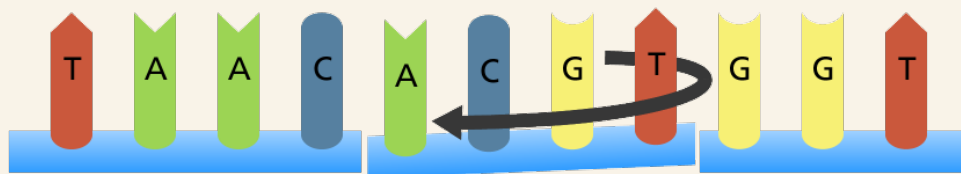
2) DNA can be *structurally rearranged*
(inversions/translocations)

2) Structural rearrangements (inversions/translocations)

Original sequence



Inversion



Can be MILLIONS of
bp long affecting the
order of many genes
simultaneously
(*Supergenes*)

What does genetic variation look like?

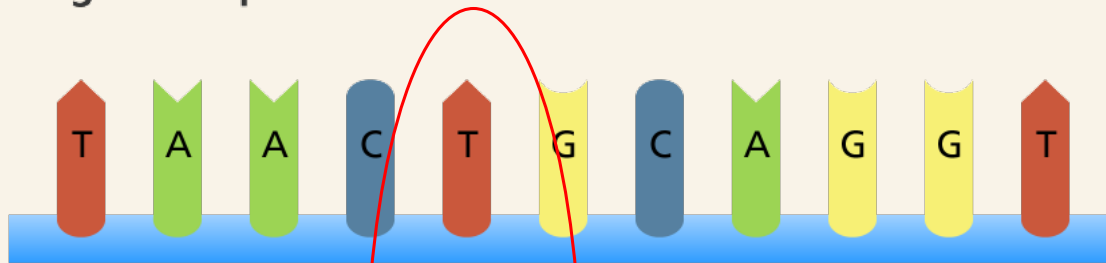
1) DNA (nucleotides) can be inserted or deleted (*indels*).

2) DNA can be *structurally rearranged* (inversions/translocations)

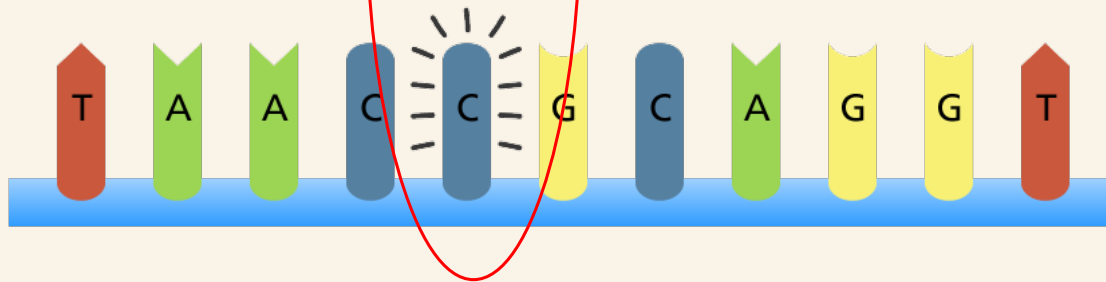
3) DNA can be *altered* at a single base pair (Single Nucleotide Polymorphism or SNP)

3) Single Nucleotide Polymorphism (SNP)

Original sequence



Point mutation



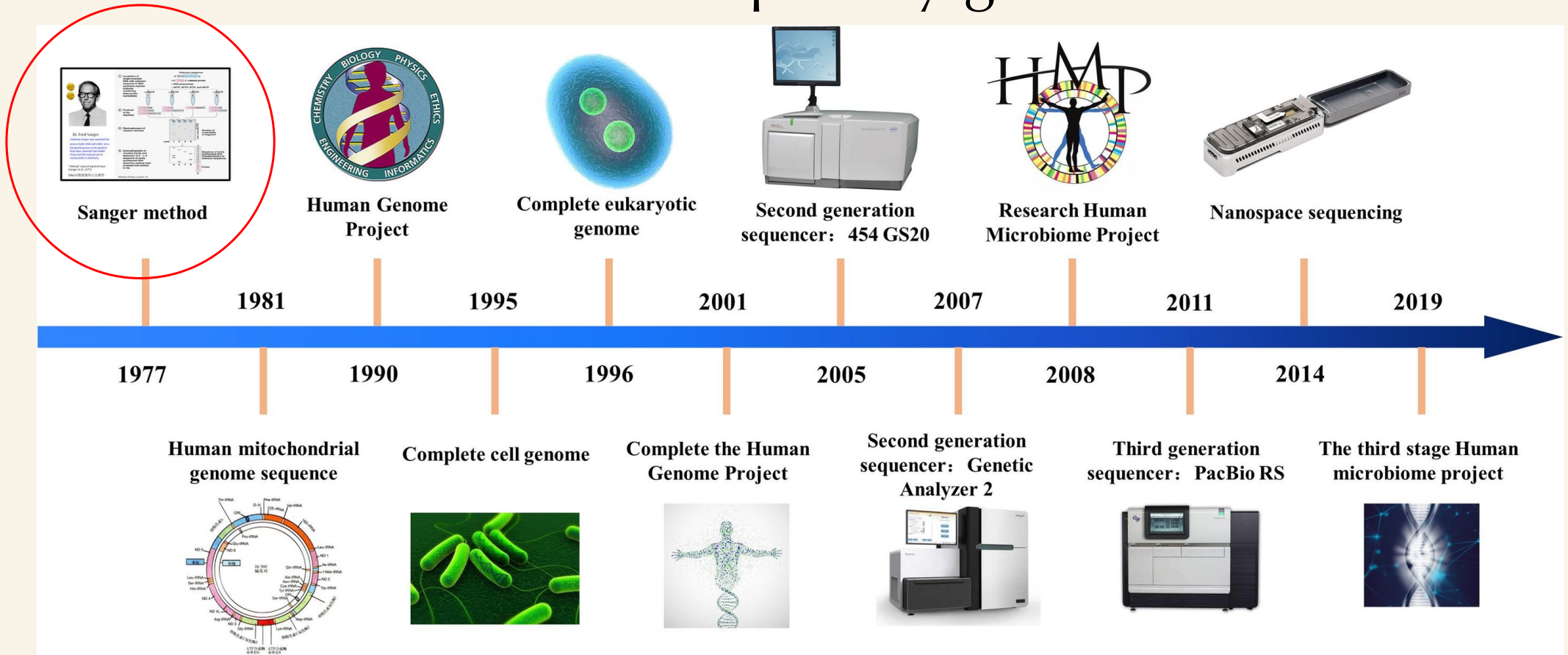
Extremely common

> 150 million known SNPs
in humans (2015)

~100 SNPs unique in
EVERY human

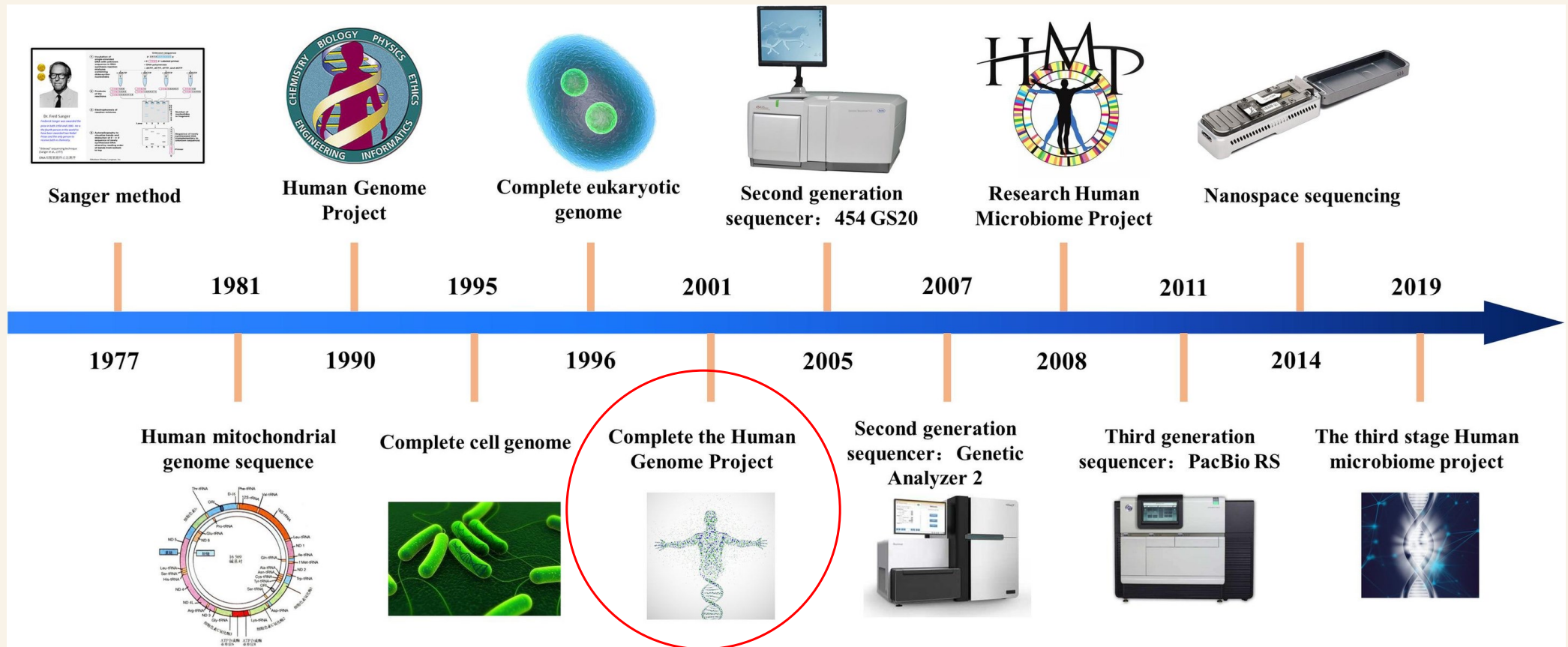
*Putatively EVERY base in
the human genome*

How do we observe and quantify genetic variation?



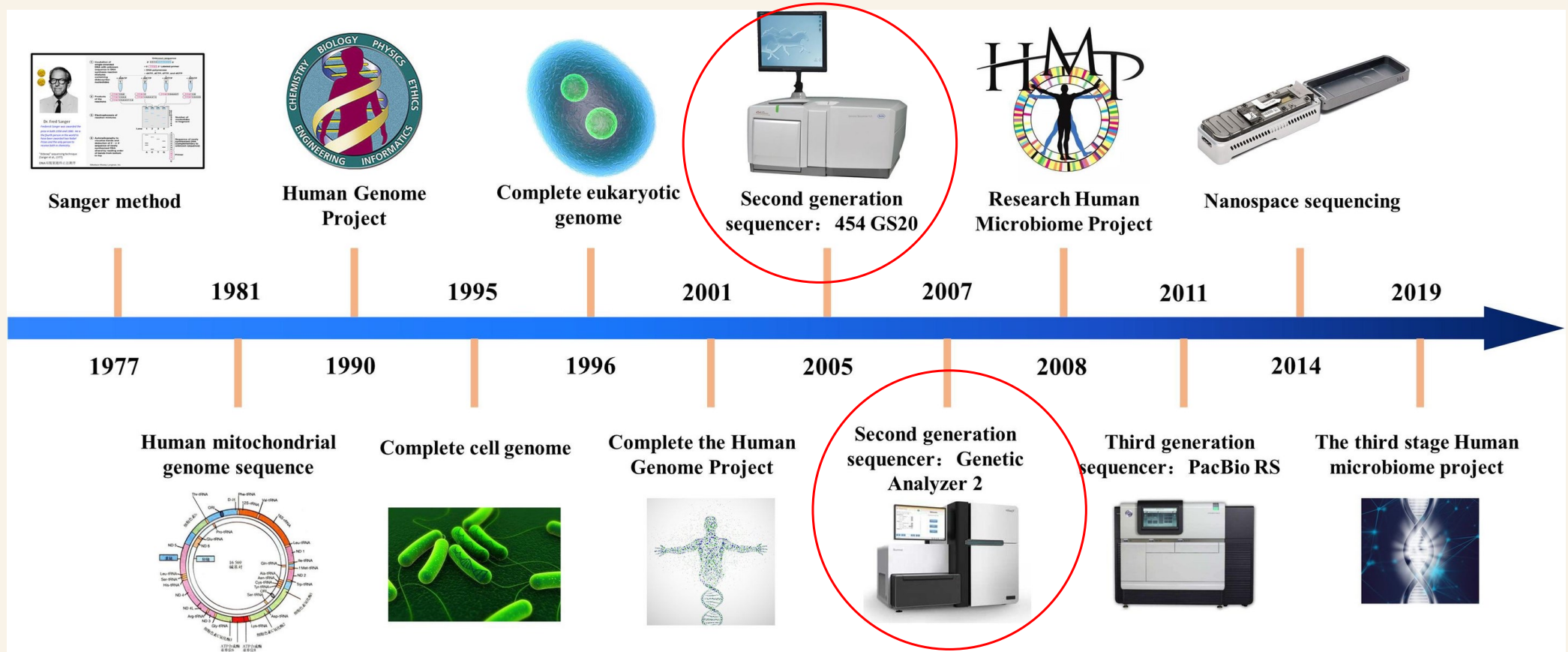
Sanger sequencing – leading sequencing technology for decades

How do we observe and quantify genetic variation?



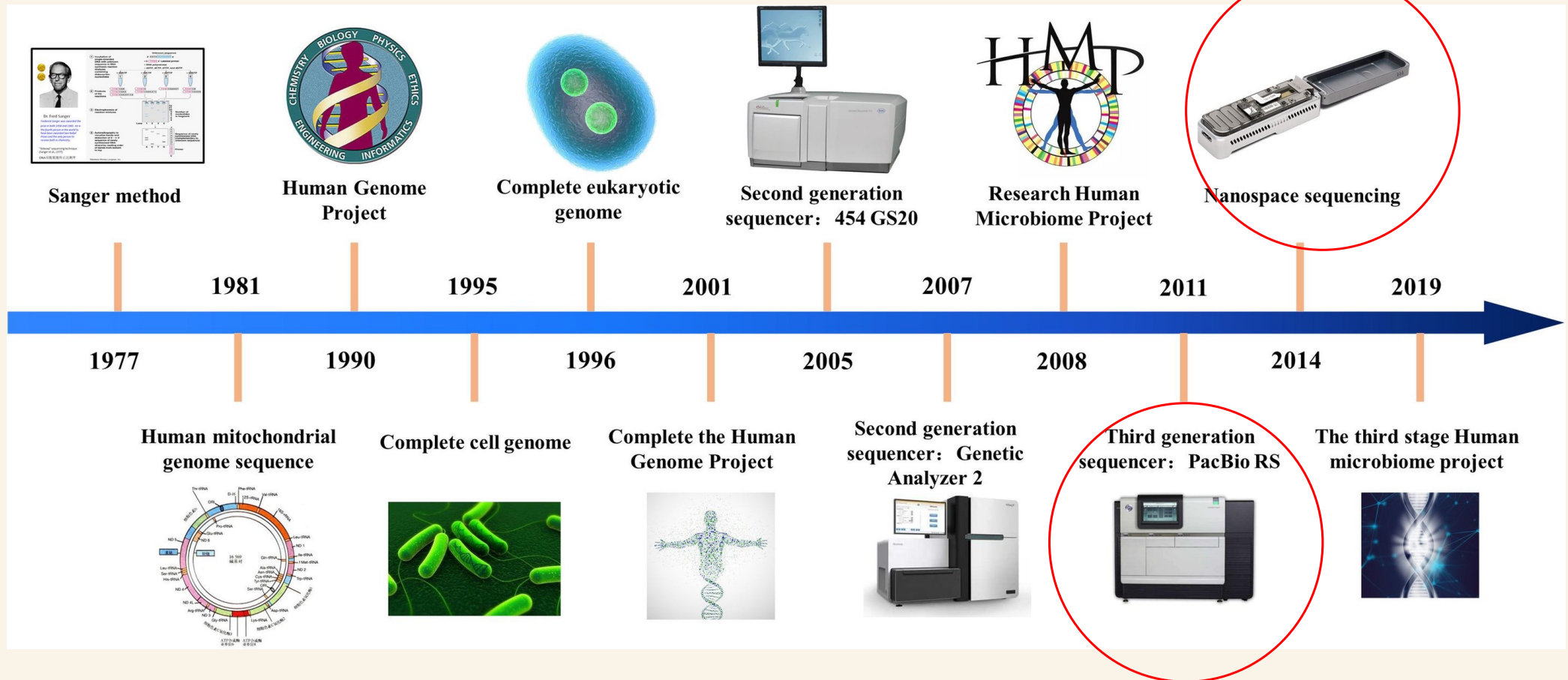
Human genome project: sparked a novel industry

How do we observe and quantify genetic variation?



“New” sequencing technologies (already outdated!)

How do we observe and quantify genetic variation?



Latest sequencing technologies that focus on long read sequencing

Two dominant technologies today



PacBio

Long read length (10k bp +)

More expensive

Specific applications

Two dominant technologies today



PacBio

Long read length (10k bp +)

More expensive

Specific applications



Illumina

Short read length (150-250 bp)

Cheap!

Workhorse of sequencing

Practical considerations: size matters!



PacBio

Long read length (10k bp +)

More expensive

Specific applications



Illumina

Short read length (150-250 bp)

Cheap!

Workhorse of sequencing

What variation can you assess with these different types of reads?

Type of variant	Short reads	Long reads
Indel	Only if small (~few bp)	Yes
Structural (inversion)	Difficult	Yes
SNP	Yes	Yes

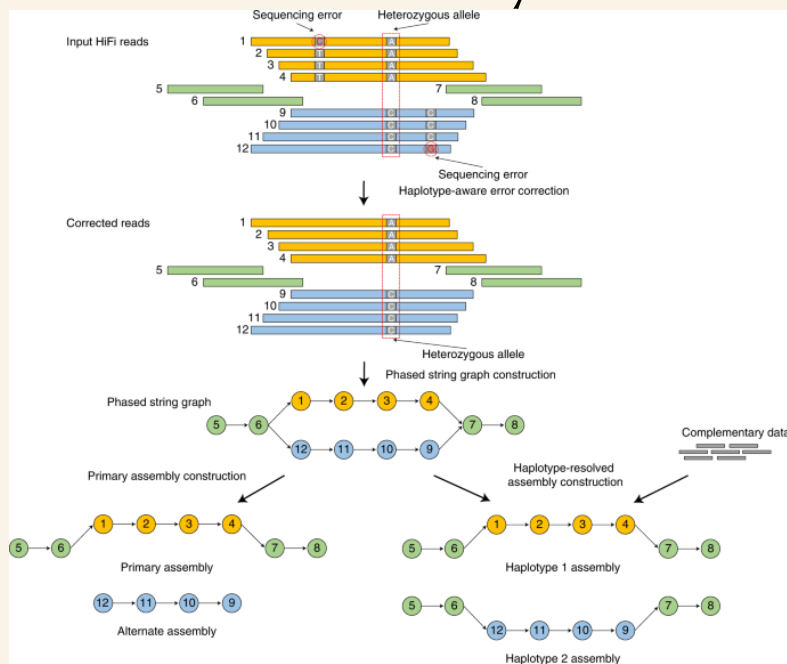
What variation can you assess with these different types of reads?

Type of variant	Short reads	Long reads
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Illumina ***re-sequencing*** domination means that SNPs are most reliably targeted and are most studied type of genetic variation

Yet there *are* different ways of assessing genetic variation

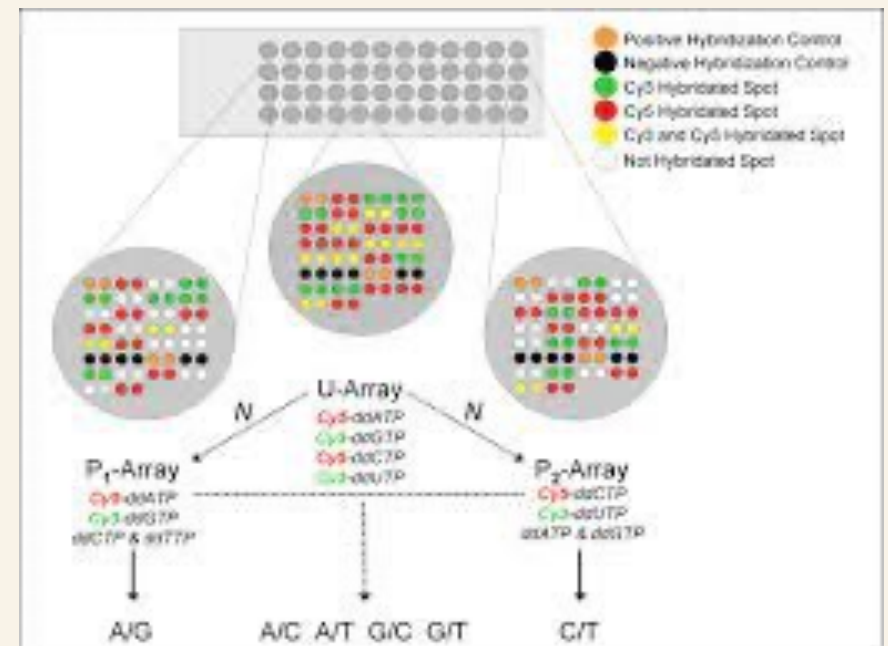
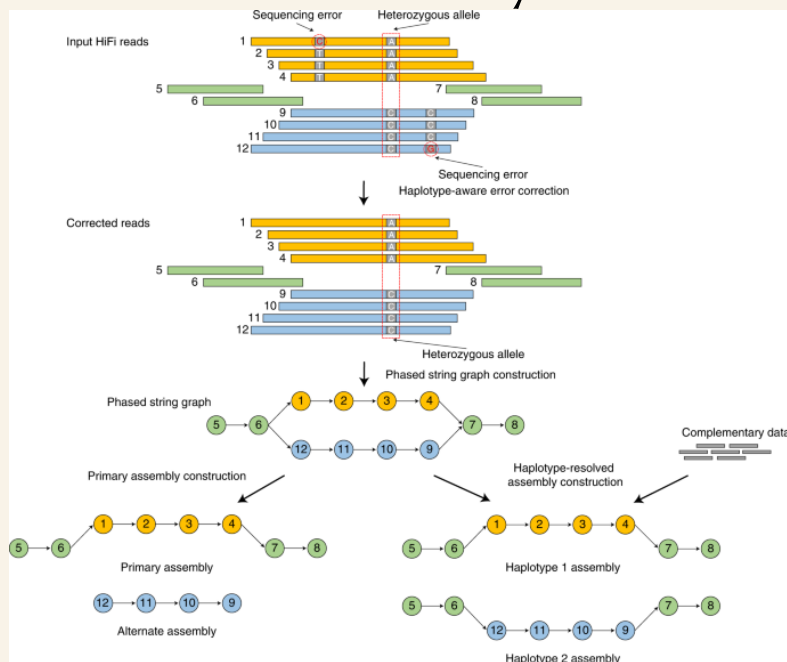
i.e. *de novo*
haplotype aware
assembly



Yet there *are* different ways of assessing genetic variation

i.e. *de novo*
haplotype aware
assembly

SNP Chip/DNA micro
array genotyping

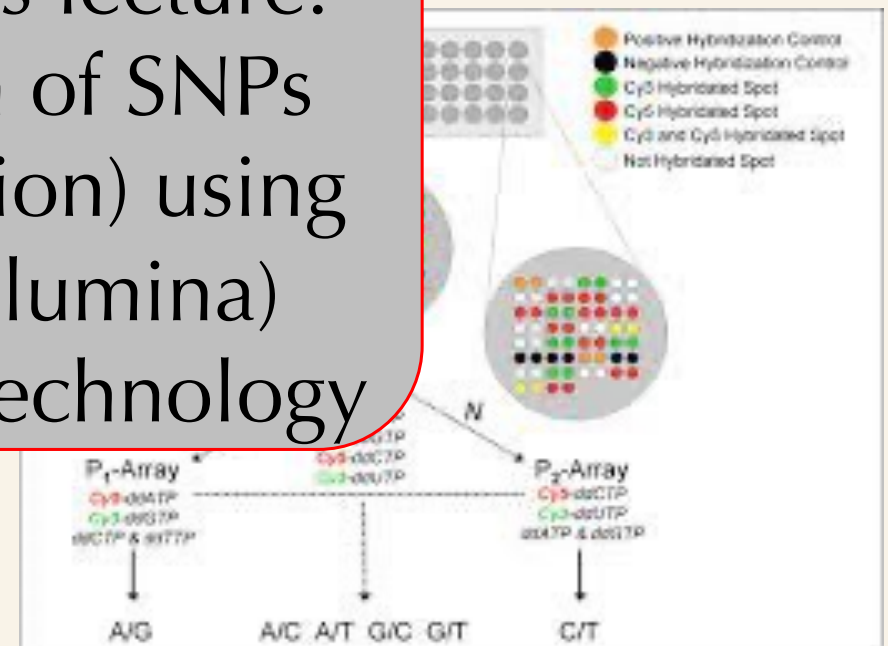
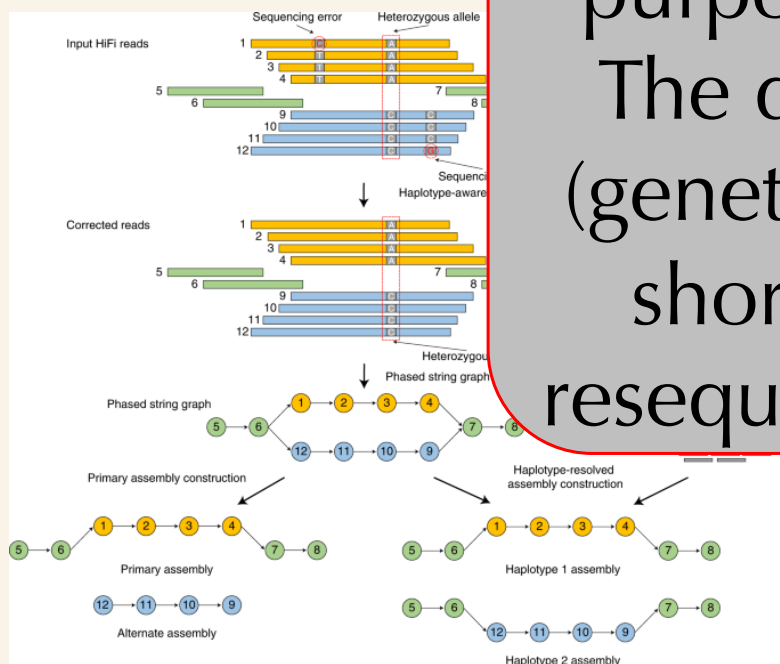


Yet there *are* different ways of assessing genetic variation

i.e. *de novo*
haplotype assembly

SNP Chip
typing

Variant calling for
purpose of this lecture:
The detection of SNPs
(genetic variation) using
short read (Illumina)
resequencing technology



Questions?



2) Variant calling pipelines/methods and limitations

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Variant calling **always** starts with a reference genome



Assembly of the first complex vertebrate genome
Human genome assembly project (2003)
Not easily repeated: it was massive task
Nowadays; much cheaper and faster

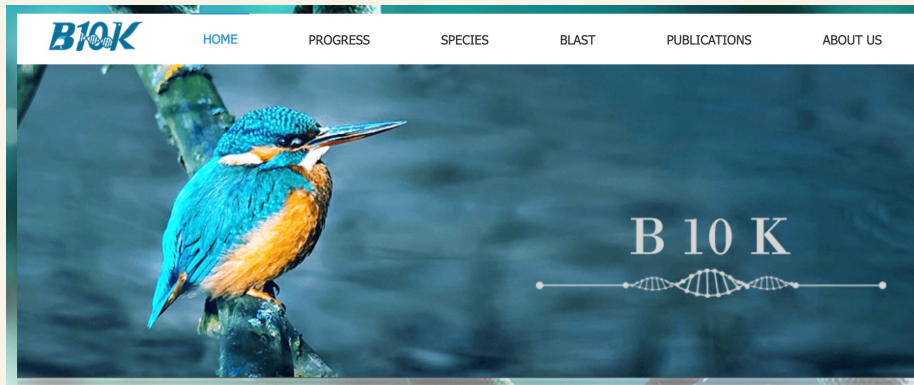
2) Variant calling pipelines/methods and limitations

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Great push to provide reference genomes for many organisms!



B10 K: 10.000 bird genomes

*Deep evolutionary understanding
of the entire living avian class*

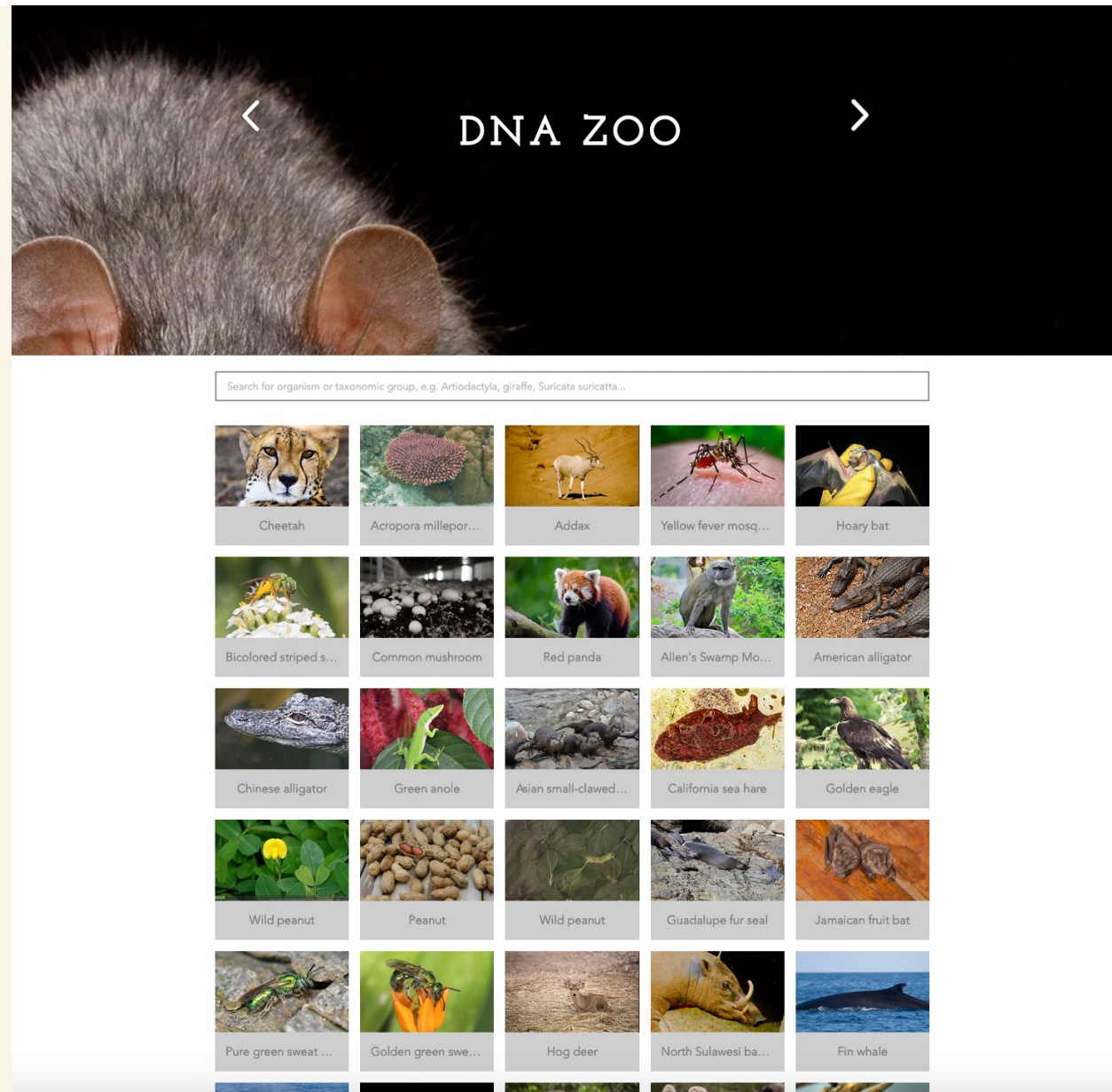
<https://b10k.genomics.cn/>



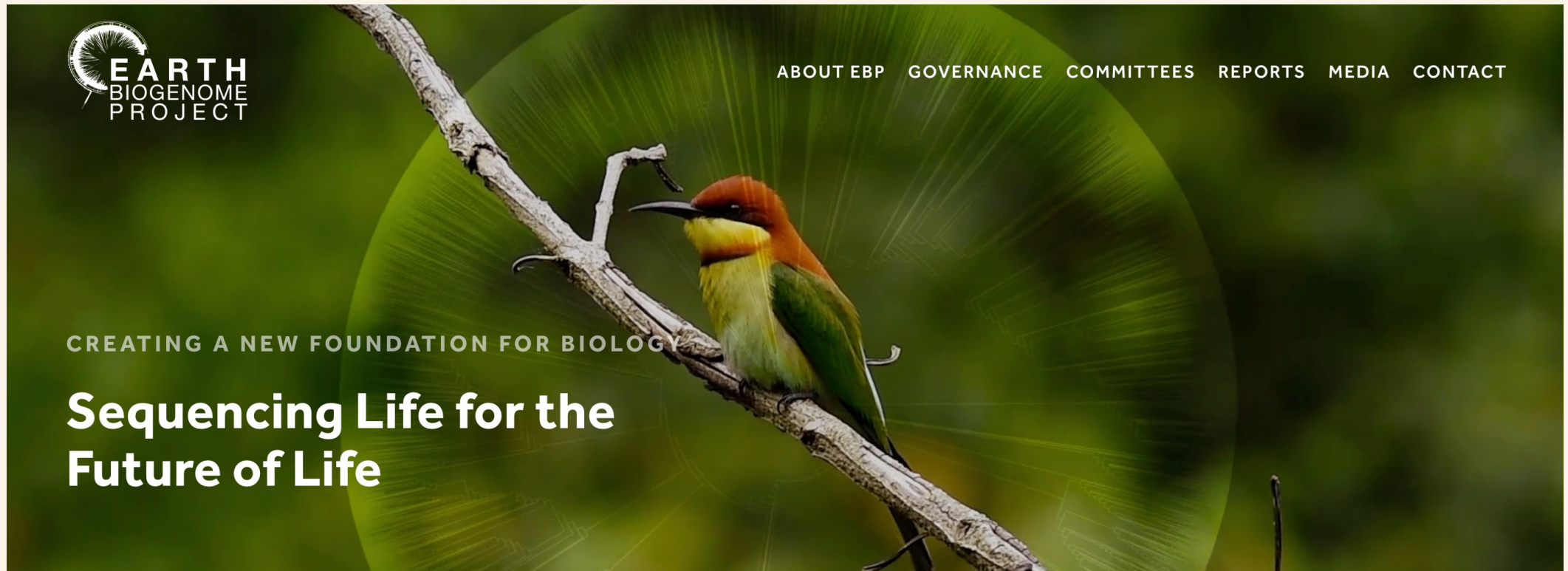
The *DNA Zoo*

facilitates conservation efforts by releasing high-quality genomics resources.

<https://www.dnazoo.org>



The most ambitious: Earth Biogenome Project



<https://www.earthbiogenome.org/>

The most ambitious: Earth Biogenome Project



ABOUT EBP GOVERNANCE COMMITTEES REPORTS MEDIA CONTACT

EBP: moonshot for biology, aims to characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.

<https://www.earthbiogenome.org/>

The most ambitious: Earth Biogenome Project



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EBP: moonshot for biology, aims to characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.

The vision: to create a new foundation for biology, with new solutions for preserving biodiversity and sustaining human societies.

<https://www.earthbiogenome.org/>

But what
is a
reference
genome?

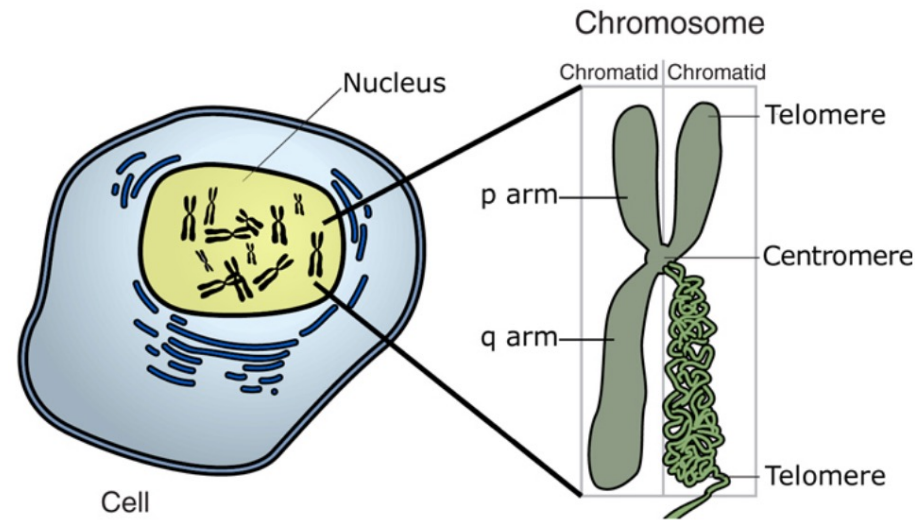
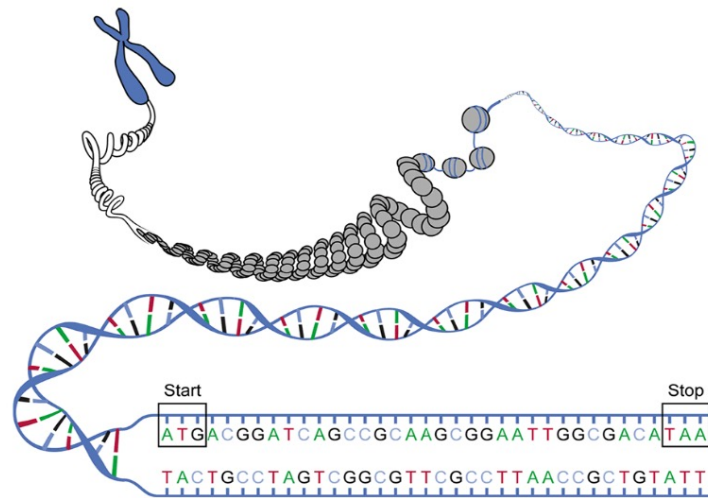
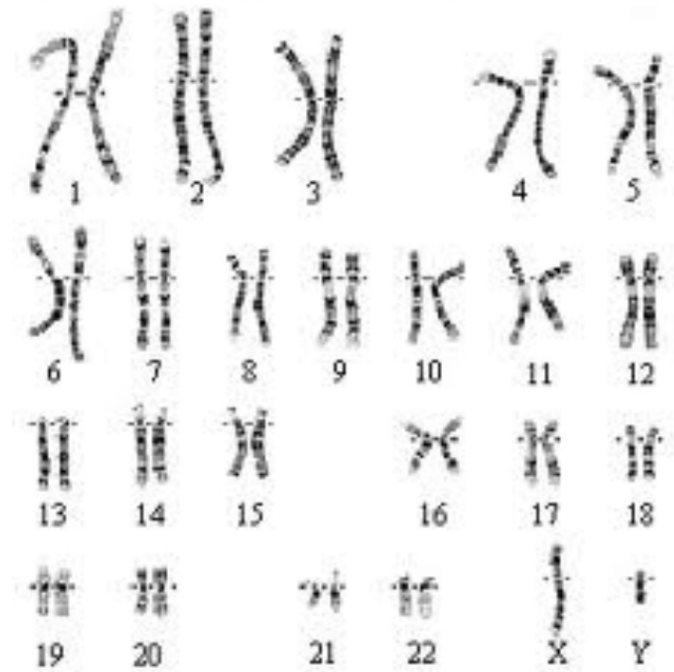


Image adapted from: National Human Genome Research Institute.



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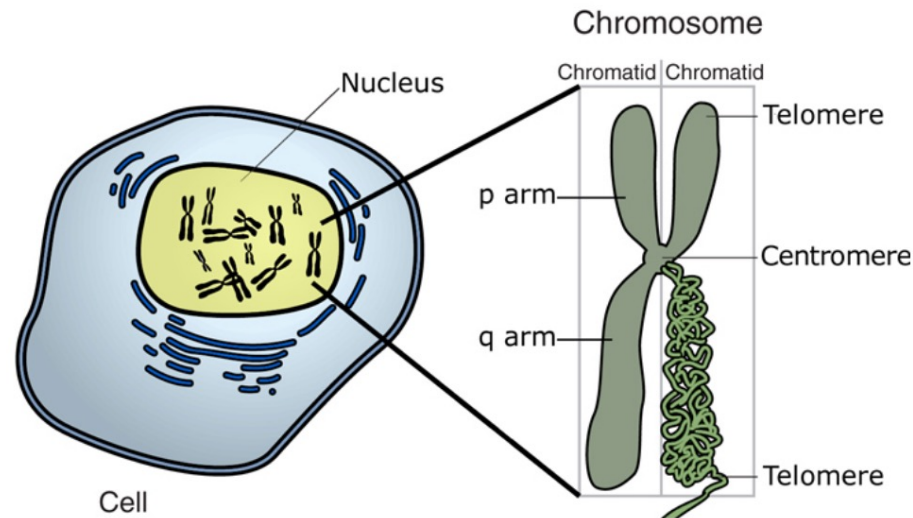
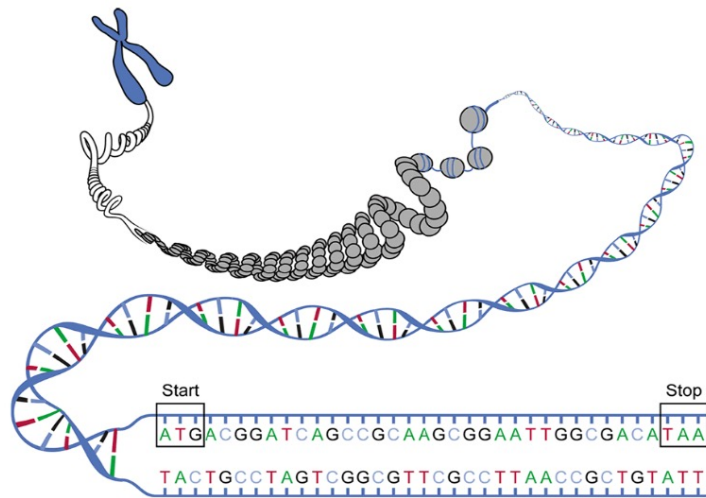
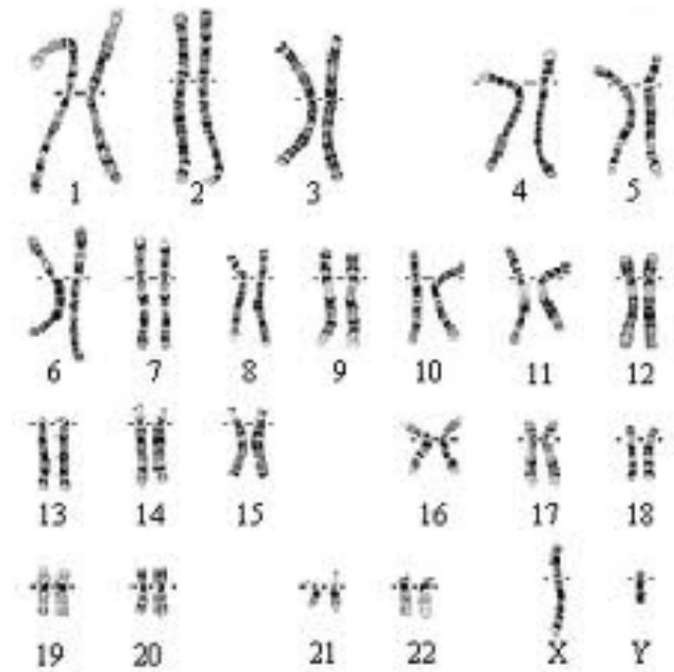


Image adapted from: National Human Genome Research Institute.



Digital representation /
abstraction of a physical,
biological phenomenon

A reference genome is ...

Usually from a single individual

Result of a genome assembly process -> errors are introduced

Of varying quality, that can vary from organism to organism

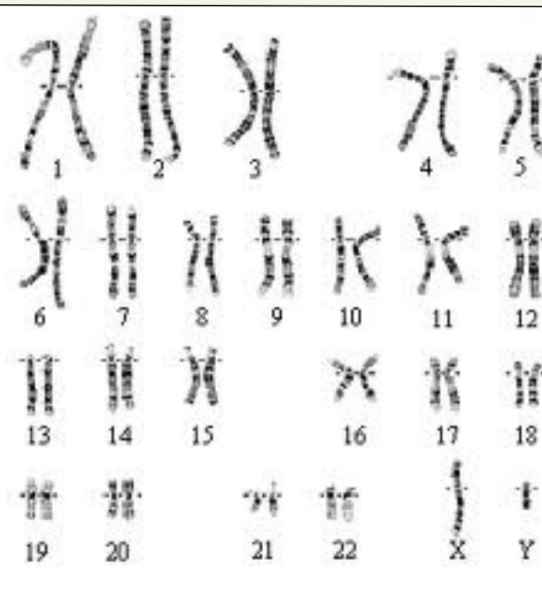
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Digital version of the genome



Sequencing
and
assembly

>Chr01
ACTACGTATATAGCATGATCATGCATGATACATGGCTAGT...
>Chr02
ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...
>Chr03
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...
>Chr04
CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...
... ..

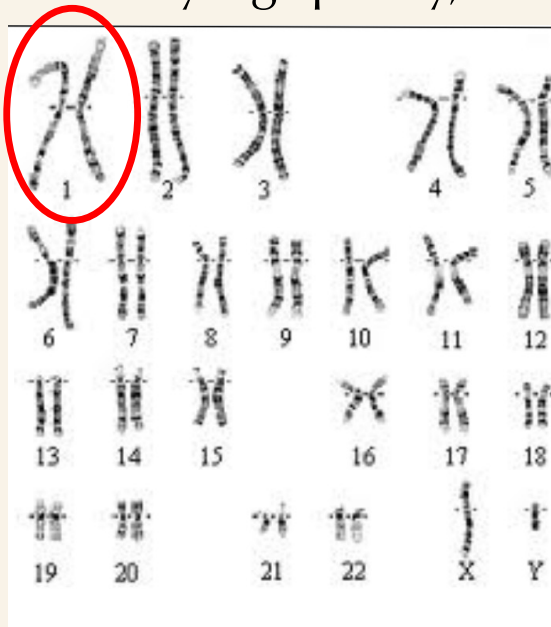
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>Chr02

ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...

>Chr03

ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...

>Chr04

CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...

... ..

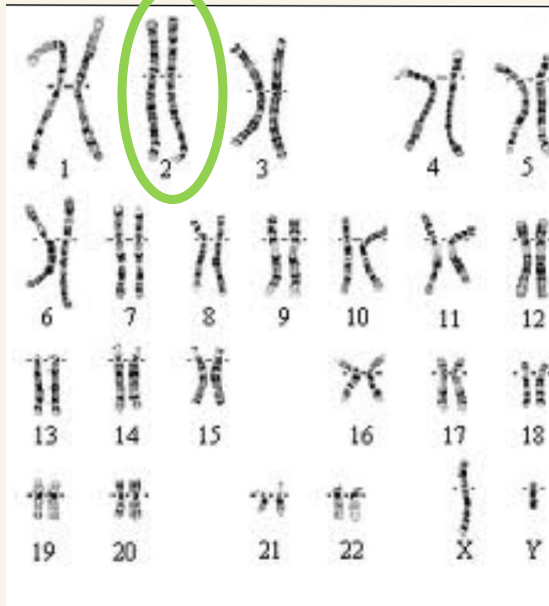
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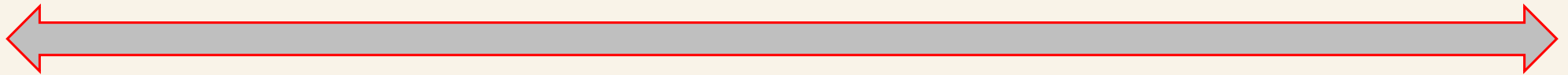
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>Chr02
ATCATGCATGATACATGGCTAGTACTACGTATATAGCATG...
>Chr03
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...
>Chr04
CGTATATAGCATGATCATGACTACATGATACATGGCTAGT...
... ..

Quality scale of reference genomes

Poor

Good

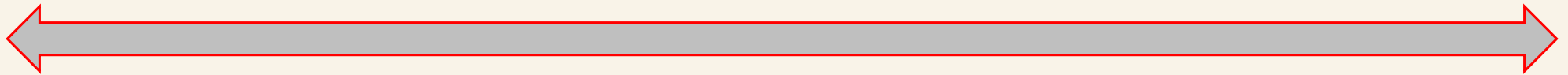


Chromosomes unclear
Thousands of loose fragments
Gaps (*nnnnn*) in sequences
Missing nucleotides

Quality scale of reference genomes

Poor

Good



Chromosomes unclear
Thousands of loose fragments
Gaps (*nnnnn*) in sequences
Missing nucleotides

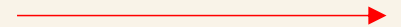
Chromosomes resolved
Continuous sequences
No gaps
Most nucleotides covered,
including centromeres and
repetitive regions

A reference genome has a 2D coordinate system

>Chr01

ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...

123456789.....



Millions of nucleotides/bases

Note: some different coordinate systems exist (i.e. starting at 0 or 1)
or using the base or space as “location”

A reference genome has a 2D coordinate system

```
>Chr01
```

```
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
```

```
123456789.....
```

Note: some different coordinate systems exist (i.e. starting at 0 or 1)
or using the base or space as “location”

i.e. A-C-T-A-C-G-T-A

```
 1 2 3 4 5 6 7 8  
 1 2 3 4 5 6 7
```

Such different systems are usually automatically recognized by different software

A reference genome has a 2D coordinate system

>Chr01
ACTACG
123456

TAGT...

This is all great, but where does the
variant calling come in?

Note: so
or using

i.e. A-C-G-T-A
1 2 3 4 5 6 7 8
1 2 3 4 5 6 7

Such different systems are usually automatically recognized by different software

A multiple alignment towards a reference

Start (12) Stop (31)

>Chr01
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...

Read 1 AGCATGATCATGCATGATGA

Read 2 GCATGATGATCATGCATGATACATGG

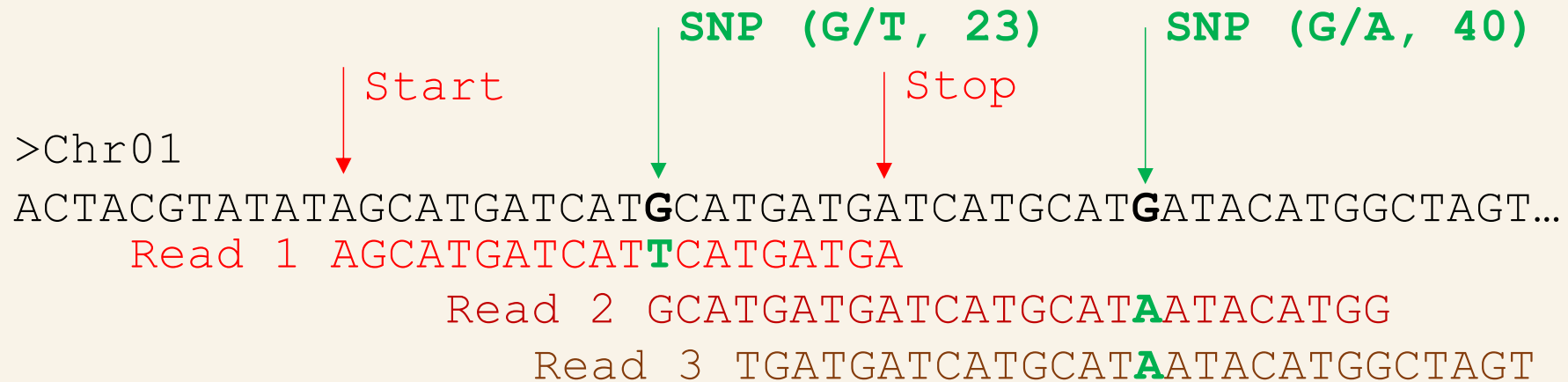
Read 3 TGATGATCATGCATGATACATGGCTAGT

The diagram illustrates the alignment of three short reads to a reference sequence. The reference sequence is shown on the first line, starting with a header '>Chr01'. The second line shows the reference sequence 'ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...'. The third line shows 'Read 1' aligned to the reference, starting at position 12 and ending at position 31. The fourth line shows 'Read 2' aligned to the reference, starting at position 13 and ending at position 30. The fifth line shows 'Read 3' aligned to the reference, starting at position 14 and ending at position 31. Red arrows point from the 'Start (12)' and 'Stop (31)' labels to the corresponding positions in the reference sequence. A black arrow points from the 'Read 1' label to the start of its sequence.

Short read sequencing data is compared to the reference (looking for a "match")

We first need such alignment before we can analyse variation

Read variation is analysed within this alignment context



>Chr01
ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCAT**G**ATACATGGCTAGT...
Read 1 AGCATGATCAT**T**CATGATGA
Read 2 GCATGATGATCATGCAT**A**ATACATGG
Read 3 TGATGATCATGCAT**A**ATACATGGCTAGT

Start

SNP (G/T, 23)

Stop

SNP (G/A, 40)

An accurate alignment is *essential* before we can trust any variant

Read variation is analysed within this alignment context

The diagram illustrates sequence alignment with two SNPs. A reference sequence is shown with two SNPs highlighted in green: 'G/T' at position 23 and 'G/A' at position 40. Red arrows indicate the 'Start' and 'Stop' of a read. Four reads are aligned below the reference sequence, with their variations highlighted in green or red.

>Chr01

Start SNP (G/T, 23) Stop SNP (G/A, 40)

ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCAT**G**ATACATGGCTAGT...

Read 1 AGCATGATCAT**T**CATGATGA

Read 2 GCATGATGATCATGCAT**A**ATACATGG

Read 3 TGATGATCATGCAT**A**ATACATGGCTAGT

Read 4 **TGATGATCATGCATAATACATGGCTAGT**

An accurate alignment is *essential* before we can trust any variant

Read variation is analysed within this alignment context

>Chr01
ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCAT**G**ATACATGGCTAGT...
Read 1 AGCATGATCAT**T**CATGATGA
Read 2 GCATGATGATCATGCAT**A**ATACATGG
Read 3 TGATGATCATGCAT**A**ATACATGGCTAGT

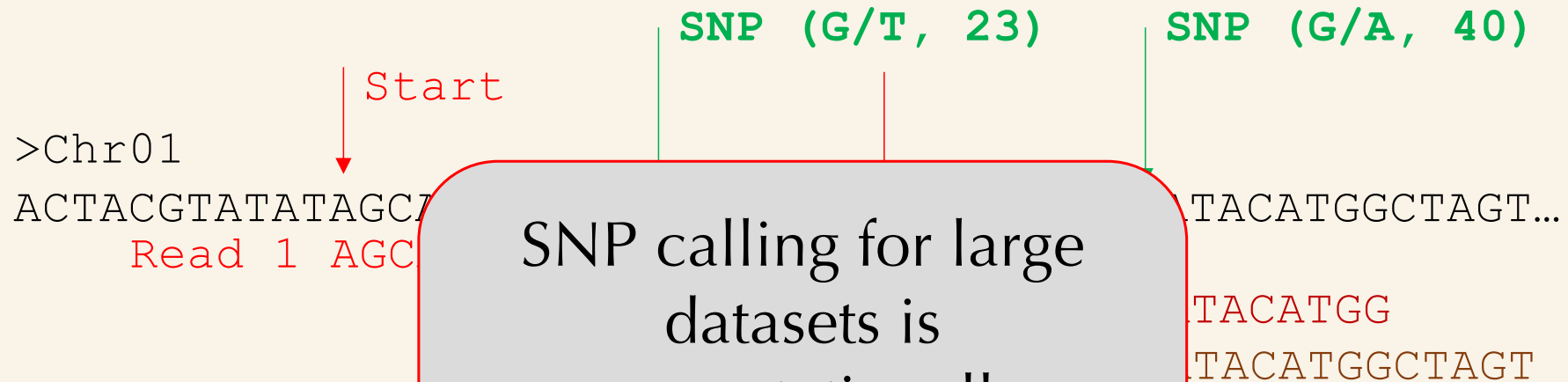
Start

SNP (G/T, 23)

SNP (G/A, 40)

We usually analyse *millions to billions* of reads and compare these to reference genomes that consist of **billions** of nucleotides/bases (Human genome ~3Gb)

Read variation is analysed within this alignment context



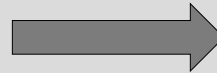
We usually compare the alignment of reads and that consist of **billions** of nucleotides/bases (Human genome ~3Gb)

Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!

Reference

Unaligned reads



Alignment program

BWA

BowTie

Reference

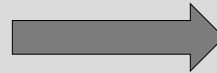
Aligned reads (nicely sorted
and tiled)

Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!

Reference

Unaligned reads



Alignment program

BWA

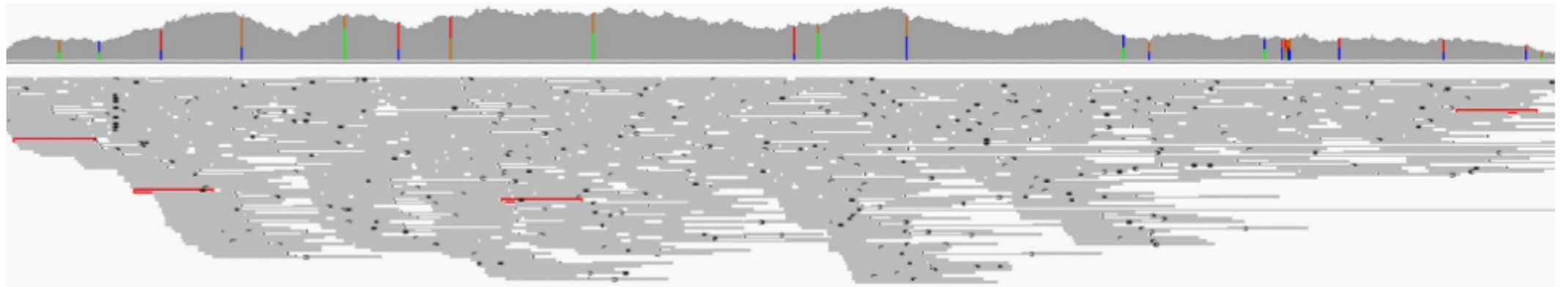
BowTie

Reference

Aligned reads (nicely sorted
and tiled)

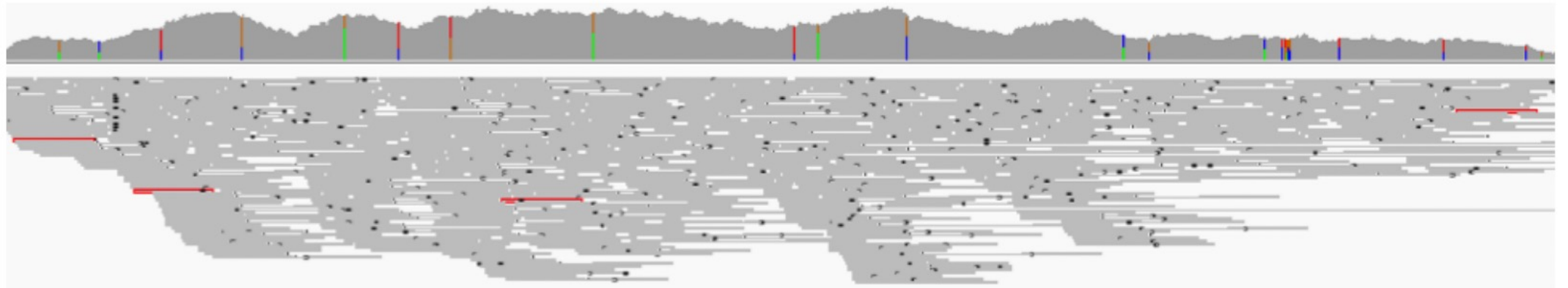
Standard program settings are usually sufficient

Visualisation of thousands of reads



Visualisation of thousands of reads

Genetic variation (SNPs) colours reflect which bases are variable (A-C, A-T, G-C, etc)



After aligning, we need another program to determine which bases are variable:

A SNP caller

SNP calling programs

Table 1. A brief summary of different tools.

caller	Bcftools	16GT	Freebayes	VarScan2	GATK
Code	C	Perl	C++	Java	Java
Model	HMM & MAQ	16-genotype probabilistic	Bayesian	heuristic algorithm	Bayesian
Sampling	Single & multiple	Single	Single	Single & multiple	Single & multiple
Variants	SNPs & indels	SNPs & indels	SNPs & indels&MNPs	SNPs & indels	SNPs & indels
Features	Sorting, indexing, etc.	easy to use, timesaving	straightforward	meet desired thresholds for read depth, base quality, variant allele frequency, and statistical significance	Realignment, per base recalibration, VQSR
Reference	Danecek et al., 2017 [15]	Luo et al., 2017 [19]	Garrison and Marth, 2012 [18]	Koboldt et al., 2012 [16]	Mckenna et al., 2010 [14]

<https://doi.org/10.1371/journal.pone.0262574.t001>

Liu J, Shen Q, Bao H (2022)

Many programs exist, and there is *continuous* development
For instance BCFtools/16GT are now recommended
Yet use of GATK is wide-spread (oldest, developed by Broad institute, good documentation)

What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

>Chr01

ACTACGTATATAGCATGATCAT**G**CATGATGATCATGCATGATACATGGCTAGT...

Read 1 AGCATGATCAT**T**CATGATGA

Is this real or not?

Discuss between each other (few mins) and come with suggestions how to know this is real or not.

What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

```
>Chr01
```

```
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
```

```
Read 1 AGCATGATCATTCATGATGA
```

Is this real or not?

Sequencing data (as any type of data) comes with errors (wrong bases called) and/or uncertainty (low quality of bases) in the call

Solution? Generate LOTS more data!

What does a variant caller do?

With more data (read), more certainty is obtained: ***fold coverage***

```
>Chr01
```

```
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
```

```
Read 1 AGCATGATCATTCATGATGA
```

```
Read 2 ATGATCATTCATGATGATCAT
```

```
Read 3 GATCATTCATGATGATCATGCATGAT
```

```
Read 4 TCATTCATGATGATCATGCAT
```

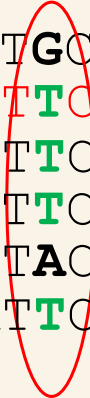
```
Read 5 CATTCATGATGATCATGCATGATACATGG
```

5-fold coverage, all the same, we are pretty certain about this call (note: we usually strive for ~20 fold coverage)

What does a variant caller do?

Another example

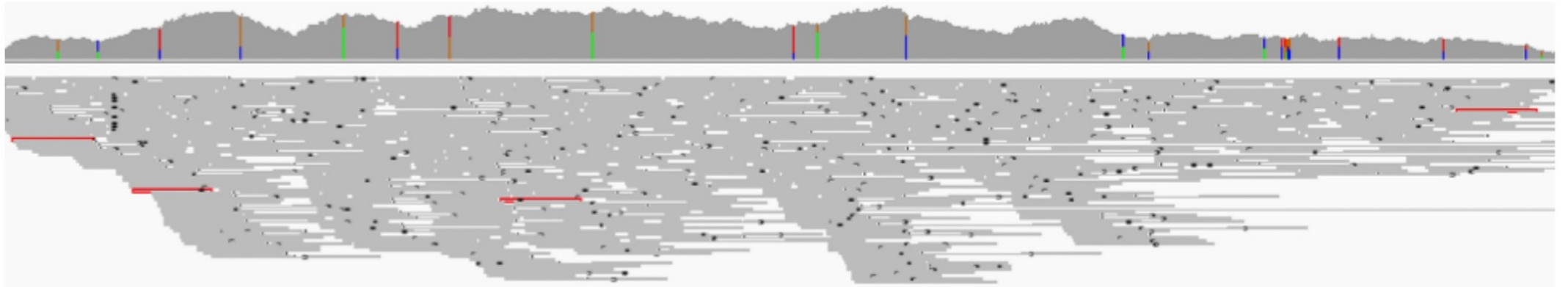
```
>Chr01
ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATACATGGCTAGT...
Read 1 AGCATGATCATTCATGATGA
Read 2 ATGATCATTCATGATGATCAT
Read 3 GATCATTCATGATGATCATGCATGAT
Read 4 TCATACATGATGATCATGCAT
Read 5 CATTCATGATGATCATGCATGATACATGG
```



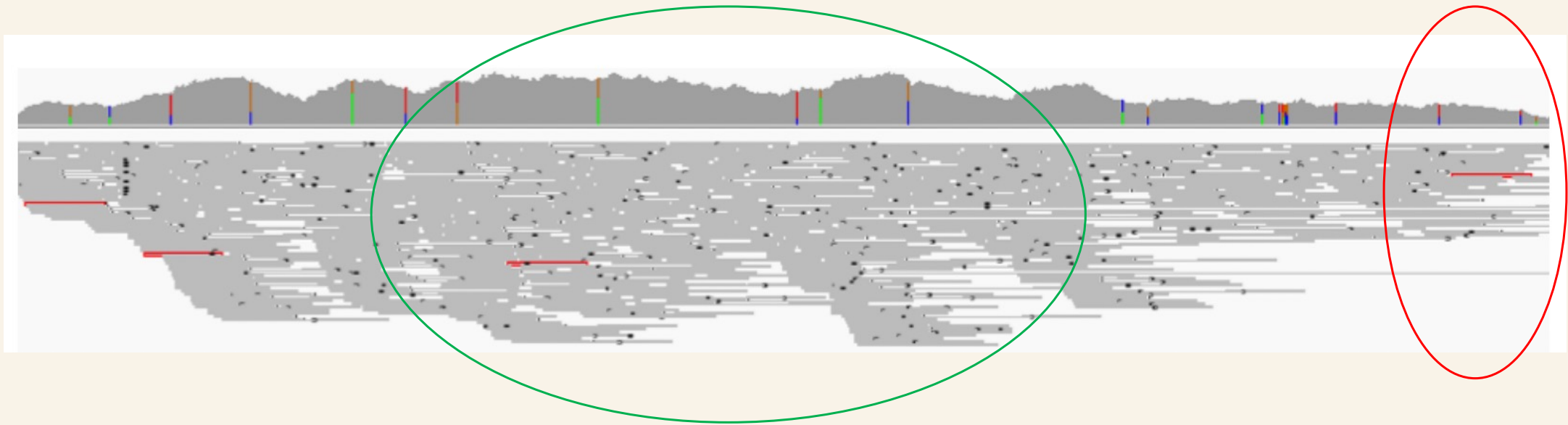
We cannot be so certain about the A, until we get more data

Coverage is the most important determinant for the quality of your data

Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity



Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity

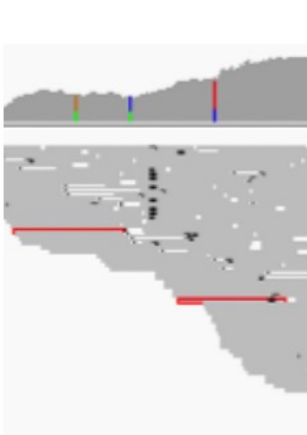


Higher coverage

Lower coverage

Yet along a reference, you'll obtain variable coverage due to random complexity

SNP callers run complex statistical models (e.g. Bayesian or HMM models) to provide confidence in SNP calls and if they are "TRUE". They often assume correct read alignment **and** require sufficient read coverage in order to provide high-quality calls



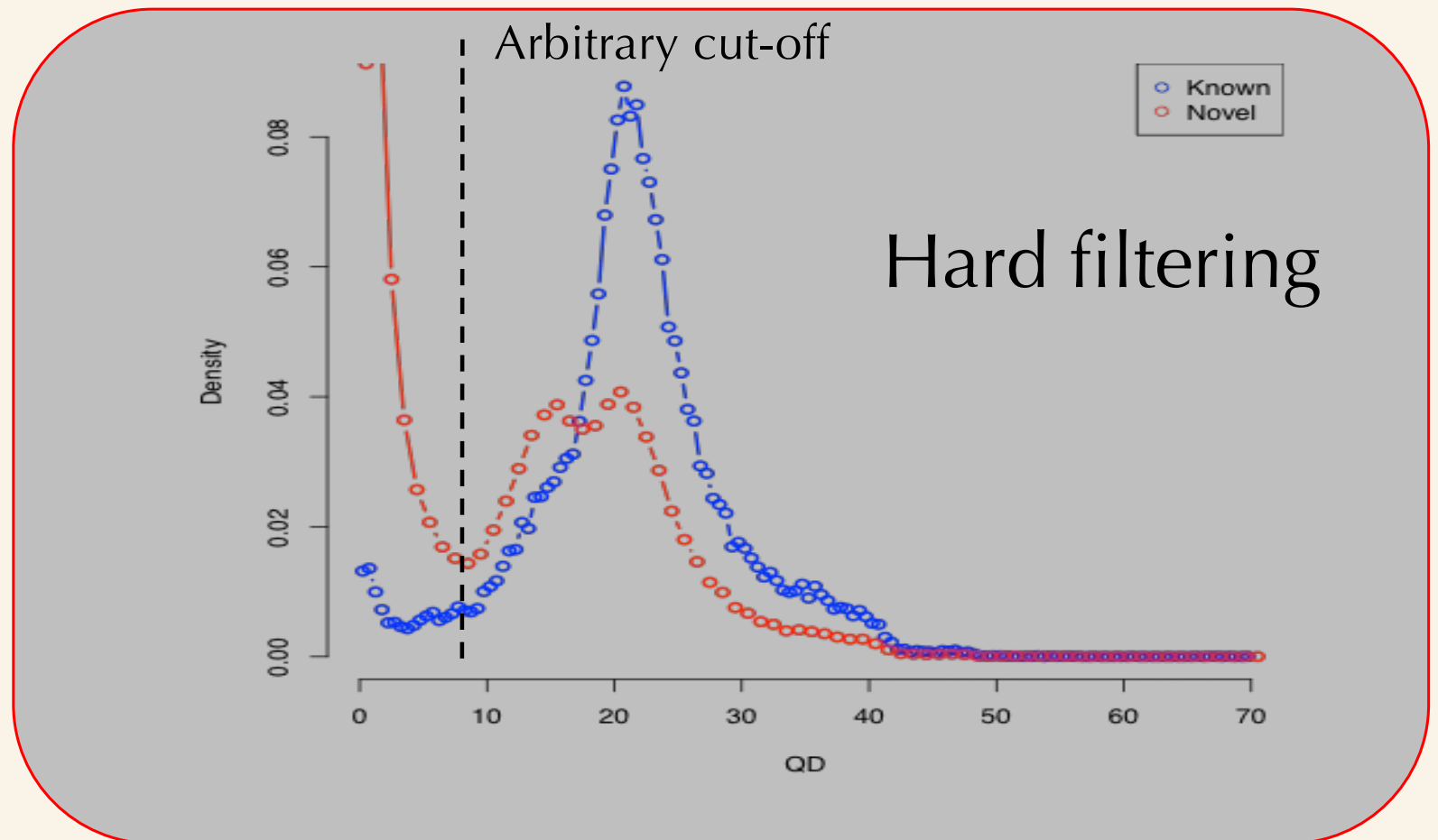
Higher coverage



Lower coverage

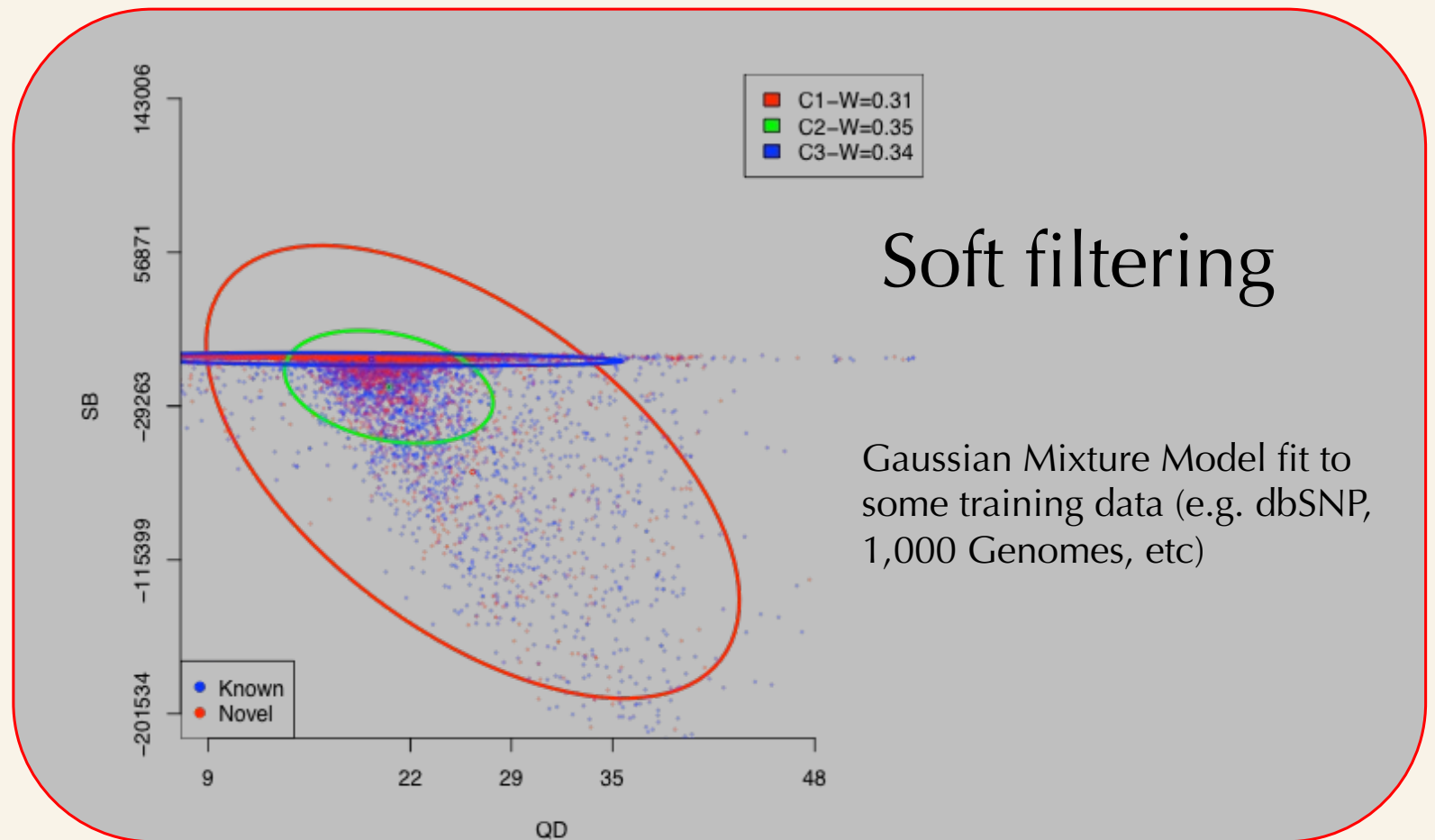
SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

We need to ***filter*** our data to only retain the high quality part of the data

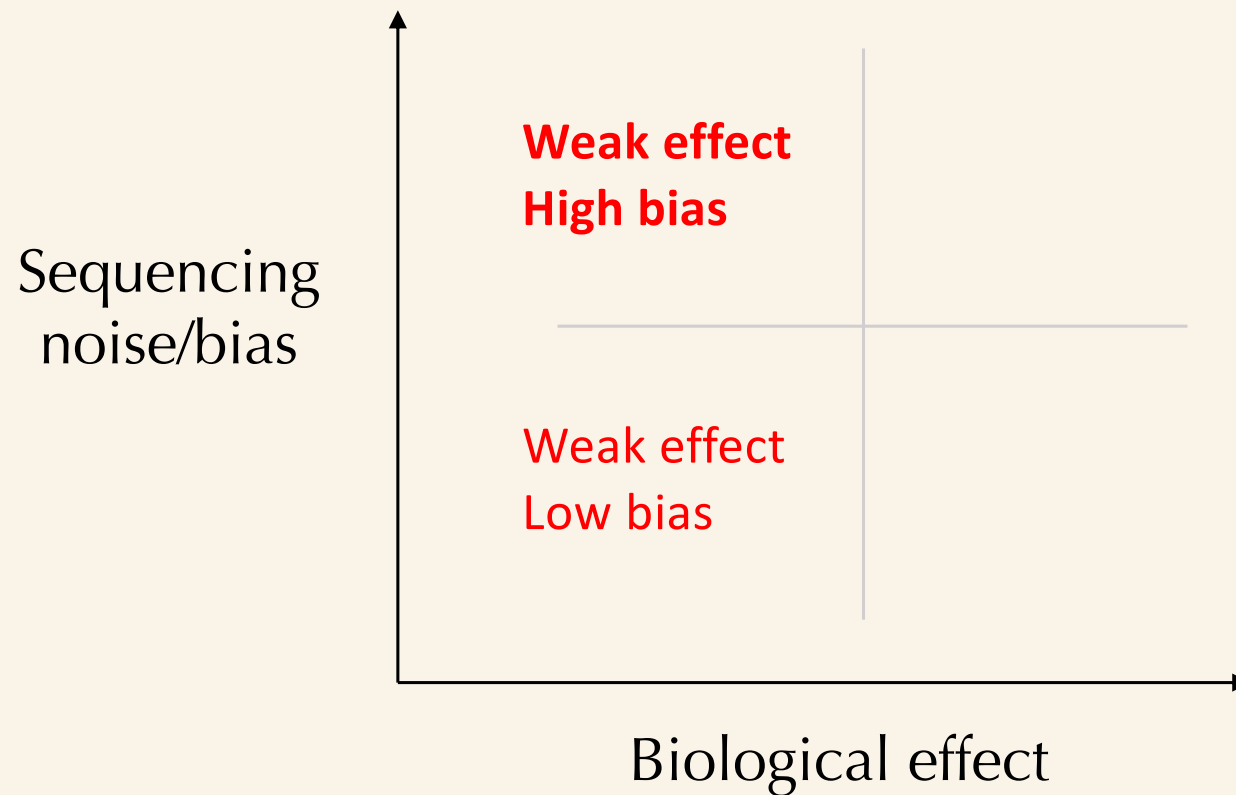


SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

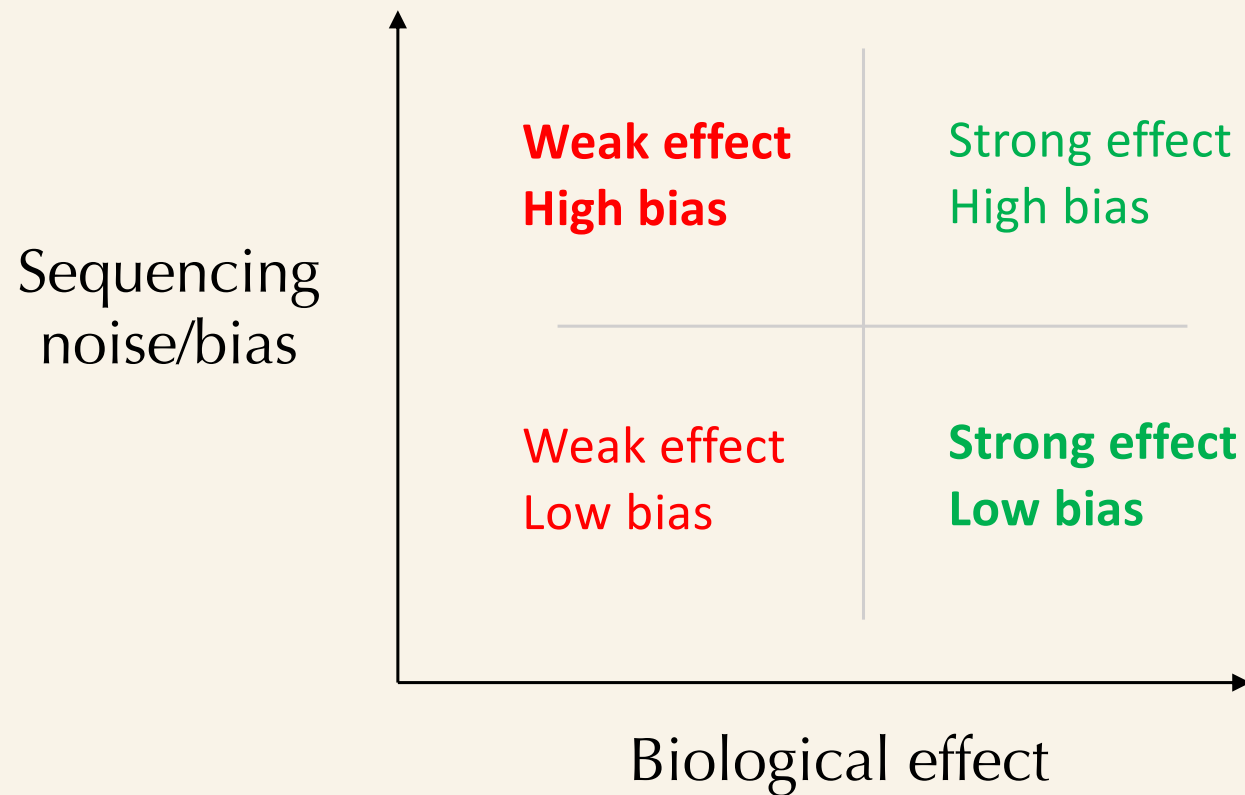
We need to ***filter*** our data to only retain the high quality part of the data



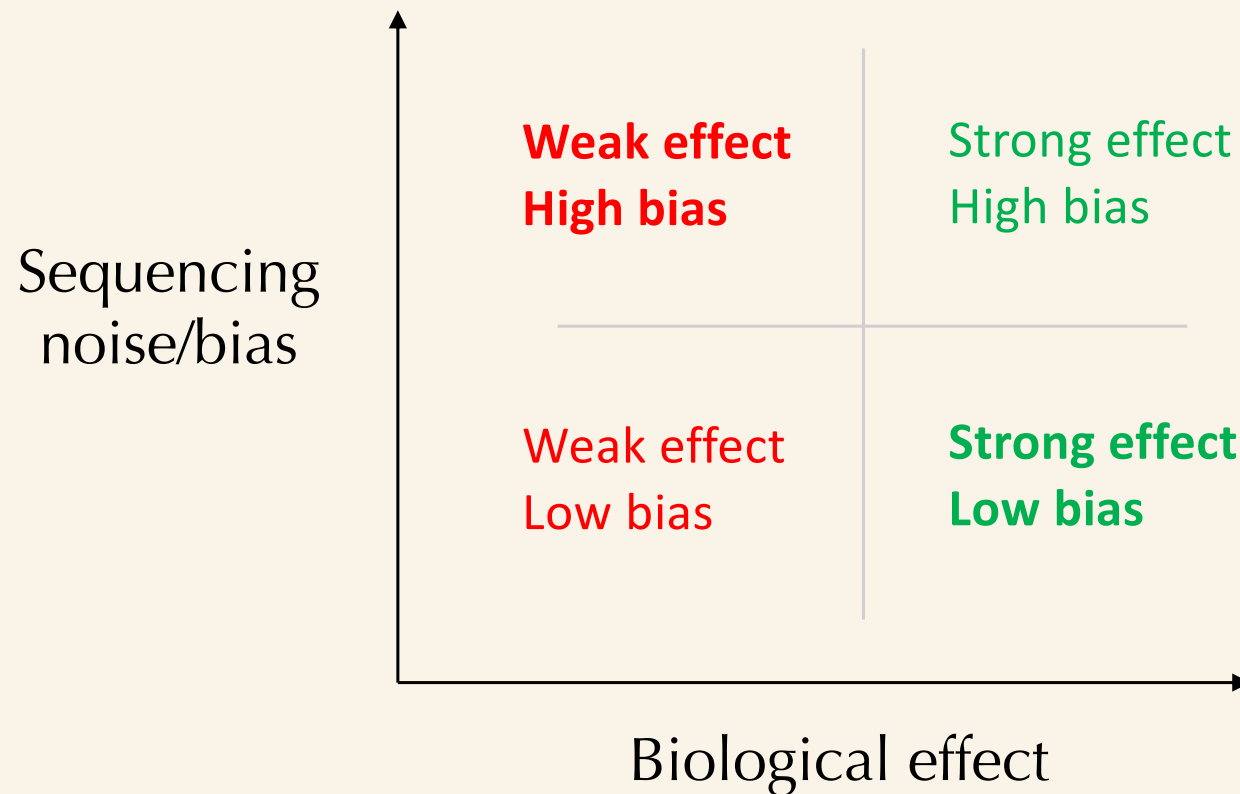
Yet there is no “fixed” approach to filtering your data



Yet there is no “fixed” approach to filtering your data



Yet there is no “fixed” approach to filtering your data



It is not always clear from the outset where you are! You need to explore your data and use preliminary analyses

Questions?



After all this, what does a variant calling pipeline look like?



Reference

Reads



e.g.
population
data

After all this, what does a variant calling pipeline look like?



Reference

Mapping/aligning

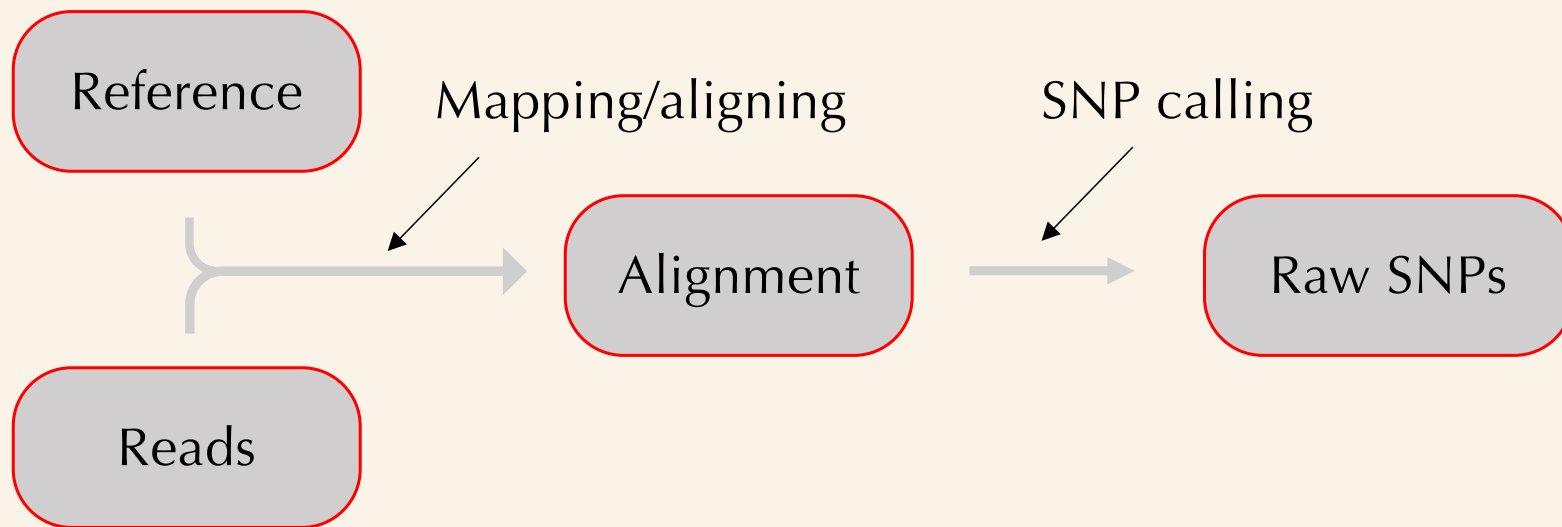
Alignment

Reads



e.g.
population
data

After all this, what does a variant calling pipeline look like?



e.g.
population
data

After all this, what does a variant calling pipeline look like?



Reference

Mapping/aligning

SNP calling

Filtering and preliminary analyses

Alignment

Filtered
SNPs

Reads

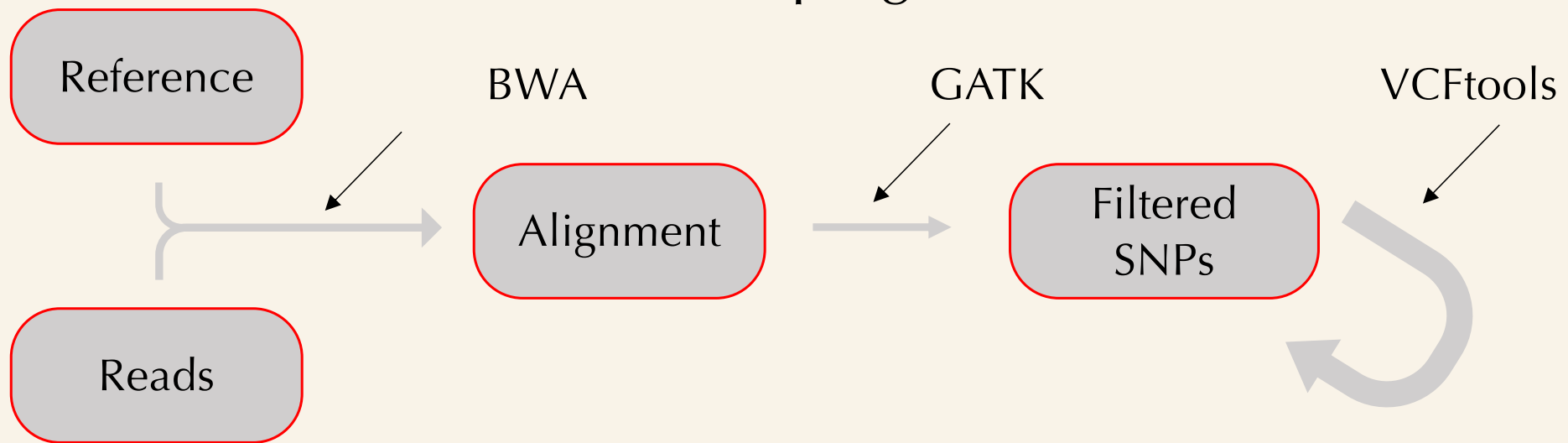


e.g.
population
data

After all this, what does a variant calling pipeline look like?

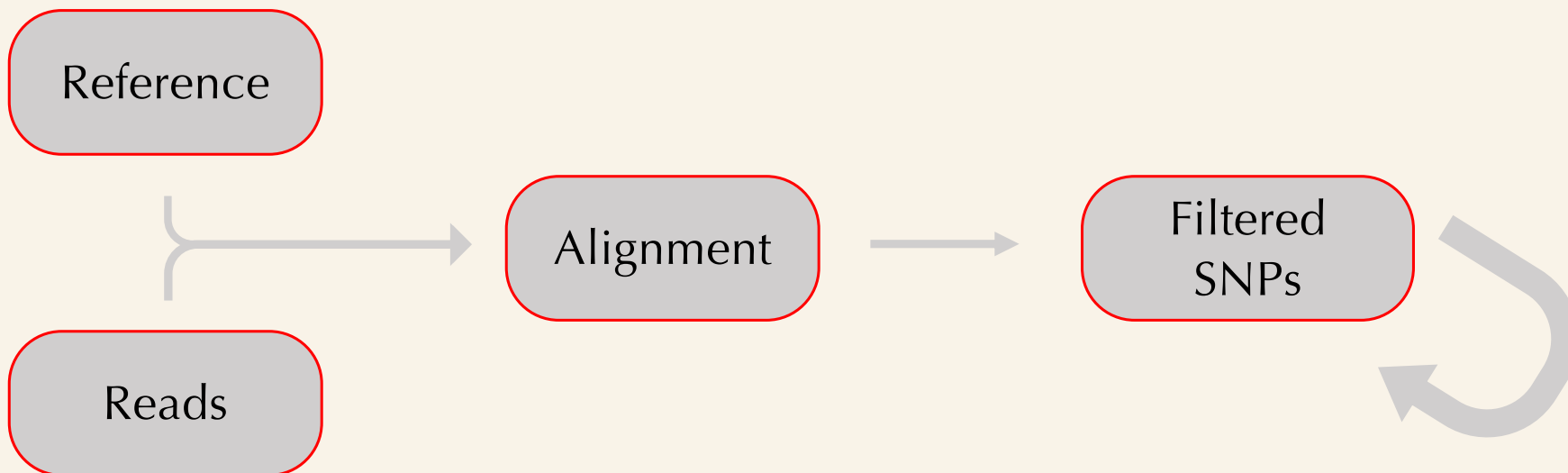


A selection of programs that can be used

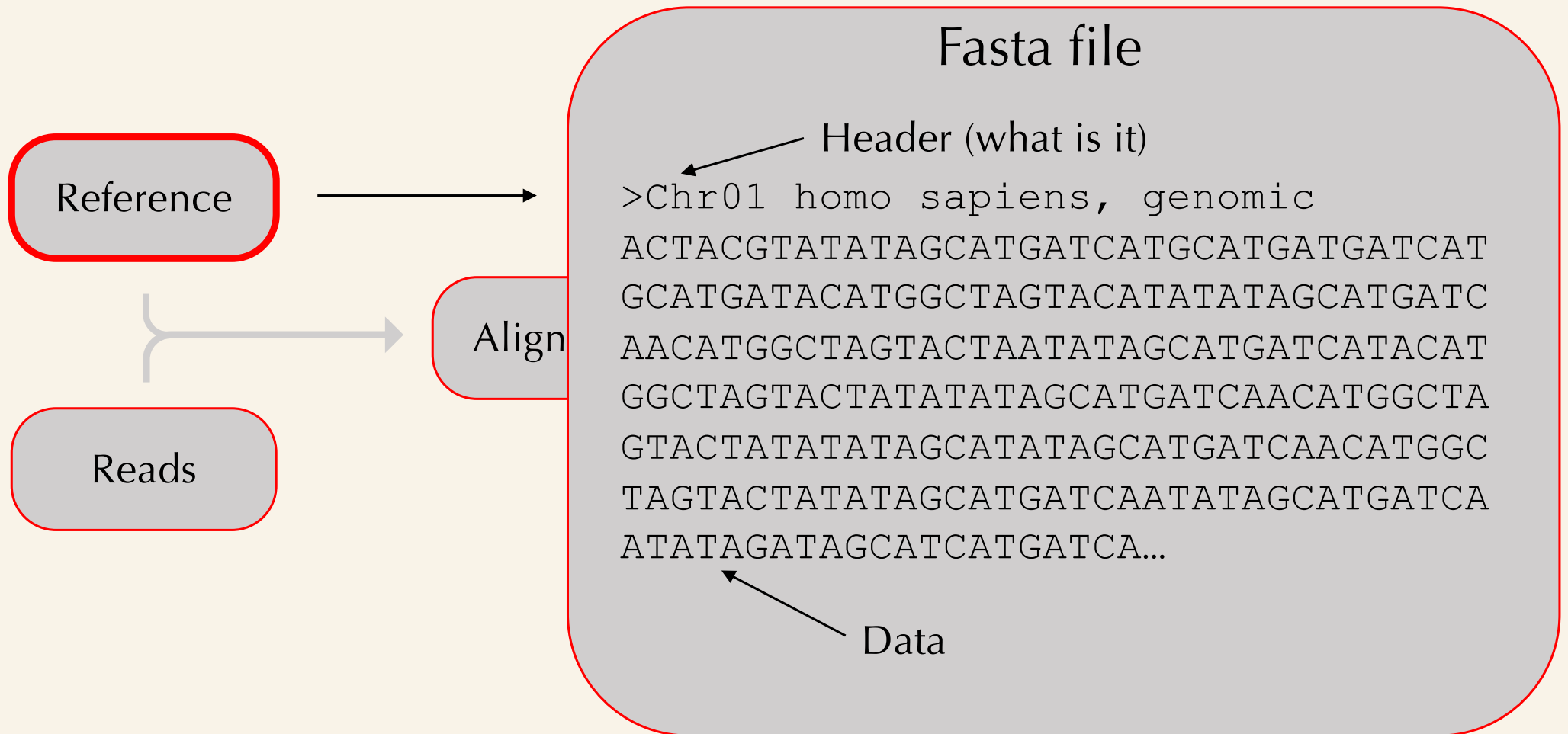


e.g.
population
data

Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!

Alignment

BAM file (binary alignment file)



Kind of data

“Header” with information about the file

```
@HD VN:1.5 GO:none SO:coordinate
@SQ SN:NC_004029.2 LN:16565
@RG ID:L1i1_AGAACCG SM:WLR001 LB:L1i1_AGAACCG PU:Nistelberger-DNA1-2016-04-15
@RG PL:ILLUMINA PG:bwa
@PG ID:bwa PN:bwa VN:0.7.17-r1188 CL:bwa samse Orosvlmt.fasta
@PG ID:GATK IndelRealigner VN:3.6-0-g89b7209 CL:knownAlleles=[] targetIntervals=WLR001/Wal_m
@PG ID:samtools CL:samtools view -H WLR001.Wal_mt.realigned.bam
```

Reference

Sample name

Each of these steps requires specific files to work with!

Alignment

BAM file (binary alignment file)

Readname

Follow by data with information about each read alignment

Start of alignment

Matching bases

M_D00564:55:C9FG3ANXX:7	0	NC_004029.2	419	37	91M	TAAAAAGCTGCCGCTAATACAAAATATACTACGAAAGTGACT
M_D00564:55:C9FG3ANXX:7	0	NC_004029.2	474	37	58M	TTACACGACAGCTAAGACCCAAACTGGGATTAGATACCCCAC
M_D00564:55:C9FG3ANXX:7	0	NC_004029.2	515	37	56M	CTATGCTTAGCCATAAACACAAATAATTTGCACAACAAAATT

Reference name

Quality of alignment (37 is max)

CIGAR string (56 matching bases)

Each of these steps requires specific files to work with!

SNP data



VCF file (Variant call format)

Again, a “Header” with lots of information about the file

```
##fileformat=VCFv4.2
##ALT=<ID=NON_REF,Description="Represents any possible alternative allele not already represented at this location">
##FILTER=<ID=LowQual,Description="Low quality">
##FILTER=<ID=PASS,Description="All filters passed">
##FORMAT=<ID=AD,Number=R,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth (reads with MQ=255 or with bad mates are
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=MIN_DP,Number=1,Type=Integer,Description="Minimum DP observed within the GVCF block">
##FORMAT=<ID=PL,Number=G,Type=Integer,Description="Normalized, Phred-scaled likelihoods for genotypes as defined in
##FORMAT=<ID=RGQ,Number=1,Type=Integer,Description="Unconditional reference genotype confidence, encoded as a phred
##FORMAT=<ID=SB,Number=4,Type=Integer,Description="Per-sample component statistics which comprise the Fisher's Exac
##GATKCommandLine=<ID=GenomicsDBImport,CommandLine="GenomicsDBImport --genomicsdb-workspace-path Walrus_DB --variar
##GATKCommandLine=<ID=GenotypeGVCFs,CommandLine="GenotypeGVCFs --output Walrus_MT.vcf.gz --variant gendb://Walrus_I
##GATKCommandLine=<ID=HaplotypeCaller,CommandLine="HaplotypeCaller --sample-ploidy 1 --emit-ref-confidence GVCF --c
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##INFO=<ID=BaseQRankSum,Number=1,Type=Float,Description="Z-score from Wilcoxon rank sum test of Alt Vs. Ref base qu
```

Each of these steps requires specific files to work with!

SNP data



VCF file (Variant call format)

Followed by the data:

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO
NC_004029.2	131	.	T	C	356.22	.	AC=1;AF=0.022;AN=45;DP=143;FS=0.000;MLEAC=1;MLEAF=C
NC_004029.2	162	.	T	C	18479.23	.	AC=15;AF=0.333;AN=45;BaseQRankSum=0.00;DP=543;FS=0.
NC_004029.2	198	.	C	T	608.22	.	AC=1;AF=0.022;AN=45;DP=410;FS=0.000;MLEAC=1;MLEAF=C
NC_004029.2	387	.	G	A	547.22	.	AC=1;AF=0.022;AN=45;DP=408;FS=0.000;MLEAC=1;MLEAF=C
NC_004029.2	616	.	T	C	235.62	.	AC=1;AF=0.022;AN=45;DP=406;FS=0.000;MLEAC=1;MLEAF=C
NC_004029.2	741	.	C	T	819.22	.	AC=1;AF=0.022;AN=45;DP=412;FS=0.000;MLEAC=1;MLEAF=C
NC_004029.2	743	.	C	T	819	.	AC=1;AF=0.022;AN=45;DP=413;FS=0.000;MLEAC=1;MLEAF=C



Reference name

Each of these steps requires specific files to work with!

SNP data

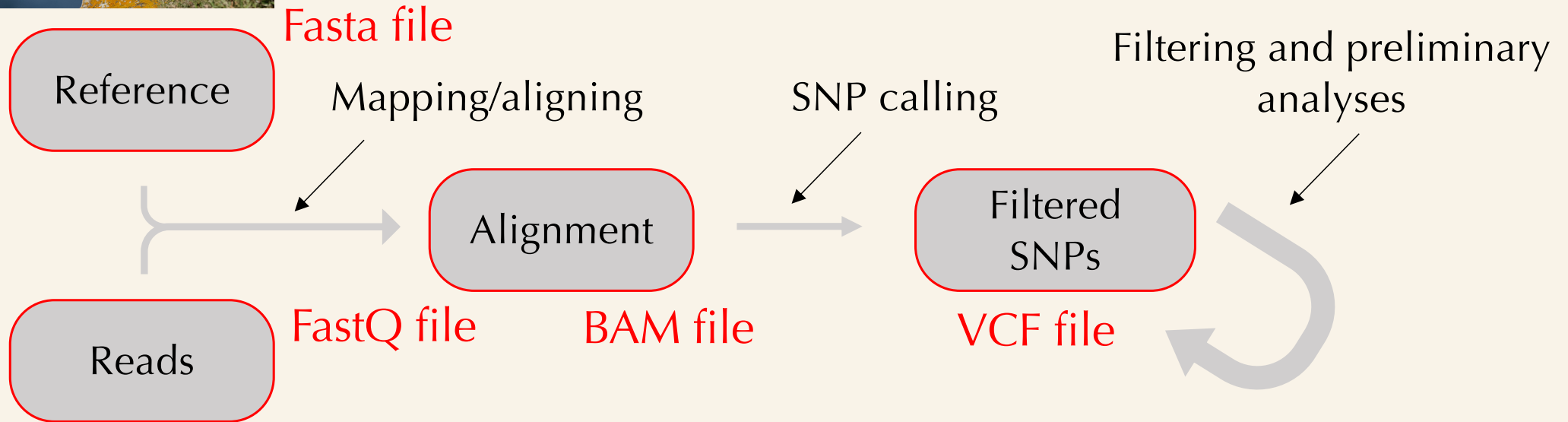
VCF file (Variant call format)

Followed by the data:

GenoType: Allele Depth: Read DePth (DP): Genotype Quality: Phred-scaled Likelihood

FORMAT	WLR001	WLR002	WLR003	WLR004	WLR005
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:2,0:2:90:0,90	0:0,0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:4,0:4:99:0,135	0:1,0:1:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:5,0:5:46:0,46	0:0,0:0:0:0:0,0	0:2,0:2:90:0,90	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0:0,0	0:2,0:2:45:0,45	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0:0,0	0:1,0:1:42:0,42	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:1,0:1:42:0,42	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:3,0:3:99:0,119	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:3,0:3:99:0,119	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:0:0,0	0:1,0:1:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:0:0,0	0:1,0:1:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0	0:0,0:0:0:0:0,0

After all this, what does a variant calling pipeline look like?



e.g.
population
data

Questions?



Then it is *Quiz*
time!



Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and pitfalls
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

